



## Potentially toxic element contamination in soil and accumulation in maize plants in a smelter area in Kosovo

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# Environmental Science and Pollution Research

## Potentially toxic element contamination in soil and accumulation in maize plants in a smelter area in Kosovo

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<b>Abstract:</b>	A biogeochemical field study was carried out in the industrial area of Kosovska Mitrovica in northern Kosovo, where agricultural soils were contaminated by potentially toxic elements due to smelting activity. Total and bioavailable contents of As, Cd, Co, Cu, Pb, Sb, U and Zn in soil and their concentrations in maize roots and grains were determined. Soil contamination by As, Cd, Cu, Pb, Sb and Zn was variable from slightly to highly contaminated soils, and influenced both the bioavailable fraction and accumulation of these potentially toxic elements in maize tissues. The comparison between potentially toxic element concentrations in roots and grains indicated that maize is able to limit the transfer of non-essential elements to edible parts. The plant-to-soil bioconcentration indices suggested that the transfer of potentially toxic elements from soil to plant was predicted better by bioavailable concentrations than by the total contents. These indices further identified some competitions and interactions among these elements in root uptake and root-to-grain translocation.
<b>Response to Reviewers:</b>	Ref.: Ms. No. ESPR-D-15-04475 Potentially toxic element contamination in soil and accumulation in maize plants in a smelter area in Kosovo  Reviewers' comments  Reviewer #1:  The manuscript Number: ESPR-D-15-04475 "Heavy element contamination in soil and accumulation in maize plants in a smelter area in Kosovo" submitted for Research Article in Environmental Science and Pollution Research presents an interesting field study on soil-plant transfers of various inorganic elements potentially toxic. Actually, the question of produce (human) safe vegetables in polluted areas is increasingly asked at the global scale. The biogeochemical field studies are needed to better understand the transfers of metal(loids) in agricultural soils. However, several changes are needed before the manuscript could be accepted.

[Click here to view linked References](#)

1 **Potentially toxic element contamination in soil and accumulation in**  
2 **maize plants in a smelter area in Kosovo**

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36 **ABSTRACT**

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38 A biogeochemical field study was carried out in the industrial area of Kosovska Mitrovica  
39 in northern Kosovo, where agricultural soils were contaminated by potentially toxic  
40 elements due to smelting activity. Total and bioavailable contents of As, Cd, Co, Cu, Pb,  
41 Sb, U and Zn in soil and their concentrations in maize roots and grains were determined.  
42 Soil contamination by As, Cd, Cu, Pb, Sb and Zn was variable from slightly to highly  
43 contaminated soils, and influenced both the bioavailable fraction and accumulation of these  
44 potentially toxic elements in maize tissues. The comparison between potentially toxic  
45 element concentrations in roots and grains indicated that maize is able to limit the transfer  
46 of non-essential elements to edible parts. The plant-to-soil bioconcentration indices  
47 suggested that the transfer of potentially toxic elements from soil to plant was predicted  
48 better by bioavailable concentrations than by the total contents. These indices further  
49 identified some competitions and interactions among these elements in root uptake and  
50 root-to-grain translocation.

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53 *Keywords:* Potentially toxic elements; Soil; *Zea mays* L.; Smelter; Bioavailability; Kosovo.

54

## 55 1. Introduction

56 Smelting activity is an important source of potentially toxic elements (PTEs) and cause of  
57 adverse environmental impacts. The emission of PTEs from smelters in form of particulate  
58 matter and gases frequently produced soil contamination (Adriano 2001; Cappuyens et al.  
59 2006; Li et al. 2006; Navarro et al. 2008; Razo et al. 2004). The PTEs released into the  
60 atmosphere by smelting activities may accumulate in topsoil and be transferred to plants  
61 through root uptake, involving a contamination risk of the food chain.

62 Contamination of agricultural soils around smelting plants is a major environmental  
63 concern as it can cause accumulation of PTEs in crops affecting plant productivity, food  
64 quality and human health (Dudka and Miller 1999; Gupta and Gupta 1998; Kachenko and  
65 Singh 2006; Lim et al. 2008; Pruvot et al. 2006; Roy and McDonald 2013; Weisło et al.  
66 2002; Yang et al., 2011; Zheng et al. 2007).

67 Maize (*Zea mays* L.) is one of most widely and intensively cultivated cereal worldwide,  
68 and it constitutes a staple food for humans and animals. For the global production,  
69 utilization and consumption of maize, the transfer of PTEs through the soil-maize system  
70 may represent an important pathway of human exposure, especially in contaminated areas  
71 (Rosas-Castor et al. 2014). Contamination of agricultural soils by PTEs may result in an  
72 increase of their uptake by maize with negative effects on the quality of crops and a  
73 potential risk for human health. For these reasons, several studies were carried out to  
74 determine levels and distribution of PTEs in maize plants grown in soils contaminated by  
75 industrial, smelting and mining activities (Awokunmi et al. 2015; Bi et al. 2009; Bini et al.  
76 2013; Liu et al. 2005; Marwa et al. 2012; Wahsha et al. 2014) or cultivated using different  
77 tillage systems (Brennan et al. 2014; Carbonell et al. 2011; Garcia-Marco et al. 2014;  
78 Lavado et al. 2007; Lu et al. 2015; Muchuweti et al. 2006). These experimental and field  
79 studies aimed to assess the impact of soil contamination and agricultural practices on  
80 uptake, translocation and accumulation of PTEs in maize tissues (roots, leaves and grains).

81 The Kosovska Mitrovica area in northern Kosovo represents a zone of interest for studies  
82 concerning the contamination of agricultural soils by PTEs caused by smelter emissions,  
83 and their transfer and accumulation in cultivated plants. This area was one of the main  
84 industrial sites in the former Yugoslavia and an important mining district in Europe.  
85 Industrial activity focused on the Zvecan Pb-Zn smelter and the Trepča factory of batteries  
86 and accumulators, caused an intense and widespread soil contamination by PTEs such as  
87 As, Cd, Cu, Pb, Sb and Zn (Borgna et al. 2009; Šajn et al. 2013). Soil contamination

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88 involved a wide agricultural area extending in the Ibar and Sitnica alluvial plains where  
89 maize is the most cultivated crop.

