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M. tuberculosis lineages: genetic diversity and its involvement on macrophage infection and on drug tolerance

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on macrophage infection and on drug tolerance**

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Summary

A central feature of *Mycobacterium tuberculosis* pathogenesis is the ability to survive within macrophages and colonize hostile environments that are acidic and rich in cholesterol and fatty acids.

The genetic variability among clinical isolates may have dramatic consequences on the outcome of infections. Many in vitro and in vivo studies have demonstrated strain-dependent variation in key aspects of virulence such as stress survival, transmission, pathology, and lethality.

This variability in MTB clinical isolates went neglected for decades; however, recently the scientific community recognized it causes important consequences on the progression of infection. As reduced host response is supposed to be the key to enabling MTB persistence and transmissibility, the immunological state of the host is also important to consider when studying the relative virulence of clinical MTB strains. In this context, the polarization states of macrophages have a huge impact on their function. It affects how macrophages can react to external signals, changing gene expression, membrane composition, receptor exposure, and cytokines production.

In our work, we focused our attention on MTB diversity and its role in pathogenesis and drug resistance acquisition. We proposed a comprehensive approach for better understanding MTB virulence taking into consideration both human (phenotypic) and MTB (genetic) variability.

We selected well-characterized strains belonging to specific MTB lineages and adopt the THP-I-derived macrophage infection model, but considering host variability by deriving M1 or M2 polarized macrophages.

Confocal live microscopy was used to study at a single-cell level different macrophage pathways during the infection. These included phagolysosomal acidification, autophagic flux, and apoptosis. All are mechanisms that MTB hijacks in order to survive within the host. Cytokine profiles were measured in response to the different MTB lineages, along

with the survival of both the macrophages and the bacteria at different time points post-infection.

In M1 macrophages, whereas less virulent/ancient are more efficient in blocking the phagosomal acidification, more virulent/modern strains are likely better at tolerating the acidic environment. However, the number of responsive macrophages remained relatively low. In M2a macrophages, whereas less virulent/ancient strains are less efficient in blocking the phagolysosomal acidification, more virulent/modern strains are blocking phagolysosomal acidification. The number of responsive macrophages was found to vary depending upon the features of the infecting strains, with the most virulent ones inducing the lowest percentages of acidifying cells.

The analysis of the autophagic flux showed less heterogeneity among both the bacterial strains and the macrophage phenotypes considered. Indeed, the induction of autophagy was found negligible in our observations.

In M1 macrophages, bystander non-infected cells of more virulent/modern showed increased apoptosis (40%). Stratification of the data by mycobacterial burden showed similar apoptotic levels irrespective of the mycobacterial load, despite a trend displaying slightly higher apoptotic levels in bystander non-infected cells or in cells infected with a lower number of mycobacteria could be noted. In contrast, none of the categories induced apoptosis in M2 macrophages at the time point considered.

In M1 cells, the infection with less virulent/ancient lineages reduced the production of pro-inflammatory IL1- β . Likewise, IL-18 levels were lower in M1 cells infected with less virulent/ancient lineages. The chemokine GRO α showed a similar pattern between MTB categories considered. In M2 cells we observed the modulation of anti-inflammatory IL-10 when comparing clinical to laboratory strains. Most of the statistically significant differences found were observed within 24h for both polarization types.

Our findings on macrophage survival corroborate the results obtained at the single-cell level, with more virulent/modern lineages causing increased cell death. Interestingly, these data seem to contrast the finding that such

strains showed lower macrophage entry in colony-forming units evaluation studies. Similarly, there was not a direct correlation between pro-inflammatory cytokine production and macrophage killing. Nevertheless, more virulent/modern strains showed increased intramacrophagic replicative capacity compared to less virulent/ancient lineages.

We also studied the genetic variability of the MTB strains during drug resistance acquisition. Among newly investigated mechanisms, the role of smallRNAs in the development of drug resistance has been considered in other bacteria but remains unexplored in MTB. We characterized the smallRNA *ncRv0842c*, cis-encoded to the *Rv0842* gene which codes for a putative efflux pump reported to be involved in rifampicin resistance (RIF-R) development. Accordingly, the present study characterized the role of *ncRv0842c* during RIF challenge and validated the role of a lineage-specific synonymous mutation on the coding region of *Rv0842* affecting the promoter region of the cis-encoded smallRNA.

