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# Ion-triggered selectivity in bacterial sodium channels

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## Abstract

Since the availability of the first crystal structure of a bacterial Na<sup>+</sup> channel in 2011, understanding selectivity across this family of membrane proteins has been the subject of intense research efforts. Initially, free energy calculations based on Molecular Dynamics simulations revealed that, while sodium ions can easily permeate the channel with their first hydration shell almost intact, the selectivity filter is too narrow for efficient conduction of hydrated potassium ions. This steric view of selectivity was subsequently questioned by microsecond atomic trajectories, which proved that the selectivity filter appears to the permeating ions as a highly-degenerate, liquid-like, environment. While this liquid-like environment looks optimal for rapid conduction of Na<sup>+</sup>, it seems incompatible with efficient discrimination between similar ion species, such as Na<sup>+</sup> and K<sup>+</sup>, through steric effects. Here, extensive MD simulations, combined with Markov State Model analyses, reveal that at positive membrane potentials, potassium ions trigger a conformational change of the selectivity toward a non-conductive metastable state. It is this transition of the selectivity filter, and not steric effects, which prevents the outward flux of K<sup>+</sup> at positive membrane potentials. This description of selectivity, triggered by the nature of the permeating ions, might have ~~wide~~ implications on the current understanding of how ion channels, and in particular bacterial Na<sup>+</sup> channels, operate at the atomic scale.

## Significance

Voltage-gated Na<sup>+</sup>-channels are essential components of the cell membrane in any realm of life. In order to perform their biological functions, Na<sup>+</sup>-channels need to conduct Na<sup>+</sup> at high rates, while at the same time blocking K<sup>+</sup>. Molecular Dynamics simulations showed that the selectivity filter of bacterial Na<sup>+</sup>-channels is a highly flexible structure. This feature favors fast Na<sup>+</sup> conduction. However, it is still unknown how such highly-flexible structure is able to select Na<sup>+</sup> over K<sup>+</sup>. In this contribution, we show that in the presence of K<sup>+</sup>, the selectivity filter switches to a non-conductive state. The effect of K<sup>+</sup> on the dynamics of Na<sup>+</sup>-channels explains how these proteins can be contemporary highly-permeable to some ion species and highly-selective for similar ones.

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## **Introduction**

Voltage-gated sodium channels are membrane proteins that open in response to depolarization of the cell membrane, and when open, they preferentially conduct  $\text{Na}^+$  over other monovalent and divalent cations. The bacterial channel NaVAb was the first experimental atomic structure of a voltage-gated  $\text{Na}^+$  channel solved by X-ray crystallography (1). The structures of two other bacterial channels, NaVRh (2) and NaVMs (3), were subsequently obtained, and more recently, in 2017, the structure of the first eukaryotic  $\text{Na}^+$  channel was solved by cryo-electron microscopy (4). Thanks to the availability of these experimental structures, at present, it is possible to investigate the mechanisms of conduction and selectivity in voltage-gated sodium channels at atomistic detail.

Sodium channels share the same fourfold architecture common to voltage-gated  $\text{K}^+$  and  $\text{Ca}^{2+}$  channels (5)(6), with the region responsible for the selective conduction, the so-called selectivity filter, at the extracellular entrance of the pore (Figure S1). Remarkably, the selectivity filter differs between eukaryotic and prokaryotic channels. In eukaryotic channels, selectivity to  $\text{Na}^+$  over  $\text{K}^+$  requires the presence of the conserved signature sequence DEKA, while in the prokaryotic homologous the DEKA signature is substituted by a conserved ring of glutamate residues (EEEE), [and -c](#)Consequently, [also](#) the mechanisms of conduction and selectivity are likely to be different between prokaryotic and eukaryotic sodium channels. The high-density of negative charge facing the lumen of the pore of bacterial  $\text{Na}^+$  channels, in combination with the width of the pore, suggest that more than one cation is likely to bind simultaneously inside the selectivity filter. In agreement with this hypothesis, free energy profiles estimated from Molecular Dynamics (MD) simulations confirmed that the selectivity filter corresponds to a deep free energy minimum for  $\text{Na}^+$ , and that conduction events with only one ion are hampered by high free energy barriers (7)(8). In contrast, when two sodium ions are considered, a low-energy pathway emerges across the selectivity filter. Free energy calculations considering permeation of two  $\text{K}^+$  or mixtures of  $\text{K}^+$  and  $\text{Na}^+$  revealed a remarkable difference between the two ion-species: the region of the selectivity filter at the intracellular side of the EEEE ring (between Leu52 and Glu53 in NaVMs), which is a free energy minimum for sodium ions, is a free energy barrier for potassium ions. This small difference in free energy, which is related to the bigger size of  $\text{K}^+$  compared to  $\text{Na}^+$ , might easily explain the selectivity for sodium ions. This description of conduction and selectivity was confirmed by extensive MD simulations in the presence of negative membrane potentials (9). The conductance values of NaVAb (10) and NaVMs (9)(11) estimated by MD simulations with negative membrane potentials are in satisfactory agreement with experimental data. The average number of ions