90 A field biogeochemical research was carried out in the K. Mitrovica area in order to define  
91 the influence of soil contamination due to smelting activity on uptake, translocation and  
92 accumulation of potentially toxic elements in maize tissues. The total contents and the  
93 bioavailable concentrations of As, Cd, Co, Cu, Pb, Sb, U and Zn in soil samples as well as  
94 their concentrations in maize roots and grains were determined.

95 This research is a contribution to the knowledge about the soil-to-plant transfer of PTEs in  
96 smelter-contaminated areas. The main objectives of the study were: i) to establish a  
97 relationship between soil contamination and bioavailability of PTEs in soil; ii) to evaluate  
98 the influence of soil contamination on PTE transfer from soil to maize; iii) to define the  
99 behaviour of PTEs in uptake, translocation and accumulation in maize roots and grains  
100 grown in soils affected by variable contamination levels.

101 Our research considered both the total content and bioavailable fraction of PTEs in soil as  
102 chemical pools that may rule the soil-to-maize transfer of these elements, and measured the  
103 levels and distribution of Sb and U in the soil-maize system. To the best of our knowledge,  
104 few field studies used the bioavailable concentrations of PTEs in soil for these purposes  
105 (Garcia-Marco et al. 2014; Lavado et al. 2007), and focused on uptake, translocation and  
106 accumulation of Sb and U in maize tissues.

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## 109 **2. Materials and methods**

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### 111 **2.1. Study area**

112 The study area is located in northern Kosovo and includes the Trepča mining and industrial  
113 complex centred on the town of Kosovka Mitrovica. The mines of the Trepča complex  
114 exploited skarn-type ore bodies mainly consisting of base-metal (Pb-Zn) massive  
115 sulphides. Industrial activity was focused on the Zvecan Pb-Zn smelter and the Trepča  
116 factory (Fig. 1).

117 The Zvecan smelter worked from 1939 to 2000 to extract metals such as Ag, Au, Bi, Pb  
118 and Zn from the crude ore of Trepča mines. Smelting processes released into the  
119 atmosphere huge quantities of contaminants (mainly SO<sub>x</sub> and potentially toxic elements)  
120 as gas and particulate matter, and produced waste disposed in large open dumps close to

121 the smelter. The Trepča factory produced battery and accumulators from 1967 to 1999. An  
122 open dump of industrial waste is present in the factory site.

123 In the study area soil was intensively and extensively contaminated by potentially toxic  
124 elements (As, Cd, Cu, Pb, Sb and Zn) due to the emissions from the Zvecan smelter and  
125 Trepča factory and, to a lesser degree, the airborne dispersion of mining and industrial  
126 waste (Borgna et al. 2009; Nannoni et al. 2011; Šajin et al. 2013). Contamination affected  
127 the upper part of soil profile (usually up to 50-60 cm deep) and extended up to 22 km north  
128 and south of the smelting plant (Borgna et al. 2009). The concentrations of As, Cd, Cu, Pb,  
129 Sb and Zn in soil progressively decreased with distance from the Zvecan smelter, and were  
130 influenced by the dominant wind directions (mainly from NNW). Soil contamination  
131 involved a wide agricultural zone extending mainly in the Sitnica alluvial plain south of the  
132 K. Mitrovica town (Fig. 1).

133 The landscape of the study area is characterized by high hills usually from 600 to 1000 m  
134 a.s.l. These reliefs are arranged in two NW-SE ridges surrounding the valleys of the Ibar  
135 and Sitnica Rivers. The Sitnica River originates about 40 km south-east of K. Mitrovica  
136 and flows north-east in a wide asymmetrical valley. Near the town of K. Mitrovica the  
137 Sitnica River merges into the Ibar River that flows northward in a narrow valley.

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## 139 **2.2. Sampling strategy**

140 Topsoil samples (20 cm deep) and maize plants were collected in 24 cultivated sites  
141 located in the alluvial plains of the Ibar and Sitnica Rivers at variable distances from the  
142 Zvecan smelter (from 2.2 to 20.7 km; Fig. 1).

143 Each topsoil sample was a composite sample consisting of three sub-samples collected a  
144 few metres apart in three subsites. Soil sub-samples were collected using a stainless steel  
145 spade. This tool was utilized to extract a 20x20x20 cm soil block containing the roots of a  
146 maize plant. By using this sampling procedure, in each sampling site three plants of maize  
147 were collected just before the harvest, and mixed to form a composite sample.

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## 149 **2.3. Laboratory treatment**

150 Soil samples were dried at +40 °C and manually sieved through a 2 mm mesh. Soil fraction  
151 <2 mm was homogenized by quartering and pulverization. Soil samples were solubilized  
152 adding 1 mL HF, 2 mL HNO<sub>3</sub>, 2 mL HCl and 1 mL HClO<sub>4</sub> to about 200 mg of powdered  
153 soil. To determine the total content of As, chlorine-free chemical reagents were used and  
154 soil digestion was carried out adding 2 mL HF, 2 mL HNO<sub>3</sub>, and 1 mL H<sub>2</sub>O<sub>2</sub> to about 200

155 mg of powdered soil. Both acid digestion procedures were performed in Teflon® bombs  
156 using a Milestone Ethos 900 microwave lab station. Ultrapure trace-grade reagents were  
157 used for soil laboratory treatment.

158 The maize plants were stored at +4 °C and roots and grains separated. Roots were  
159 preliminarily washed with running tap water and then immersed in an ultrasonic bath for  
160 30 min and rinsed with deionized water. Grains were washed with running tap water and  
161 rinsed with deionized water. Both roots and grains were dried at +30 °C and then finely  
162 powdered before elemental analysis. The roots and grains were solubilized by acid  
163 digestion adding in Teflon® bombs 6 mL HNO<sub>3</sub> and 1 mL H<sub>2</sub>O<sub>2</sub> (ultrapure reagents) to  
164 about 500 mg of powdered sample. Acid digestion was performed using a Milestone Ethos  
165 900 microwave lab station.