In order to understand the role of the smallRNA in modulating the efflux pump, we characterized its expression during challenge experiments using sub-inhibitory concentrations of rifampicin. We then generated overexpressing mutants from MTB-selected ancient and modern strains.

qPCR showed basal downregulation of *ncRv0842* in ancient lineages compared to modern lineages. This result confirmed the hypothesis that the synonymous mutation in *Rv0842* (L45L) specific for the ancient lineages, and abrogating the -10 promoter region of the antisense smallRNA is severely affecting the expression of the smallRNA. Under RIF-induced stress, we observed upregulation of *Rv0842*, and downregulation of the smallRNA only in H37Rv (modern lineage) whereas in L5 (ancient lineage) the smallRNA expression was not affected. MABA assay performed on modern lineage mutants overexpressing *ncRv0842c* showed a 1-dilution reduction of the MIC in comparison with their respective control (mock) while the ancient lineage strains did not show any MIC shift despite the overexpression of the smallRNA. Our analysis showed that the unraveling

of smallRNA may provide new insights on MTB lineage-specific adaptation to drug-related stress and uncover the role of silent mutations in determining different phenotypes in MTB.

Our study can contribute to better understanding lineage-specific pathogenicity and highlights the importance of strain- and lineage-specific rational design and development of effective diagnostic tools and vaccines.

List of abbreviations

- **AM** Alveolar Macrophage
- **AMK** Amikacin
- **BDQ** Bedaquiline
- **CAP** Capreomycin
- **CCL5** CC-chemokine ligand 5
- **CFP** Culture Filtrate Protein
- **CFZ** Clofazimine
- **CXCL** Chemokine CXC ligand
- **DLM** Delamanid
- **EMB** Ethambutol
- **ESAT-6** Early Secretory Antigenic Target 6 (EsxA)
- **ETO** Ethionamide
- **FCR** Fc Receptors
- **FQ** Fluoroquinolone
- **GM-CSF** Granulocyte-Macrophage Colony Stimulating Factor
- **IFN** Interferon
- **IL** Interleukin
- **INH** Isoniazid
- **KAN** Kanamycin
- **LAM** Lipoarabinomannan
- **LEV** Levofloxacin
- **LZN** Linezolid
- **ManLAM** Mannose-capped Lipoarabinomannan
- **M-CSF** Macrophage Colony Stimulating Factor
- **MDR** Multi-drug Resistance
- **MHC** Major Histocompatibility Complex
- **MMP** Matrix Metalloproteinase
- **MOI** Multiplicity of Infection
- **MTBC** Mycobacterium tuberculosis complex

- **NO** Nitric Oxide
- **NOS** Nitrogen Reactive Species
- **ORF** Open Reading Frame
- **OriC** Origin of Replication
- **PAMPS** Pathogen Associated Molecular Patterns
- **PDIM** Phthiocerol dimycocerosate
- **PGL** Phenolic Glycolipid
- **PI3K** Phosphoinositide 3-kinase
- **PIM** Phosphatidylinositol Mannoses
- **PRR** Pattern Recognition Receptors
- **PTKA** Protein Tyrosine Kinase A
- **PZA** Pyrazinamide
- **RIF** Rifampicin
- **RNAP** RNA polymerase
- **ROS** Reactive Oxygen Species
- **SR** Scavenger Receptor
- **STM** Streptomycin
- **TB** Tuberculosis
- **TGF- β** Transforming growth factor beta
- **TH1/2** T helper1/2
- **TLR** Toll Like Receptor
- **TNF** Tumor Necrosis Factor
- **TSS** Transcription Start Site