occupying the selectivity filter in these trajectories ranges between 1.7 and 2.0, confirming that conduction is likely to involve the presence of two  $\text{Na}^+$  inside the selectivity filter. Moreover, in simulations with  $\text{K}^+$ , the conductance of the channel was lower than in simulations with  $\text{Na}^+$ , in agreement with the experimental behavior of the channel. The main difference between the two ionic species was ascribed to a region in the middle of the selectivity filter, with much lower density for  $\text{K}^+$  than for  $\text{Na}^+$  (9). This low-density region corresponds to the free energy barrier observed for  $\text{K}^+$  but not  $\text{Na}^+$ , at the intracellular side of the EEEE ring in free energy calculations, confirming the hypothesis that selectivity for  $\text{Na}^+$  over  $\text{K}^+$  results from the exclusion of (bigger) potassium ions from the core of the selectivity filter.

The mechanism of  $\text{Na}^+/\text{K}^+$  selectivity described in the previous paragraph, and based on steric effects, was questioned by equilibrium MD simulations of the NaVA channel in the microsecond time scale (12), where it was shown that residues of the EEEE motif might switch between two states (Figure S1), with the side chains directed respectively to the extracellular entrance of the channel (out-facing) or the pore lumen (dunked). The relative probability of these two configurations is strongly correlated with the number of  $\text{Na}^+$  inside the selectivity filter, with higher ion occupancies favoring the dunked state. The same degeneracy of states, and correlation between ion occupancy and structure of the filter, was later observed in free energy calculations where more than two permeating ions and longer MD trajectories were considered (13)(14). Not surprisingly, transitions of the glutamate residues between the out-facing and the dunked states have an effect on the free energy barriers of conduction events, and consequently on selectivity. When the degeneracy of states is considered, the difference in free energy between  $\text{Na}^+$  and  $\text{K}^+/\text{Na}^+$  mixtures largely disappears (13). As a consequence of the high-mobility of the side chains lining the pore, the selectivity filter appears to the permeating ions as a disordered region. This liquid-like environment looks optimal for efficient conduction of  $\text{Na}^+$ . However, it is less obvious how a highly-flexible selectivity filter might discriminate between sodium and similar potassium ions.

The ring of glutamate residues of the EEEE motif is located in the region where most of the voltage drop between the extracellular and the intracellular compartment is focused (15)(16)(17). Thus, the membrane potential is likely to impact on the transitions between the out-facing and the dunked state, and as a result, on conduction and selectivity. In agreement with this hypothesis, MD simulations revealed differences between inward (negative membrane potential) and outward (positive membrane potential) conduction. The number of sodium ions inside the selectivity filter is higher in simulations of outward conduction, as well as the probability of the dunked state of residues in the EEEE motif (11). These simulations of outward conduction were performed with membrane potentials close to 500 mV, and only in the presence of sodium ions. In contrast, the

behavior of the EEEE motif at positive membrane potentials in a physiological range, and in the presence of potassium ions, is still unexplored by MD simulations.

The role of the EEEE motif on conduction and selectivity is investigated in the present study by an extensive set of MD simulations with membrane potentials at  $\pm 100$  mV in the presence of sodium or potassium ions. The mechanisms of ion selectivity at positive and negative membrane potentials are found to be different. At negative membrane potentials, selectivity to  $\text{Na}^+$  over  $\text{K}^+$  arises from the exclusion of  $\text{K}^+$  from the central region of the selectivity filter, as previously suggested. Instead, at positive membrane potentials, the presence of  $\text{K}^+$  gives rise to a non-conductive metastable state of the selectivity filter, which is absent in analogous simulations with  $\text{Na}^+$ . This mechanism of selectivity – where transitions among metastable states are triggered by the nature of the permeating ion – is exploited to discriminate similar ion species, with ~~possible~~<sup>important</sup> implications on the current understanding of ion channels at the atomic level.

## Results

### Inward conduction of sodium ions

The estimated conductance for the inward flux of  $\text{Na}^+$  in MD simulations with membrane potential at  $-100$  mV is  $32 \pm 14$  pS, in qualitative agreement with experimental data and previous MD simulations (Table 1 and Table S1) (9). The selectivity filter is most likely occupied by two (57.7%) or one sodium (39.8%) ion (Table S2). However, complete conduction events might involve the temporary binding of a third ion (Figure 1 and S2). These triple occupancy states are short-lived, and they represent less than 3% of the simulated time. On average, the number of ions interacting with atoms of the selectivity filter is 1.6, distributed among three main binding sites (Figure 1), in agreement with previous calculations (7)(9). The side chains of Glu53 residues do not deviate significantly from the experimental structure for the entire simulated time (1.5  $\mu\text{s}$ ). Rapid movements of Glu53 toward the dunked configuration were observed, but these states were never stable for more than a few nanoseconds. In accordance with this description, the position of the carbon atom of the carboxyl group of Glu53 residues (atom CD in Figure S1) exhibits a single density peak, corresponding to the out-facing configuration (Figure 1).