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#### 167 ***2.4. Analytical determinations***

168 The hydrometer method was used to determine the size distribution of soil particles  
169 expressed as percent content of the sandy, silty and clayey fractions. Soil pH (pH<sub>H<sub>2</sub>O</sub>) and  
170 the effective cation exchange capacity (CECe) were measured following the EPA method  
171 9045/D and the procedure of Hendershot and Duquette (1986), respectively. The method of  
172 Walkley and Black (1934) was employed to determine the organic carbon content, and De  
173 Astis calcimeter to measure the carbonate content (as % CaCO<sub>3</sub>).

174 Inductively coupled plasma-mass spectrometry (Perkin Elmer Sciex Elan 6100  
175 spectrometer) was employed to measure the total contents and bioavailable concentrations  
176 of As, Cd, Co, Cu, Pb, Sb, U and Zn in soil samples as well as the concentrations in maize  
177 roots and grains.

178 The analytical accuracy of the total contents of the analyzed elements was established  
179 using the NIST 2709 (San Joaquin Soil) and NIST 2710 (Montana Soil) reference  
180 materials. The analytical precision was evaluated by five replicate analyses of each soil  
181 sample and expressed as percent relative standard deviation (% RSD).

182 A chemical extraction procedure was used to determine the concentrations of As, Cd, Co,  
183 Cu, Pb, Sb, U and Zn in the bioavailable (extractable) fraction of soil samples. This  
184 extraction based on the protocol (step A) of the Standards, Measurements and Testing  
185 Programme (formerly BCR, Bureau Community of Reference) of the European  
186 Commission, was performed adding 40 mL of a 0.11M CH<sub>3</sub>COOH (acetic acid) solution to  
187 1 g of powdered soil sample. The mixture was shaken for 16 h at room temperature and



188 centrifuged. To validate the analytical results of extraction procedure, the concentrations of  
189 Cd, Cr, Cu, Ni, Pb and Zn were determined in the BCR-701 reference material.  
190 The GBW 07603 (Bush branches and leaves) and NIST RM 8413 (Corn Kernel-Zea mays)  
191 reference materials were utilized to check the accuracy of analytical determinations in  
192 maize samples. The analytical precision was determined by means of five replicate  
193 analyses of each maize sample.

## 194 195 **2.5. Geochemical and biological indices**

196 To assess the overall soil contamination level, the Pollution Load Index (PLI) developed  
197 by Tomlinson et al. (1980), was employed. The value of PLI was calculated by the  $n$ -root  
198 of the product of the Concentration Factor (CF) of the  $n$  chemical elements considered:  $PLI$   
199  $= n\sqrt{(CF_1 \times CF_2 \times CF_3 \times \dots \times CF_n)}$ . The CF of a chemical element was given by  
200  $[C]_{\text{element}}/[C]_{\text{background value}}$ , where:  $[C]_{\text{element}}$  = element concentration in soil,  $[C]_{\text{background value}}$   
201 = average value of local geochemical background of the element in soil. According to  
202 Singh et al. (2003), PLI values vary as follows:  $PLI = 0$  background concentration;  $0 < PLI$   
203  $\leq 1$  uncontaminated;  $1 < PLI \leq 2$  moderately to uncontaminated;  $2 < PLI \leq 3$  moderately  
204 contaminated;  $3 < PLI \leq 4$  moderately to highly contaminated;  $4 < PLI \leq 5$  highly  
205 contaminated;  $PLI > 5$  very highly contaminated.

206 The mobility in soil of the analyzed chemical elements was assessed using the Mobility  
207 Factor Index (MFI; Kabala and Singh 2001; Narwal et al. 1999) calculated as the ratio  
208 between the bioavailable concentration of the element and its total content in soil.

209 In the current scientific literature, the plant-to-soil bioconcentration ratios are indices used  
210 to evaluate the transfer of a chemical element from soil to plant, comparing concentrations  
211 in vegetation parts to concentrations in soil. In this study the Biological Concentration  
212 Factor (BCF; Yoon et al. 2006) was calculated as the ratio of the element concentration in  
213 maize roots to its total content ( $BCF_{\text{tot}}$ ) and bioavailable concentration ( $BCF_{\text{bio}}$ ) in soil.  
214 Similarly, the Biological Accumulation Coefficient (BAC; Cui et al. 2007; Li et al. 2007)  
215 was determined as the ratio of the element concentration in maize grains to its total content  
216 ( $BAC_{\text{tot}}$ ) and bioavailable concentration ( $BAC_{\text{bio}}$ ) in soil.

217 The Translocation Factor (TF) as the ratio of the element concentration in plant shoots to  
218 that in roots (Cui et al. 2007; Li et al. 2007), was used to evaluate the effectiveness of the  
219 maize plants in translocating As, Cd, Co, Cu, Pb, Sb, U and Zn from roots to grains.

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## 222 **2.6. Statistical analysis**

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2 223 The analytical data were treated statistically using the software Statistica 8 (Statsoft,  
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4 224 Padova). The Shapiro-Wilk W normality test was used to check the normal distribution of  
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6 225 the element contents in soil and maize samples. The test results showed that most of data  
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8 226 had a non normal distribution ( $p < 0.05$ ). The non-parametric Mann-Whitney U test was  
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10 227 used to check the difference of the median values of the analyzed chemical elements  
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12 228 between the two groups of soils and maize plants defined according to soil contamination  
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14 229 level. The Spearman's correlation coefficient was employed to measure the correlation  
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16 230 between the variables. A multivariate statistical approach was applied to verify the  
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18 231 geochemical characterization of soil samples. Principal Component Analysis (PCA) was  
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20 232 applied to the standardized data of element concentrations (total and bioavailable) in soil  
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22 233 sample. Soil samples were clustered based on the case coordinates in respect to the  
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24 234 principal axis (components 1 and 2) of PCA analysis. The distances between objects were  
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26 235 defined by the Euclidean distances and the objects were linked together by Ward's method.  
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## 30 **3. Results and discussion**

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### 32 **3.1. Potentially toxic elements in soil**

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34 241 The sampled soils were loam soils formed from the Quaternary alluvial sediments of the  
35 242 Ibar and Sitnica Rivers, and, to a lesser extent, clay loam soils derived from the Pliocene  
36 243 sediments cropping out in the Sitnica valley. Soil pH usually ranged between 6.9 and 7.9  
37 244 (neutral to alkaline soils) and carbonate content between 0.5 and 2%  $\text{CaCO}_3$  (slightly  
38 245 calcareous soils). Soil samples had a very high organic carbon content ( $C_{\text{org}} > 2.2\%$ ) and a  
39 246 high cation exchange capacity (CEC usually from 24 to 42  $\text{cmol}_{(+)}\text{kg}^{-1}$ ). The cation  
40 247 exchange capacity increased as the organic carbon content and clayey fraction increased.