Chapter 1

Introduction

1.1 Tuberculosis

Tuberculosis (TB) is an airborne infectious disease caused by a group of bacteria belonging to the *Mycobacterium tuberculosis* complex (MTBC) family, of which *M. tuberculosis* (MTB) is the principal etiological agent of the human disease. Recent studies suggest that species belonging to the MTBC were originated in Africa and then diffuse around the globe adapting to the host, in a sort of co-evolutionistic process ¹. This ancient disease has accompanied mankind during its evolution, adapting to the social, demographic changes, and migratory flows that have characterized human history ². The history of TB across the centuries is indeed strictly connected with human demographic growth, migrations, and social behaviors, and, in Europe, was considered as a “real plague” during the industrial revolution. TB in 2021 still represents a global emergency with a high impact on the public health system; furthermore, together with the spread of HIV, the use immunosuppressive therapies, the increasing immigration from high TB endemic countries, and the spread of multidrug-resistant (MDR) strains, TB remains public health concern even in industrialized areas (Fig. 1).

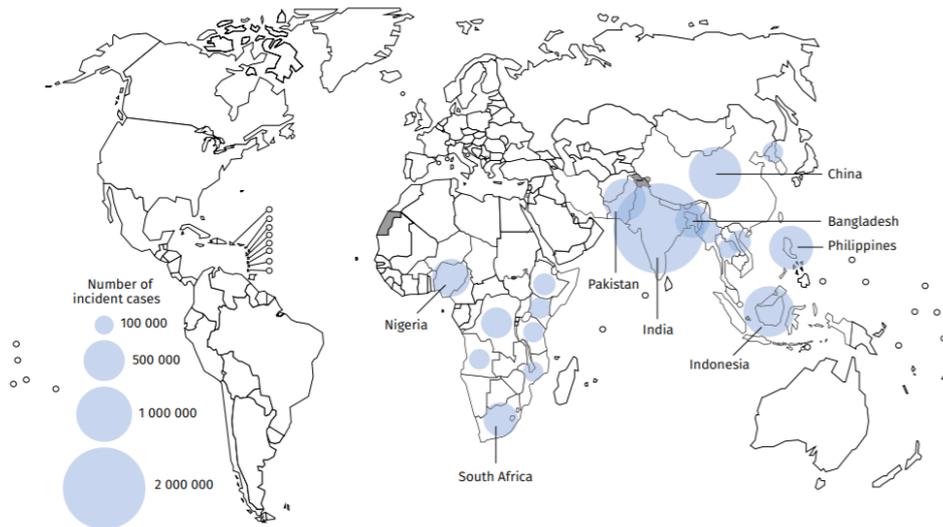


Fig. 1: Estimated TB incidence in 2020, for countries with at least 100,000 incident cases ⁵.

The COVID-19 pandemic has invalidated years of progress we achieved in fighting TB and reducing the TB disease burden. The biggest impact is a large drop in the global number of people newly diagnosed with TB, with a decrease from 7.1 million in 2019 to 5.8 million in 2020 with a substantial fall of 18%, and far short of the approximately 10 million people who developed TB in 2020 ³. COVID-19 reduced the access to TB diagnosis and treatment, increasing the TB death rate. 2021 WHO report estimates for 2020 1.3 million TB deaths among HIV-negative people and an additional 214000 cases among HIV-positive people ³.

1.2 *Mycobacterium tuberculosis*: the etiological agent of TB

Mycobacteria are ubiquitous since they are easily found in water, soil, food and as a pathogen/commensal for animals and humans. The first identification of MTB was attributed to Robert Koch at the end of the XIX century when he demonstrated the correlation and the etiological origin of MTB in TB illness.

Mycobacteria are part of the Mycobacteriaceae family and belong to the Actinobacteria order. The genus *Mycobacterium* includes over 188 species, some of which are pathogenic to humans (MTB, *M. bovis*, *M. leprae*) some others are specific for animals. The genus *Mycobacterium* is categorized into complexes, which are grouped considering similar phenotypical characteristics as their principal hosts and the type of pathology caused. The MTBC includes: MTB *sensu stricto*, *M. africanum*, and *M. canettii* which are responsible for the majority of human TB cases, plus the animal-adapted species *M. bovis*, *M. microti*, *M. caprae*, *M. pinnipedii* ⁴. In addition, *M. canetti*, and other smooth TB bacilli have been added to the complex because of the similarities of the clinical manifestations of the infection, even though their grouping within the complex in terms of phylogenesis is still debated ⁵.