### Inward conduction of potassium ions

In MD simulations with  $\text{K}^+$  and membrane potential at  $-100$  mV, the conductance is roughly half of the value estimated for sodium ions under the same conditions, being 17 pS with  $\text{K}^+$  versus 32 pS with  $\text{Na}^+$  (Table 1). The ion occupancy of the selectivity filter also differs between  $\text{K}^+$  and  $\text{Na}^+$  (Table S2). For both ions, the selectivity filter hosts two ions most likely. However, the probability of 3-ion states is higher for  $\text{K}^+$  than for  $\text{Na}^+$  (11.9 versus 2.5), as well as the average number of ions

inside the selectivity filter (1.9 versus 1.6). The difference between the two ion species emerges clearly at the extracellular side of the selectivity filter. The high-density peak between Leu52 and Glu53 in simulations with  $\text{Na}^+$  disappears in simulations with  $\text{K}^+$  (Figure 1). The extracellular entrance of the selectivity filter is still an attractive region for  $\text{K}^+$ , but ions preferentially bind between Glu53 and Ser54. Despite these differences in ion binding, the dynamics of the selectivity filter is similar in simulations involving both ion species, with side chains of Glu53 residues stable in the out-facing configuration (Figure 1 and S2). This outward facing configuration of Glu53 is critical for the selectivity mechanism that was proposed on the basis of free energy calculations, as it creates an energy barrier between Leu52 and Glu53 for  $\text{K}^+$  but *not* for  $\text{Na}^+$ . In agreement with this selectivity mechanism, the ion density has a deep minimum at the intracellular side of the EEEE ring for  $\text{K}^+$  but not for  $\text{Na}^+$  (Figure S2). Exclusion of  $\text{K}^+$  from the core of the selectivity filter impairs ion movements across the channel, rendering natural selectivity for  $\text{Na}^+$  over  $\text{K}^+$  for inward fluxes.

### **Outward conduction of sodium ions**

Outward conduction of sodium ions is slightly different from inward conduction, as evidenced by the different number of ions inside the selectivity filter and the dynamic behavior of Glu53 residues. The presence of three ions inside the selectivity filter, which was a rare event in simulations with negative membrane potential, has a probability of 16.7% in simulations with membrane potential at +100 mV (Table S2). As a consequence, the average number of ions in the selectivity filter increases from 1.6 to 2.0. This increase in ion-occupancy is associated with a higher mobility of Glu53 residues. The side chains of Glu53 switch between out-facing and dunked configurations on the same time scale of conduction events (Figure 2 and S3), with the dunked state more likely associated with the binding of three ions inside the selectivity filter. The position of the carbon atom of the carboxyl group of Glu53 residues exhibits a wider distribution compared to simulations with negative membrane potential (compare Figure 1c with 2c), which is representative of the degeneracy of states of the selectivity filter. Despite this different behavior of the selectivity filter between positive and negative membrane potentials, the position of the binding sites for sodium ions is almost unaffected, and the estimated conductance of the channel is also close to the value calculate at -100 mV (Table 1). Therefore, while the mechanisms of conduction exhibit some minor differences, the NaVMs channel conducts  $\text{Na}^+$  at similar rates in inward and outward directions.

### **Outward conduction of potassium ions**

In simulations with  $\text{K}^+$  and membrane potential at +100 mV, the rate of permeation showed high variability among different trajectories (Table S1). Therefore, in order to better explore the causes responsible for this variability, a higher number of independent trajectories was simulated for

outward conduction of  $K^+$  than for the other tested conditions. The average conductance, as estimated from eight independent MD simulations and a cumulative simulation time of 3.4  $\mu$ s, is 24 pS (Table 1). This value is ~30% lower than the conductance estimated for  $Na^+$  under the same conditions. However, while in simulations with sodium ions the value of the conductance is relatively stable among independent simulations (24-40 pS), in simulations with potassium ions the conductance ranges from 3 pS (1 conduction event in a 500 ns trajectory) to 61 pS (19 conduction events in a 500 ns trajectory). The average ion occupancy of the selectivity filter is greater in simulations with  $K^+$  than in simulations with  $Na^+$  (Table S2). In the most likely configuration, the selectivity filter is occupied by three ions (46.9%), but states with four ions are also observed with significant probability (6.9%). Moreover, the behavior of Glu53 residues is also different from the one observed in any other tested condition. The dunked state of Glu53 has higher probability than in simulations with  $Na^+$ , and these states are stable for hundreds of nanoseconds (Figure 2 and S4). The differences between  $K^+$  and  $Na^+$  observed in simulations with a non-polarizable force field (CHARMM36) were confirmed by two independent set of simulations with the Drude polarizable force field (Figure S5).

Visual inspection of the trajectories in simulations with  $K^+$  and positive membrane potential immediately suggests a link between the rate of ion conduction and the configuration of Glu53 residues. For instance, in the trajectory shown in Figure 2b, all the conduction events take place when the carbon atoms of the carboxyl group of Glu53 residues are above  $z = 5 \text{ \AA}$ . In contrast, long-lived states with these atoms below this threshold correspond to non-conducting periods of the channel. A similar behavior is observed in the other trajectories (Figure S4). Therefore, a possible explanation for the high variability of the estimated conductance in this set of MD simulations is that the selectivity filter exists in separate metastable states with different functional properties. This hypothesis is investigated in the next section.