41 248 In order to define the contaminants and the contamination levels in soils of K. Mitrovica  
42 249 area, the total contents of As, Cd, Co, Cu, Pb, Sb, U and Zn in soil samples were first  
43 250 compared with the respective local natural variability in soil (geochemical background)  
44 251 using the Concentration Factor (CF). The geochemical background of As, Cd, Co, Cu, Pb,  
45 252 Sb, U and Zn in soils of the study area (alluvial plains of the Ibar and Sitnica Rivers) was  
46 253 estimated using the total contents of these PTEs in uncontaminated subsoil samples (below  
47 254 90 cm depth) derived from the alluvial sediments of the Ibar and Sitnica Rivers (Borgna et  
48 255 al. 2009). This approach was applied as soil contamination by PTEs extended for tens of

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256 kilometers involving the topsoil of the study area, and affected soil profiles usually up to  
257 50-60 cm deep (Borgna et al. 2009; Šajin et al. 2013).

258 Using the CF scale proposed by Sutherland (2000), Cd, Pb and Sb were moderately to  
259 significantly enriched elements in soil as the CF values were usually above 2 (0.8 to 17.5  
260 for Cd, 0.9 to 20.4 for Pb, 0.4 to 7.5 for Sb). The CFs of As (0.4-21.3), Cu (0.7-2.7) and Zn  
261 (0.4-7.5) indicated geogenic abundances to moderate enrichments. Co and U were not  
262 enriched in soil samples as CF values were below 2 (0.6 to 2.2 for Co, 0.4 to 1.4 for U).

263 The highest enrichments of As, Cu, Pb and Sb were found in soils collected around the  
264 Zvecan smelter within a distance of about 5 km from the plant. Cd and Zn were more  
265 enriched in soils close the Trepča factory. As a general trend, the enrichment levels of As,  
266 Cd, Cu, Pb, Sb and Zn in soil decreased as the distance from the industrial district of K.  
267 Mitrovica (Zvecan smelter and Trepča factory) increased.

268 According to the scale proposed by Singh et al. (2003), the values of Pollution Load Index  
269 (PLI) pointed out that soil samples were usually contaminated by PTEs (mainly Cd, Pb and  
270 Sb), and contamination levels were variable from slightly to highly contaminated soils.  
271 Based on the PLI values, it was possible to divide soil samples in two groups: the first  
272 group included uncontaminated to moderately contaminated soils ( $0 < \text{PLI} \leq 2$ ), hereafter  
273 named slightly contaminated soils ( $n=14$ ); the second group included moderately to highly  
274 contaminated soils ( $2 < \text{PLI} < 5$ ), hereafter named contaminated soils ( $n=10$ ).

275 Table 1 reports the geochemical features of both slightly contaminated and contaminated  
276 soils expressed by min, max, mean and median of As, Cd, Co, Cu, Pb, Sb, U and Zn total  
277 contents. Statistically significant differences between the median of PTE total contents in  
278 soils of two groups ( $p < 0.05$ ) were found.

279 Multivariate statistical techniques such as Principal Component Analysis (PCA) and  
280 cluster analysis were also applied for soil characterization. PCA indicated that the first  
281 factor was defined by the soil contaminants as the following order:  $\text{Sb} > \text{Pb} > \text{Cd} > \text{Cu} > \text{Zn} > \text{As}$   
282 (Fig. 2a). Co and U described the second factor suggesting a geogenic origin of these  
283 elements. Soil samples (cases) coordinates were used as variables for soil classification by  
284 cluster analysis. The results confirmed the above described division of soil samples in  
285 slightly contaminated and contaminated soils (Fig. 2b).

286 As shown in Table 1, slightly contaminated and contaminated soils had also distinctive  
287 bioavailable concentrations of the analyzed PTEs. The differences were statistically  
288 significant ( $p < 0.05$ ) for the contaminants (As, Cd, Cu, Pb, Sb and Zn). On the contrary, the

289 concentrations of the geogenic elements (Co and U) were rather uniform in both soil  
290 groups.

291 The multivariate statistical approach (PCA and cluster analyses) applied to the bioavailable  
292 concentrations of As, Cd, Co, Cu, Pb, Sb, U and Zn confirmed the same grouping of soil  
293 samples in slightly contaminated and contaminated soils obtained using the total contents  
294 of these PTEs.

295 The bioavailable fraction of PTEs was constantly higher in contaminated soils than in the  
296 slightly contaminated ones. The most significant difference was found for Zn, whose  
297 bioavailable percentage amount in contaminated soils was on average double than in  
298 slightly contaminated soils (13% vs 6.5%).

299 On the basis of Mobility Factor Index (MFI), the order of mobility of the potentially toxic  
300 elements in contaminated soils was: Cd>>Zn>Co>Pb>As≈Cu>Sb>U, similar to that in  
301 slightly contaminated soils: Cd>>Co>Zn>Pb>As≈Cu>Sb>U. In both groups of soils Cd  
302 was by far the most bioavailable PTE.