The observation that the human-adapted MTBC population is phylogeographically organized has led to the hypothesis that the different lineages might be adapted to particular human populations; MTBC strains can be classified into generalists and specialists depending on the host range and the bacterial capacity to survive in a determined population. In particular, generalist strains are more globally spread and do not show a particular prevalence on a specific human population; specialists are predominantly found in restricted areas of the globe with a prevalence on a particular human population ⁶. Further details are provided in Section 1.3 “MTB genomic diversity and pathogenicity”. Phylogeny studies suggest the presence of a common ancestor to the MTBC strains. In particular, there is the evidence between a specific absence of a repeated region RD9 and

strains infecting exclusively animals ⁷. The high level of conservation in housekeeping genes revealed that between 15000 and 20000 years ago there was a bottleneck event of speciation promoting the selection of mycobacterial species present nowadays ⁸. Another bottleneck effect seems to have also occurred for the phylogenetic branch that includes the species *M. africanum*, *M. microti*, and *M. bovis* ⁹.

When observed at the microscope, they appear filament-shaped, about 2-4 μm in length and 0.5 μm in width; they are not motile and free of cilia or flagella ¹⁰. The presence of a cell wall rich in mycolic acids gives mycobacteria alcohol-acid-resistance and using Gram staining techniques they are classified as weakly gram-positive. Mycobacteria are non-spore-forming, catalase-positive bacilli with aerobic or microaerophilic metabolism, and they are characterized by a high lipid content. Indeed, lipids in mycobacteria are up to 60% of the dry weight making them the microorganisms with the highest lipid content in the Bacteria kingdom.

Numerous characteristics of mycobacteria (including virulence) have been linked to the uniqueness of their wall, which is characterized by an asymmetrical double layer rich in waxes that acts as a permeability barrier to polar molecules ¹¹. The cell wall can be divided into two layers: the first one composed of peptidoglycan, arabinogalactan, and a mycolic acid core, and a second one with a layer of amphipathic lipids, non-covalently linked to mycolic acids (Fig. 2). The anchoring proteins, phosphatidylinositol mannoses (PIMs), and lipoarabinomannan (LAM) of the outer parts of the MTB cell wall are responsible, among others, for the development of clinical manifestations and can modulate the immune response of the host. In particular, LAM can inhibit protein kinase C activity and can promote the production of numerous macrophage-associated cytokines such as Tumor Necrosis Factor (TNF)- α , Interleukin (IL)-1, IL-6, and IL-10 ¹².

Among cell components, the chordal factor trehalose 6,6-dimycolate is present in virulent *Mycobacterium* species strains. Studies have demonstrated that the chordal factor is a virulence factor, inducing the release of chemokines by macrophages, and granulomatous response ¹³.