### **Metastable states of the selectivity filter**

Use of Markov State Models (MSM) is a possible strategy to identify metastable states in MD simulations (18) (19). The MD trajectories with  $K^+$  and positive membrane potential were converted into sequences of discrete microstates using the configuration of the Glu53 residues. The absence of significant memory effects, necessary for an accurate estimate of the MSM, was tested by comparing the relaxation times calculated with increasing sampling periods (20). The relaxation times were marginally affected by the sampling period for time intervals above 10 ns (Figure S6a), which was consequently used as lag-time for estimating the MSM. The ordered sequence of relaxation times computed from the transition matrix of the MSM exhibits a clear gap between the



first and the second relaxation time (Figure 3a). Therefore, a hidden Markov State Model with two states was estimated using the discretized MD trajectories (21).

The hidden states of the MSM correspond to different metastable states of the selectivity filter, which were named respectively  $E_{OUT}$  and  $E_{IN}$ , in agreement with the most likely configuration of the glutamate residues of the EEEE ring. In state  $E_{OUT}$ , Glu53 residues are preferentially directed toward the extracellular entrance of the channel (Figure 3b-c), but excursions of one glutamate residue to the dunked state are frequently observed (Figure S6b). This configuration of the selectivity filter is most likely occupied by two or three ions, respectively with probability 54.9% and 38.1% (Table S2), and the binding sites are remarkably similar to the ones observed at negative membrane potential (compare Figure 3c with Figure 1d). Instead, in metastable state  $E_{IN}$ , the distribution of ions inside the selectivity filter differs from the one observed at negative membrane potential, or with sodium ions (Figure 3d). The selectivity filter is mainly occupied by three ions (68.4%), and the probability of four ions interacting with the filter is 26.1% (Table S2). The major difference between ion distributions in the two metastable states emerges at the center of the selectivity filter, which in state  $E_{IN}$  is completely depleted from  $K^+$ . This non-canonical distribution of ions is related to a structural change of Glu53 residues. In metastable state  $E_{IN}$ , the most likely configuration of the selectivity filter has two glutamate residues protruding toward the axis of the channel (Figure S6b). These configurations of Glu53 residues prevent the binding of ions at the center of the selectivity filter. Ions, instead, accumulate in an off-axis position, where they directly interact with the carbonyl oxygen atoms of the glutamate residues (Figure S6c).

The different structures and ion occupancies of the selectivity filter in the two metastable states suggest an effect on the conduction properties. Indeed, the vast majority of the outward conduction events of  $K^+$  occur when  $E_{OUT}$  is the most likely metastable state of the channel. In detail,  $E_{OUT}$  is the most likely metastable state for approximately 2.5  $\mu$ s, and during this period of time, 49 conduction events are observed, while, a single conduction event is observed during the 0.9  $\mu$ s when  $E_{IN}$  is the most likely metastable state (Table 1). Therefore, an approximate estimate of the channel conductance in two metastable states is respectively 31 pS for  $E_{OUT}$  and less than 2 pS for  $E_{IN}$ . The conductance of state  $E_{OUT}$  is remarkably similar to the value estimated in analogous simulations with  $Na^+$  (35 pS). This lack of selectivity is the natural consequence of the high mobility of Glu53 residues in simulations with positive membrane potential. Indeed, it is important to remark that in state  $E_{OUT}$ , the out-facing configuration of residues Glu53 is more likely than in state  $E_{IN}$ , but Glu53 residues are not confined to this out-facing configuration, instead, they are free to sample several different microscopic states (Figure S6b). In agreement, the distribution of the carbon atom of the carboxyl group of Glu53 residues in state  $E_{OUT}$  closely resembles the analogous distribution

in simulations with  $\text{Na}^+$  and positive membrane potential (Figure 2c). The side chains of Glu53 residues rapidly interconvert among different states, and this liquid-like environment is not suited for selecting between two ionic species with identical charge and similar radius, as  $\text{K}^+$  and  $\text{Na}^+$  are. Therefore, the lack of selectivity of this degenerate state is not surprising. The apparent contradiction between a liquid-like environment and the ability of the selectivity filter to discriminate similar ionic species is resolved by conformational transitions to the metastable state  $E_{\text{IN}}$ . In state  $E_{\text{IN}}$ , the mobility of Glu53 residues is much lower than in state  $E_{\text{OUT}}$ . The side chains of Glu53 residues in two opposing subunits are blocked in an inward configuration, where they prevent the passage of ions. This locked state is stable for hundreds of nanoseconds. Similar transitions of the selectivity filter were not observed in simulations with sodium ions. While it is not possible to exclude that similar states could emerge also in the presence of sodium ions, the consistent behavior observed among  $\text{Na}^+$  simulations, and the different ion occupancy of the selectivity filter between the two ion-species, also confirmed by simulations with an alternative force field, seem to suggest that  $\text{Na}^+$  and  $\text{K}^+$  can actually interfere with the dynamics of the selectivity filter. In the presence of sodium ions, a unique – conductive – stable state of the selectivity filter is observed. In contrast, in the presence of potassium ions, an alternative – non-conductive – metastable state of the selectivity filter emerges. Switching between the conductive ( $E_{\text{OUT}}$ ) and the non-conductive ( $E_{\text{IN}}$ ) metastable states (Figure 3b) could reduce the permeability of  $\text{K}^+$ , making the channel selective for  $\text{Na}^+$ .