303

### 304 **3.2. Potentially toxic elements in maize**

305 Table 2 reports the main statistical indices of As, Cd, Co, Cu, Pb, Sb, U and Zn  
306 concentrations in roots and grains of maize specimens grouped according to the  
307 contamination level of soil. Analytical data indicated that the analyzed PTEs had the  
308 highest concentrations in the roots of maize plants grown in contaminated soils.  
309 Nevertheless, statistical analysis by means the Mann-Whitney U test revealed that  
310 significant differences ( $p<0.05$ ) were only between the concentrations of contaminants (As,  
311 Cd, Cu, Pb, Sb and Zn) in maize roots from slightly contaminated and contaminated soils.  
312 In maize roots grown in slightly contaminated soils, the order of PTE concentrations  
313 (Zn>>Cu>Pb>>Co>As>Cd>U>Sb) partly differed from that of maize plants from  
314 contaminated soils (Zn>>Pb>Cu>As>>Co>Cd>Sb>U). These findings suggested that soil  
315 contamination caused a substantial increase of root uptake of PTEs resulting in root  
316 reservoir for toxic metals, especially As, Cd, Pb and Sb.

317 No statistical difference ( $p<0.05$ ) was found for the concentrations of PTEs in grains of  
318 maize plants cultivated in slightly contaminated and contaminated soils (Table 2).  
319 However, the average concentrations of contaminants were higher in maize grains from  
320 contaminated sites.

321 In all maize plants the concentrations of As, Cd, Co, Pb, Sb and U in grains were  
322 significantly lower than in roots, suggesting that maize was able to limit the transfer of

323 these non-essential elements from root to edible part (Table 2). Conversely, Cu and Zn  
324 concentrations were comparable in maize grains and roots likely because these elements  
325 are plant micronutrients.

326 The concentrations of As, Cd and Pb in maize grains were compared with the maximum  
327 permissible levels set by the European Commission Regulation for foodstuff (European  
328 Commission 2006) and feedstuff (European Commission 2002). European Commission  
329 established the maximum permissible level in cereals for human consumption at 0.1 mg kg<sup>-1</sup>  
330 for Cd and 0.2 mg kg<sup>-1</sup> for Pb (fresh weight). The maize grains sampled in this study had  
331 always Pb concentrations over the legislation limit of 0.2 mg kg<sup>-1</sup>, while 36% of Cd  
332 concentrations in grains from slightly contaminated soils and 50% from contaminated soils  
333 exceeded the maximum permissible level of 0.1 mg kg<sup>-1</sup>.

334 European limits in products for animal feed are given for As (2 mg kg<sup>-1</sup>), Cd (1 mg kg<sup>-1</sup>)  
335 and Pb (10 mg kg<sup>-1</sup>). As these values are referred to foods with 12% humidity, As, Cd and  
336 Pb concentrations in maize grains were normalized to this level of humidity to allow a  
337 correct comparison. Analytical data showed that no grain samples exceeded As, Cd and Pb  
338 European limits for feedstuff.

339 As suggested by Andersson and Pettersson (1981) and Carbonell et al. (2011), the  
340 concentration of Zn in maize grains should be less than 34 mg kg<sup>-1</sup> to be fit for human  
341 consumption. Zn exceeded this indicative threshold in 50% of grains from contaminated  
342 soils.

343

### 344 ***3.3. Potentially toxic elements from soil to maize***

345 The values of the Biological Concentration Factor (BCF) and Biological Accumulation  
346 Coefficient (BAC) showed a large variability depending on the chemical element, maize  
347 part, use of total or bioavailable concentration and soil contamination level (Tab. 3).  
348 Nevertheless, it has to be noted that the BCF<sub>tot</sub> and BCF<sub>bio</sub> values of the analyzed PTEs  
349 (except for Zn) were higher than the respective BAC<sub>tot</sub> and BAC<sub>bio</sub>. This finding confirmed  
350 an important restriction of the internal transport of PTEs from roots to grains in maize  
351 plants, likely due to the metal-binding capacity of root system (Carbonell et al. 2011; Singh  
352 and Agrawal 2007).

353 There was no statistically significant difference between the BCF<sub>tot</sub> and BCF<sub>bio</sub> of As, Cd,  
354 Co, Cu, Pb, Sb and U in maize roots grown in slightly contaminated and contaminated  
355 soils. The only exception is given by Zn, as the BCF<sub>tot</sub> and BCF<sub>bio</sub> values of this element  
356 were significantly higher in maize plants from slightly contaminated soils (Tab. 3). A

357 possible explanation is that the high concentrations of Cd and Cu in contaminated soils  
358 competitively inhibited the uptake of Zn by roots (Choudhary et al. 1995; Hart et al. 2002;  
359 Kabata-Pendias and Pendias 1984; Mullins and Sommers 1986; Wang et al. 2007).  
360 This study confirmed that the bioavailable concentrations and  $BCF_{bio}$  predict the transfer of  
361 PTEs from soil to plant better than the total content and  $BCF_{tot}$ . Statistical analysis  
362 indicated significant inverse correlations ( $p < 0.05$ ) between  $BCF_{bio}$  and bioavailable  
363 concentration in soil for the analyzed PTEs (except for Cd), while correlations between the  
364  $BCF_{tot}$  and total concentration in soil were found only for As, Cu, U and Zn. The lack of  
365 significant correlation for Cd could be due to its high geochemical mobility in soil and the  
366 ability of cultivated plants to tolerate high concentrations of the element without any  
367 symptom of toxicity (Dudka et al. 1995). Likewise,  $BAC_{bio}$  values were negatively  
368 correlated with bioavailable concentration in soils for all the analyzed PTEs.  
369 As a whole, the Translocation Factor (TF) of PTEs was comparable in maize plants from  
370 slightly contaminated and contaminated soils (Tab. 3). Only the TF values of Cu and Pb  
371 were statistically different and higher in maize grown in slightly contaminated sites (Tab.  
372 3). This evidence could be attributed to greater competition among PTEs during  
373 translocation from roots to grains at high concentrations (Greger 1999).  
374 The TF values in maize plants from slightly contaminated and contaminated soils varied as  
375 the following order:  $Zn > Cu > Cd > U > Sb > Pb > Co > As$ . FTs were usually  $< 0.5$  and only Zn  
376 had values about 1. These results highlighted that maize plants are able to store higher  
377 amounts of PTEs (except Zn) in roots than in grains.  
378 As immobilization of PTEs in roots is mainly related to exclusion strategy (Baker 1981;  
379 Bose and Bhattacharyya 2008), the highest TF values of Cu and Zn can be attributed to  
380 their essential functions in plant tissues. Cu is necessary for carbohydrate and nitrogen  
381 metabolism, and Zn plays a key role as structural constituent or regulatory cofactor of a  
382 wide range of different enzymes and proteins in many important biochemical pathways.  
383 Among the non-essential elements, Cd showed the highest TFs as more than 40% of Cd  
384 taken up by maize roots was translocated to grains. Translocation of Cd in maize plants  
385 was significantly greater than for the other non-essential elements. This fact could be  
386 explained by the chemical similarity of Cd and Zn which can lead to their interaction in  
387 root uptake, transport from roots to aboveground parts, and accumulation in edible parts  
388 (Das et al. 1997). The lowest TFs of As proved that the movement of this element along  
389 the conductive system of maize was strongly limited. This evidence is likely related to the  
390 presence of a self-detoxification mechanism in maize plant which prevents As