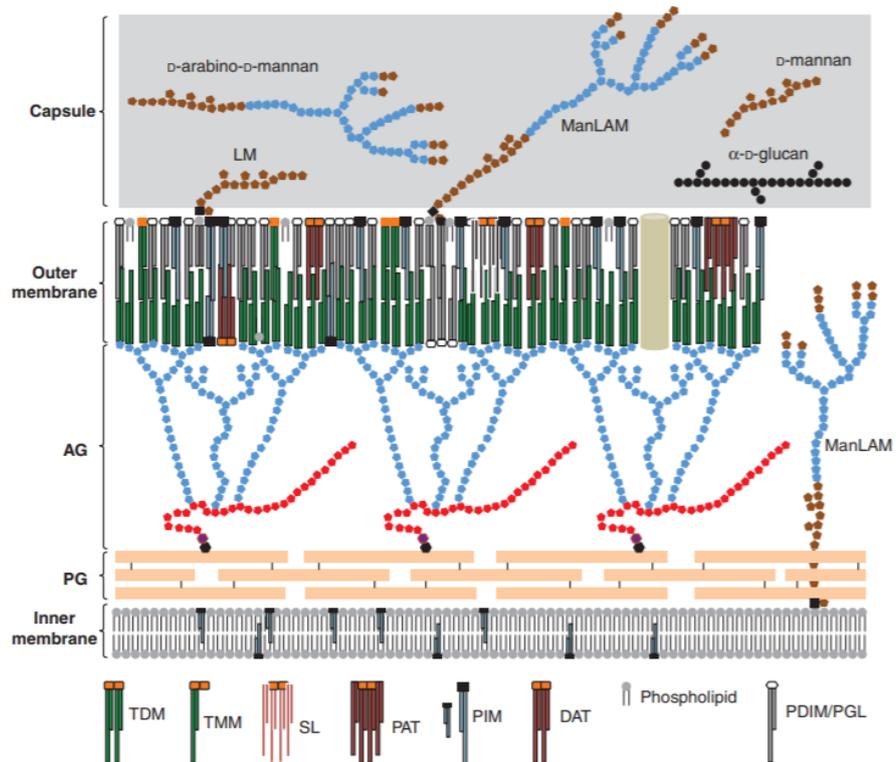


Fig.2: Schematic representation of the MTB cell envelope. Light blue symbols represent arabinose residues, red symbols represent galactose residues, brown symbols represent mannose residues, and black circles represent glucose residues. D-arabino D-mannan, D-glucan, and D-mannan are shown in dark green. LM, lipomannan; ManLAM, manose-capped lipoarabinomannan; AG, arabinogalactan; PG, peptidoglycan ¹¹.

The most widely used MTB reference strain in research laboratories is MTB H37Rv, the genome of which was sequenced in 1998 ¹⁴. MTB H37Rv genome is 4,411,529 base pairs long with high GC content (65.6%). Ribosomal *rrn* RNA gene is present in a single operon, close to the origin of replication site (OriC); this characteristic is common for slow-growing bacteria since having multiple copies of ribosomal operons close to OriC is typical for rapid growing bacteria. Approximately 4000 open reading frames (orf) have been identified by genomic analysis, but recent

proteomic studies revealed the presence of some other possible genes and transcripts not identified before ¹⁵.

MTB can synthesize essential amino acids, vitamins, and cofactors, and it is able to metabolize carbohydrates, hydrocarbons, alcohol ketones, and carboxylic acids. MTB is naturally resistant to several antibiotic compounds. Indeed, its strongly hydrophobic cell wall acts as a permeability barrier that prevents antibiotics to enter the cell. In addition, the organism is provided with many potential drug efflux systems including 14 major facilitator superfamily (MFS) members and numerous ABC transporters that further affect drug uptake ^{14, 16}. Some hydrolytic and modifying enzymes (e.g., β -lactamase and aminoglycoside-acyltransferases) are then encoded in the genome, further reducing the spectrum of antibiotics effective on MTB.

Several virulence factors have been described to be involved in the metabolism of fatty acids, in the synthesis of lipids and mycolic wall acids, in proteins and lipoproteins expressed in the membrane, in enzymes involved in attenuating the oxidative stress in macrophages and secretion systems (Supplementary Table) ¹⁶.

In MTB approximately 2000 putative non-coding smallRNAs have been identified by *in silico* prediction and/or RNA sequencing approaches. However, only a few have been confirmed and characterized in details (Table 1).