## Discussion

At present, the overall consensus about conduction and selectivity in bacterial  $\text{Na}^+$  channels extracted from MD simulations is primarily focused on two atomic models that are, at least apparently, at conflict with each other. The first atomic model entails a stable selectivity filter, where the glutamate residues of the EEEE motif do not deviate significantly from the experimental structure. In this model, selectivity for  $\text{Na}^+$  over  $\text{K}^+$  arises from steric effects (7)(8). In the second atomic model, the glutamate residues of the EEEE motif rapidly switch among different configurations (12). As a result of this degeneracy of states, the selectivity filter mimics a liquid-like environment for the permeating sodium ions. This liquid-like environment guarantees high sodium permeability. However, it is currently unknown how such a flexible selectivity filter might discriminate  $\text{Na}^+$  over  $\text{K}^+$ . In this study, permeation in bacterial  $\text{Na}^+$  channels was investigated, for the first time, by a set of MD simulations with different ion species and physiological membrane potentials ( $\pm 100$  mV). The MD trajectories confirmed that both the hypothesized behaviors of the selectivity filter, either rigid or flexible, are possible. At negative membrane potentials, the selectivity filter does not deviate considerably from the experimental structure, and the lower

permeability to  $K^+$  over  $Na^+$  might easily be explained by steric effects. Instead, at positive membrane potentials, the glutamate residues of the EEEE motif move between different configurations. The high mobility of the selectivity filter has marginal effects on the permeability of sodium ions, which is similar for negative and positive membrane potentials. In contrast, the presence of potassium ions stabilizes alternative structures of the selectivity filter with remarkably low conduction rates. These transitions between different metastable states of the selectivity filter, rather than steric effects, are the main determinants of selectivity for  $Na^+$  over  $K^+$  at positive membrane potentials.

The presence of alternative mechanisms of selectivity for inward and outward ion fluxes is in qualitative agreement with electrophysiological experiments, which demonstrated that the selectivity of bacterial  $Na^+$  channels depends on the direction of the concentration gradients (22). Despite this qualitative agreement, the quantitative comparison between MD simulations and experimental data is hampered by several issues. Firstly, in electrophysiological experiments, the selectivity for  $Na^+$  over  $K^+$  is usually estimated by measuring the reversal potential with the channel exposed to different concentration gradients for the two ion-species. Instead, in MD simulations, selectivity was analyzed by comparing  $Na^+$  and  $K^+$  trajectories at the same membrane potential and without concentration gradients. The transitions between metastable states observed in trajectories with  $K^+$  might be the result of a direct effect of the positive membrane potential on the glutamate residues, or of ion movements in the outward direction. The second hypothesis is supported by steered MD simulations, which showed that the selectivity filter of bacterial  $Na^+$  channels behaves asymmetrically for inward and outward fluxes, even at null membrane potential (23). The quantitative comparison with experimental data requires a thorough understanding of the individual contributions of the direction of ion fluxes and of the membrane potential to selectivity, and this is not possible with the simulation protocol employed in the present study. Algorithms to mimic concentration gradients in MD simulations have been proposed (24)(25). However, in order to estimate reversal potentials, which could then be directly compared with experimental data, simulations of ion fluxes under different electrochemical gradients, some of them close to equilibrium, are required. The high computational cost of MD simulations makes the estimate of reversal potentials in voltage-gated channels a daunting task. Consequently, MD simulations at constant ion concentrations were performed.

The second issue that might hamper a quantitative comparison between simulations and experiments is the accuracy of the force fields adopted for MD simulations of biomolecules. The  $Na^+/K^+$  permeability ratio critically depends on two factors: (i) the relative permeability of the two ion-species when the glutamate residues of the EEEE motif are in the out-facing configuration, and

(ii) the probabilities of the different metastable states of the selectivity filter. At present, it is uncertain if classical force fields can render accurate estimates of these relative probabilities. The variability among simulations of bacterial Na<sup>+</sup> channels with different force fields well represents the current situation. For instance, the Na<sup>+</sup>/K<sup>+</sup> permeability ratio at -100 mV is close to 10 when ion parameters by Joung et al are adopted (9), while a much lower ratio was estimated here, using the ion parameters by Roux et al (26). Despite these quantitative differences, it is important to remark that the picture emerging from independent MD simulations is coherent. Analogous binding sites have been identified, regardless of the simulation method and the adopted force field, and these binding sites are in good agreement with experimental data (7)(8). The number of ions inside the selectivity filter, and the differences between inward and outward permeation pathways for Na<sup>+</sup>, coincide in several computational studies (10)(14). Moreover, the sodium conductance estimated from MD simulations is conserved in different force fields, and it is in agreement with experimental data (9)(10)(11). Although, current simulation protocols and the classical force fields might not be good enough for a quantitative comparison with experimental data about selectivity, the agreement with experiments for sodium ions, together with the coherent picture emerging from simulations using different force fields, strongly support the hypothesis that the mechanisms of conduction and selectivity extracted from these computational studies are indeed representative of how ion channels operate at atomic resolution.