391 translocation to the aboveground parts of plant. Baig et al. (2010) reported that As  
392 translocation in different tissues of maize was in the order roots>shoots>grains.

393

394

#### 395 **4. Conclusions**

396 This biogeochemical field study carried out in the industrial area of Kosovska Mitrovica  
397 highlighted that soil contamination influenced uptake, translocation and accumulation of  
398 potentially toxic elements in maize plants.

399 In the study area agricultural soil was contaminated by potentially toxic elements (As, Cd,  
400 Cu, Pb, Sb and Zn) mainly due to the emissions from industrial plants such as the Zvecan  
401 smelter and Trepča factory. Soil contamination by As, Cd, Cu, Pb, Sb and Zn was variable  
402 from slightly to highly contaminated soils, and influenced both the bioavailable fraction  
403 and accumulation of these PTEs in maize roots and grains.

404 The comparison of PTE concentrations in roots and grains suggested that maize was able  
405 to limit the translocation of non-essential elements (As, Cd, Pb and Sb) to edible parts,  
406 likely due to the metal-binding capacity of plant roots. Conversely, the levels of  
407 micronutrients (Cu and Zn) were comparable in maize roots and grains.

408 The plant-to-soil bioconcentration indices suggested that the bioavailable concentrations  
409 predict better the transfer of the potentially toxic elements from soil to plant, and identified  
410 some competitions and interactions among these elements in root uptake and root-to-grain  
411 translocation.

412 In the study area contamination of agricultural soils by potentially toxic elements affected  
413 the quality of maize crops: Cd and Pb concentrations in maize grains exceeded the  
414 maximum permissible limits for human consumption established by the European  
415 Commission Regulation for foodstuff.