The mechanism of selectivity observed in simulations with positive membrane potential is intrinsically different from the ones previously proposed for bacterial Na<sup>+</sup> channels, or even other ion channels. Previous mechanisms of selectivity proposed on the bases of MD simulations can be classified into two main categories, here, referred to as local selectivity and multi-ion selectivity for clarity. The term local selectivity is used when selectivity can be accounted for by the presence of a specific region along the pore that favors some particular ionic species. The selectivity for Na<sup>+</sup> over K<sup>+</sup> for inward fluxes at negative membrane potentials in bacterial Na<sup>+</sup> channels is an example of local selectivity. Sodium ions are favored over potassium ions because they better fit in the region at the intracellular side of the EEEE ring. In other ion channels, it is not possible to explain selectivity by the presence of (local) selective binding sites. For instance, the selectivity filter of K<sup>+</sup> channels exhibits a sequence of binding sites selective for K<sup>+</sup>, alternated by a sequence of binding sites selective for Na<sup>+</sup> (27). Selectivity depends on the multi-ion conduction mechanism: single files of K<sup>+</sup> are energetically favored over single files of ion mixtures (Na<sup>+</sup>/K<sup>+</sup>) (28)(29). Therefore, in K<sup>+</sup> channels, selectivity cannot be ascribed to any specific position along the pore, because it emerges as a global property of the channel due to the multi-ion conduction mechanism. The mechanism of selectivity proposed here for outward ion fluxes in bacterial Na<sup>+</sup> channels is neither a local property

nor due to multi-ion conduction. Instead, it is caused by the effect of the permeating ions on the dynamics of the selectivity filter, and consequently it will be referred to as ion-triggered selectivity. This concept of ion-triggered selectivity has been proposed before to describe how the selectivity filter of ion channels reacts to the permeating ion species. For instance, the selectivity filter of some  $K^+$  channels collapses in the presence of sodium ions, and this might add a second layer of selectivity over the one created by the multi-ion conduction mechanism described above (30). In addition, an effect of the permeating ion species on the dynamics of ion channels has been observed in previous MD simulations of other ion channels, and also related to selectivity (31)(32)(33). However, to the best of our knowledge, this is the first time that selectivity is explained by a transition of the selectivity filter to a different metastable state induced by a particular permeating ion species. In this respect, ion-triggered selectivity could easily explain why an ion channel might appear as a liquid-like, highly-conductive, environment to some ion species, while at the same time blocking other ion species with high efficiency. As selectivity is the fingerprint of ion channels, ion-triggered selectivity might be a common mechanism, playing an important role in the function of this class of membrane proteins.

## Methods

MD simulations were performed using NAMD, version 2.11 (34). The simulation systems consisted of the pore domain of NaVMs (PDB code: 3ZJZ), that was aligned along the z-axis and embedded in a POPC lipid membrane. The CHARMM36 force field for protein and lipids was used (35)(36)(37)(26)(38)(39), and water molecules were described with the TIP3P model (40). The set of simulations included several independent trajectories in the NVT ensemble with different ion species, NaCl or KCl, and membrane potentials,  $\pm 100$  mV, as listed in Tables S1. In order to check for the dependency of the results on the force field, a set of simulations with the Drude polarizable force field was also performed (41)(42)(43). The membrane potential was mimicked by a constant electric field acting along the z-axis. PyEMMA was used to estimate the MSM (44), using as discrete features the coordinates of the carbon atoms of the carboxyl group of Glu53 residues. For additional details about simulation and analyses protocols see SI Methods.

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## References

1. Payandeh J, Scheuer T, Zheng N, Catterall WA (2011) The crystal structure of a voltage-gated sodium channel. *Nature* 475(7356):353–359.
2. Zhang X, et al. (2012) Crystal structure of an orthologue of the NaChBac voltage-gated sodium channel. *Nature*. doi:10.1038/nature11054.
3. McCusker EC, et al. (2012) Structure of a bacterial voltage-gated sodium channel pore reveals mechanisms of opening and closing. *Nat Commun* 3:1102.
4. Shen H, et al. (2017) Structure of a eukaryotic voltage-gated sodium channel at near-atomic resolution. *Science* (80- ):eaa14326.
5. Doyle DA, et al. (1998) The structure of the potassium channel: molecular basis of K<sup>+</sup> conduction and selectivity. *Science* (80- ) 280(5360):69–77.
6. Wu J, et al. (2015) Structure of the voltage-gated calcium channel Cav1.1 complex. *Science* (80- ) 350(6267):aad2395-aad2395.
7. Furini S, Domene C (2012) On conduction in a bacterial sodium channel. *PLoS Comput Biol* 8(4):e1002476.
8. Corry B, Thomas M (2012) Mechanism of ion permeation and selectivity in a voltage gated sodium channel. *J Am Chem Soc* 134(3):1840–1846.
9. Ulmschneider MB, et al. (2013) Molecular dynamics of ion transport through the open conformation of a bacterial voltage-gated sodium channel. *Proc Natl Acad Sci* 110(16):6364–9.
10. Stock L, Delemotte L, Carnevale V, Treptow W, Klein ML (2013) Conduction in a biological sodium selective channel. *J Phys Chem B* 117(14):3782–3789.
11. Ke S, Timin EN, Stary-Weinzinger A (2014) Different Inward and Outward Conduction Mechanisms in NaVMs Suggested by Molecular Dynamics Simulations. *PLoS Comput Biol* 10(7). doi:10.1371/journal.pcbi.1003746.
12. Chakrabarti N, et al. (2013) Catalysis of Na<sup>+</sup> permeation in the bacterial sodium channel Na(V)Ab. *Proc Natl Acad Sci U S A* 110(28):11331–6.
13. Boiteux C, Vorobyov I, Allen TW, Na M (2014) Ion conduction and conformational flexibility of a bacterial voltage-gated sodium channel. *Proc Natl Acad Sci* 111(9):3454–3459.
14. Domene C, Barbini P, Furini S (2015) Bias-Exchange Metadynamics Simulations: An Efficient Strategy for the Analysis of Conduction and Selectivity in Ion Channels. *J Chem Theory Comput* 11(4):1896–1906.
15. Contreras JE, et al. (2010) Voltage Profile along the Permeation Pathway of an Open Channel. *Biophys J* 99(9):2863–2869.
16. Berneche S, Roux B (2003) A microscopic view of ion conduction through the K<sup>+</sup> channel. *Proc Natl Acad Sci* 100(15):8644–8648.
17. Furini S, Zerbetto F, Cavalcanti S (2007) Role of the Intracellular Cavity in Potassium Channel Conductivity. *J Phys Chem B* 111(50):13993–14000.
18. Bowman GR, Pande VS, Noé F (2014) An Introduction to Markov State Models and Their Application to Long Timescale Molecular Simulation. *Springer* 797:148.
19. Prinz J-H, et al. (2011) Markov models of molecular kinetics: Generation and validation. *J Chem Phys* 134(17):174105.

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20. Swope WC, et al. (2004) Describing Protein Folding Kinetics by Molecular Dynamics Simulations. 2. Example Applications to Alanine Dipeptide and a  $\beta$ -Hairpin Peptide <sup>†</sup>. *J Phys Chem B* 108(21):6582–6594.
21. Noé F, Wu H, Prinz JH, Plattner N (2013) Projected and hidden Markov models for calculating kinetics and metastable states of complex molecules. *J Chem Phys* 139(18). doi:10.1063/1.4828816.
22. Finol-Urdaneta RK, et al. (2014) Sodium channel selectivity and conduction: prokaryotes have devised their own molecular strategy. *J Gen Physiol* 143(2):157–71.
23. Ngo V, Wang Y, Haas S, Noskov SY, Farley RA (2016) K<sup>+</sup> Block Is the Mechanism of Functional Asymmetry in Bacterial Nav Channels. *PLoS Comput Biol* 12(1):e1004482.
24. Khalili-Araghi F, Ziervogel B, Gumbart JC, Roux B (2013) Molecular dynamics simulations of membrane proteins under asymmetric ionic concentrations. *J Gen Physiol* 142(4):465–475.
25. Berti C, Furini S, Gillespie D (2016) PACO: PArticle COunting Method to Enforce Concentrations in Dynamic Simulations. *J Chem Theory Comput* 12(3):925–929.
26. Beglov D, Roux B (1994) Finite representation of an infinite bulk system: Solvent boundary potential for computer simulations. *J Chem Phys* 100(12):9050–9063.
27. Kim I, Allen TW (2011) On the selective ion binding hypothesis for potassium channels. *Proc Natl Acad Sci* 108(44):17963–17968.
28. Egwolf B, Roux B (2010) Ion Selectivity of the KcsA Channel: A Perspective from Multi-Ion Free Energy Landscapes. *J Mol Biol* 401(5):831–842.
29. Furini S, Domene C (2011) Selectivity and Permeation of Alkali Metal Ions in K<sup>+</sup>-channels. *J Mol Biol* 409(5):867–878.
30. Zhou Y, Morais-Cabral JH, Kaufman A, Mackinnon R (2001) Chemistry of ion coordination and hydration revealed by a K<sup>+</sup> channel-Fab complex at 2.0 Å resolution. *Nature* 414(6859):43–48.
31. Dixit PD, Merchant S, Asthagiri D (2009) Ion selectivity in the KcsA potassium channel from the perspective of the ion binding site. *Biophys J* 96(6):2138–2145.
32. Burykin A, Kato M, Warshel A (2003) Exploring the origin of the ion selectivity of the KcsA potassium channel. *Proteins Struct Funct Genet* 52(3):412–426.
33. Darré L, Furini S, Domene C (2015) Permeation and dynamics of an open-activated TRPV1 channel. *J Mol Biol* 427(2):537–549.
34. Phillips JC, et al. (2005) Scalable molecular dynamics with NAMD. *J Comput Chem* 26(16):1781–1802.
35. MacKerell AD, et al. (1998) All-Atom Empirical Potential for Molecular Modeling and Dynamics Studies of Proteins <sup>†</sup>. *J Phys Chem B* 102(18):3586–3616.
36. MacKerell AD, Feig M, Brooks CL (2004) Improved Treatment of the Protein Backbone in Empirical Force Fields. *J Am Chem Soc* 126(3):698–699.
37. Klauda JB, et al. (2010) Update of the CHARMM All-Atom Additive Force Field for Lipids: Validation on Six Lipid Types. *J Phys Chem B* 114(23):7830–7843.
38. Luo Y, Roux B (2010) Simulation of Osmotic Pressure in Concentrated Aqueous Salt Solutions. *J Phys Chem Lett* 1(1):183–189.
39. Venable RM, Luo Y, Gawrisch K, Roux B, Pastor RW (2013) Simulations of anionic lipid