416

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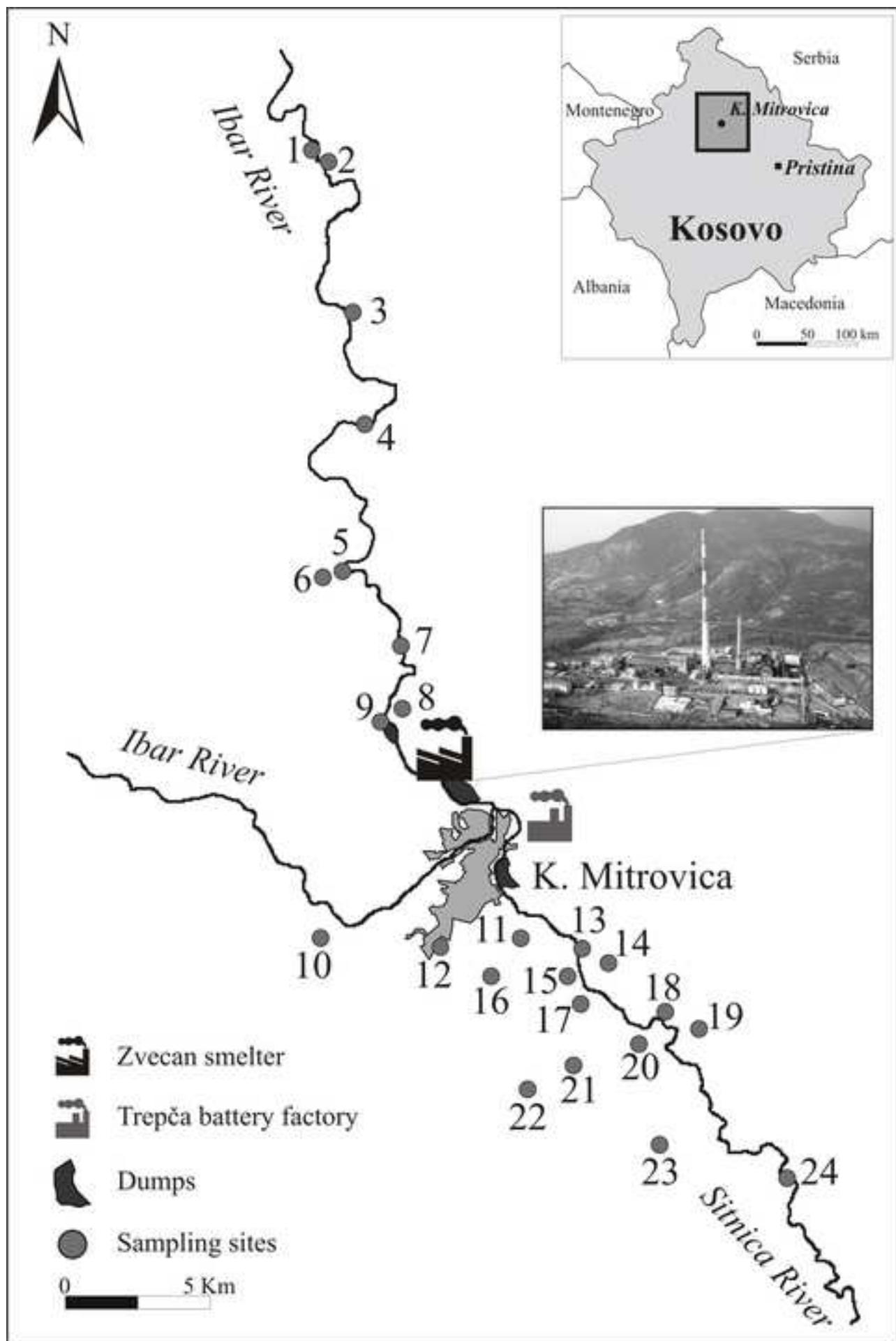
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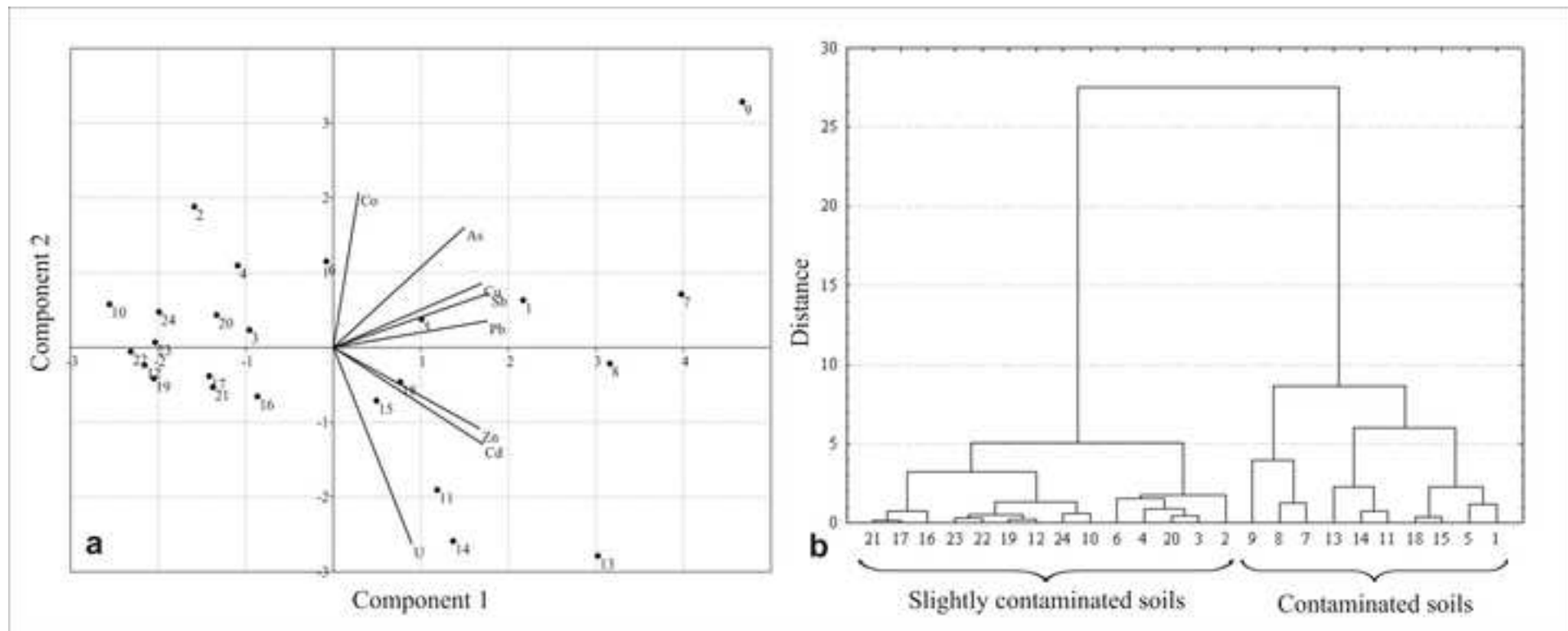
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## FIGURE CAPTIONS

**Fig. 1.** Sketch map of study area. Figure reports the geographic distribution of sampling sites (grey circle), as well as the location of Zvecan smelter, Trepča factory and urban area of Kosovska Mitrovica

**Fig. 2.** Soil grouping by multivariate statistical approach. (a) PCA biplot shows a projection of the variables (potentially toxic elements) onto the scatter plot (projection of soil samples on the factorial plain); (b) Tree-clustering of soil samples based on the case coordinates in respect to the principal axis (components 1 and 2) of PCA analysis. The distances between objects were defined by the Euclidean distances and the objects were linked together by Ward's method





1 **Table 1** Descriptive statistic of As, Co, Cd, Cu, Pb, Sb, U and Zn total contents and  
 2 bioavailable concentrations in soil samples grouped according to the Pollution Load Index  
 3 (data in mg/kg<sup>-1</sup>; *n* = number of soil samples)  
 4

Element		Slightly contaminated soils ( $0 < \text{PLI} \leq$ ; $n = 14$ )					Contaminated soils ( $2 < \text{PLI} < 5$ ; $n = 10$ )				
		Minimum	Maximum	Mean	Median	St. Dev.	Minimum	Maximum	Mean	Median	St. Dev.
As	tot	10.2	54.9	26	23.4 *	11.8	30	501	148	84.5 *	162
	bio	0.05	1.2	0.38	0.27 *	0.33	0.37	2.4	1.6	1.7 *	0.6
Cd	tot	0.37	1.8	1.1	1.2 *	0.45	2.9	8.2	4.5	4.3 *	1.4
	bio	0.11	0.68	0.36	0.35 *	0.18	0.76	2.6	1.5	1.5 *	0.59
Co	tot	12.8	51.6	25.9	20.7	11.7	14.2	43.5	25.9	20.6	11.5
	bio	0.4	4	1.8	1.6	1.2	0.1	5	1.8	1.2	1.6
Cu	tot	22.3	53.8	34.5	32.8 *	10.7	42.7	79.3	60.8	59.3 *	12.9
	bio	0.26	0.54	0.36	0.37 *	0.08	0.52	2.9	1.2	0.81 *	0.89
Pb	tot	72.1	698	293	220 *	182	512	1677	915	756 *	425
	bio	0.49	22.3	7.6	4.2 *	7.8	2.9	200	44.6	22.4 *	59.4
Sb	tot	1.5	8.9	4.8	4 *	2	7.4	26.2	13.4	10.5 *	6.9
	bio	0.01	0.16	0.05	0.04 *	0.04	0.05	0.63	0.24	0.17 *	0.22
U	tot	1	2.4	1.9	1.8 *	0.37	1.6	3.4	2.5	2.4 *	0.61
	bio	0.005	0.03	0.01	0.008	0.007	0.005	0.02	0.01	0.01	0.007
Zn	tot	66.6	377	233	240 *	77.3	473	1131	711	647 *	218
	bio	1.3	41.6	15.5	12.6 *	12.1	31	230	99.3	85.8 *	60.6

tot: total potentially toxic element contents in soil.

bio: bioavailable potentially toxic element concentrations in soil.