membranes: Development of interaction-specific ion parameters and validation using NMR data. *J Phys Chem B* 117(35):10183–10192.

40. Jorgensen WL, Chandrasekhar J, Madura JD, Impey RW, Klein ML (1983) Comparison of simple potential functions for simulating liquid water. *J Chem Phys* 79(2):926–935.
41. Lopes PEM, et al. (2013) Polarizable force field for peptides and proteins based on the classical drude oscillator. *J Chem Theory Comput* 9(12):5430–5449.
42. Lamoureux G, Harder E, Vorobyov I V., Roux B, MacKerell AD (2006) A polarizable model of water for molecular dynamics simulations of biomolecules. *Chem Phys Lett* 418(1–3):245–249.
43. Li H, et al. (2015) Representation of Ion-Protein Interactions Using the Drude Polarizable Force-Field. *J Phys Chem B* 119(29):9401–9416.
44. Scherer MK, et al. (2015) PyEMMA 2: A Software Package for Estimation, Validation, and Analysis of Markov Models. *J Chem Theory Comput* 11(11):5525–5542.



## Figure legends

**Figure 1. Conduction with membrane potential at -100 mV. (a-b)** Trajectory of ions and Glu53 residues in simulations with  $\text{Na}^+$  and  $\text{K}^+$  respectively. Top-panels: ion positions along the axis of the channel (z-axis). The trajectories of ions that cross the channel from side to side are shown using bold lines in different colors. Thin grey lines are used for any other ion in the system that is closer than 12 Å from the channel axis. Other ions are not shown. The black lines show the position of the center of mass of Ala90 residues (intracellular entrance of the channel), and of the side chain oxygens of Ser54 residues (extracellular entrance of the selectivity filter). The zero of the z-axis corresponds to the center of the carbonyl oxygen atoms of Thr51 residues. Representative snapshots of the selectivity filter (residues 51 to 54 of only two opposing subunits in licorice representation), with the permeating ions (VDW representation), and surrounding water molecules (licorice representation) are shown on the top. The colors used for the ions correspond to the ones adopted in the plot. Middle/bottom-panels: trajectories of the carbon atom of the carboxyl group of Glu53 residues respectively along the z-axis and in the radial direction. The radial coordinate was defined as the distance on the x-y plane from the center of mass of the carbonyl oxygen atoms of residues Thr51. **(c-d)** Distribution of ions and Glu53 residues in simulations with  $\text{Na}^+$  and  $\text{K}^+$  respectively. MD trajectories were sampled with a period of 10 ps. The average positions of the carbonyl oxygen atoms of Thr51 and Leu52, the carbon atoms of the carboxyl group of Glu53, and of the side-chain oxygen atoms of Ser54 are highlighted by horizontal dashed lines. The z- and the r-axis were discretized in bins with side equal to 0.25 Å, and the average number of ions, and carbon atom of the carboxyl group of Glu53 (CD atom) residues was computed for each bin. This average number of particles per bin was divided by the volume of the bin to get the densities of ions (blue maps), and of CD atoms (red contour plots).

**Figure 2. Conduction with membrane potential at +100 mV.** Same legend as Figure 1. **(a-b)** The dashed lines in middle/bottom panels show the average position of the carbon atom of the carboxyl group of [Glu53](#) residues ~~Glu53~~ along the axial and radial directions respectively, in simulations with the same ion species and membrane potential at -100 mV. The color bar above panel (b) shows the most likely metastable state of the MSM; green corresponds to  $E_{\text{OUT}}$  and red to  $E_{\text{IN}}$ .

**Figure 3. Markov State Models of the selectivity filter. (a)** Estimated relaxation times of the MSM. **(b)** Representative configurations of the selectivity filter in metastable states  $E_{\text{IN}}$  (red box) and  $E_{\text{OUT}}$  (green box). Two alternative views of residues 51 to 54 (rotated by 90 degrees around the axis of the channel) are shown for two opposite subunits. [Mean first passage times between the two metastable states are provided.](#) **(c-d)** Distribution of ions (blue maps) and of carbon atom of the carboxyl group of Glu53 residues (red contour plots) in metastable states  $E_{\text{OUT}}$  and  $E_{\text{IN}}$  respectively, calculated as described in the legend of Figure 1.