\* Significant differences between the median contents of potentially toxic elements in slightly contaminated and contaminated soils ( $p < 0.05$ ).



6 **Table 2** Descriptive statistic of As, Co, Cd, Cu, Pb, Sb, U and Zn concentrations in roots  
 7 and grains of maize plants grown in slightly contaminated and contaminated soils (data in  
 8 mg kg<sup>-1</sup> fresh weight; *n* = number of soil samples)  
 9

Element		Slightly contaminated soils ( $0 < \text{PLI} \leq 2$ ; $n = 14$ )					Contaminated soils ( $2 < \text{PLI} < 5$ ; $n = 10$ )				
		Minimum	Maximum	Mean	Median	St. Dev.	Minimum	Maximum	Mean	Median	St. Dev.
As	roots	0.23	1.3	0.51	0.43 *	0.28	0.44	4.3	1.8	1.4 *	1.4
	grains	0.005	0.05	0.02	0.02	0.01	0.01	0.28	0.06	0.03	0.08
Cd	roots	0.11	1.2	0.32	0.23 *	0.32	0.14	2.5	0.83	0.47 *	0.81
	grains	0.02	0.25	0.1	0.07	0.08	0.04	0.3	0.13	0.1	0.08
Co	roots	0.24	1.4	0.78	0.74	0.32	0.36	1.7	0.93	0.83	0.44
	grains	0.02	0.31	0.06	0.04	0.08	0.02	0.11	0.06	0.06	0.04
Cu	roots	1	11.3	4.2	3.9 *	2.3	3.9	10.8	6.3	5.4 *	2.3
	grains	1.4	3.3	2.3	2.2	0.57	1.4	4.6	2.6	2.3	1.1
Pb	roots	1.4	11.8	3.7	2.6 *	2.9	3.0	25.3	9.7	8.3 *	6.9
	grains	0.22	0.65	0.44	0.39	0.13	0.25	1.5	0.58	0.5	0.36
Sb	roots	0.03	0.13	0.06	0.05 *	0.03	0.07	0.48	0.22	0.15 *	0.17
	grains	0.001	0.04	0.01	0.01	0.01	0.003	0.06	0.02	0.01	0.02
U	roots	0.06	0.33	0.11	0.08	0.08	0.07	0.16	0.11	0.09	0.03
	grains	0.02	0.04	0.03	0.03	0.01	0.02	0.03	0.03	0.03	0.002
Zn	roots	12.8	55	26.7	23.6 *	12.3	22.2	56.4	36	32.6 *	12.2
	grains	14.7	38.7	26.9	26.8	5.5	21.7	59.7	33.9	30.8	12.2

\* Significant differences between the median contents of potentially toxic elements in roots and grains of maize plants grown in slightly contaminated and contaminated soils ( $p < 0.05$ ).

11 **Table 3** Bioconcentration, accumulation and transfer of potentially toxic elements from  
 12 soil to roots and grains of maize plants from slightly contaminated and contaminated soils  
 13

Maize from:	BCFs		BACs		TFs		
	Slightly contaminated soils	Contaminated soils	Slightly contaminated soils	Contaminated soils	Slightly contaminated soils	Contaminated soils	
As	tot	0.02	0.02	0.001	0.001	0.06	0.06
	bio	2.7	1	0.14	0.04		
Cd	tot	0.28	0.17	0.09	0.03	0.43	0.45
	bio	0.92	0.53	0.35	0.09		
Co	tot	0.03	0.04	0.003	0.003	0.09	0.09
	bio	0.76	1.3	0.05	0.16		
Cu	tot	0.13	0.11	0.07	0.05	0.67 *	0.44 *
	bio	12.3	7.3	6.5	3.3		
Pb	tot	0.02	0.01	0.002	0.001	0.18 *	0.09 *
	bio	1.5 *	0.51 *	0.22	0.03		
Sb	tot	0.02	0.01	0.003	0.002	0.28	0.16
	bio	1.7	1.1	0.41	0.16		
U	tot	0.06	0.04	0.02	0.01	0.35	0.3
	bio	14.1	12.6	3.8	3.6		
Zn	tot	0.14 *	0.05 *	0.14	0.05	1.1	0.96
	bio	3.5 *	0.48 *	3.8	0.45		

Bioconcentration, accumulation and transfer of potentially toxic elements from soil to maize were evaluated in terms of Biological Concentration Factor (BCF), Bioaccumulation Coefficient (BAC) and Translocation Factor (TF).

$$\text{BCF} = [\text{element}]_{\text{root}} / [\text{element}]_{\text{soil}}$$

$$\text{BAC} = [\text{element}]_{\text{grain}} / [\text{element}]_{\text{soil}}$$

$$\text{TF} = [\text{element}]_{\text{grain}} / [\text{element}]_{\text{root}}$$

tot: BCF<sub>tot</sub> and BAC<sub>tot</sub> calculated using the total potentially toxic element contents in soil.

bio: BCF<sub>bio</sub> and BAC<sub>bio</sub> calculated using the bioavailable potentially toxic element concentrations in soil.

\* Significant differences between BCFs, BACs and TFs values of maize plants grown in slightly contaminated and contaminated soils ( $p < 0.05$ ).