Table 1: MTB smallRNAs with a known mechanism of action

MTB smallRNA	Classification	Strand	Surrounding genes	Role
Mcr7 (MTB000067)	Trans-encoded	+	<i>aprABC</i> locus	Mcr7 expression was found to be regulated by the two-component PhoPR system and modulates the translation of <i>tatC</i> gene-regulating its expression and affecting Tat secretory complex ¹³³
MrsI (MTB000142, ncRv11846)	Trans-encoded	+	<i>blal</i> - <i>Rv1847</i>	MrsI was found to respond to several stresses such as lack of iron, oxidative stress, acidic pH, and nutrient deficiency. Deletion of this smallRNA was found to be lethal in an iron-deficient medium ¹³⁴
B11 (ncRv13660c, MTS2822, 6C)	Trans-encoded	-	<i>Rv3660c</i> - <i>Rv3661</i>	B11 secondary structure contains two hairpins, each with 6 consequent cytosine residues in the loop. Its expression is induced by oxidative stress and low pH. Its overexpression is lethal for <i>M. tuberculosis</i> ¹³⁵
F6 (ncRv10243, MTS194, MTB000051)	Trans-encoded	+	<i>fadA2</i> - <i>fadE5</i>	F6 smallRNA expression is upregulated in oxidative stress, hypoxia, low pH, macrophagic infection, and nutrient deficiency ¹³⁶
Mcr11 (MTS0997, MTB000063)	Trans-encoded	-	<i>Rv1264</i> - <i>Rv1265</i>	Mcr11 levels increase upon transition to the stationary growth phase in MTB. Its overexpression causes MTB growth inhibition ¹³⁷
MTS1338 (Drr5, MTB000077)	Trans-encoded	+	<i>Rv1733c</i> - <i>Rv1734c</i>	MTS1338 (Drr5), as for the former smallRNA, is upregulated in the stationary phase of growth and is regulated by the DosR regulon ¹³⁸
MTS2823 (MTB000078)	Trans-encoded	+	<i>Rv3661</i> - <i>Rv3662c</i>	MTS2823 concentration is increased during chronic mice infection. Its overexpression decrease the expression of many genes involved in MTB central metabolism ¹³⁹
ncRv12659 (MTS2048)	Cis-encoded	+	<i>Rv2660c</i>	ncRv12659 is accumulated in mouse infection and during starvation. Its overexpression reduces bacterial growth and changes the transcription of at least 50 MTB gene ¹⁴⁰

These non-coding RNAs with a length ranging from 50 to 400 nucleotides are able to regulate transcription, translation, stability of mRNAs and proteins. smallRNAs can be encoded in cis or in trans to their target genes, and they can act both as activator and repressor (Fig. 3)¹⁷.

These RNAs can act both regulating virulence-associated processes and drug tolerance mechanisms as reported for many bacteria¹⁸. For example, as outlined in Fig. 4, in *E. coli* several sRNAs are involved in the regulation of key processes such as cell envelope synthesis, biofilm formation, antibiotic uptake, and efflux. Among other pathogenic bacteria, main studies are focusing on *Staphylococcus aureus*, *Pseudomonas aeruginosa*, and *Salmonella typhimurium* and participate in complex networks which adapt metabolism in response to external stimuli (Table 2). Despite the evidence derived from other bacterial species about the contribution of non-coding RNAs to both drug resistance and pathogenesis, the data

available on MTB remains very limited and scattered throughout a few pioneer studies.

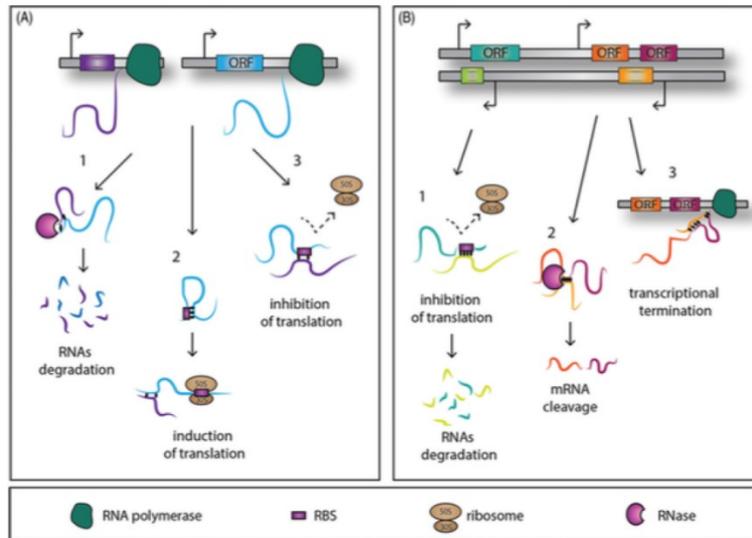


Fig. 3: Simplified model of action of trans-encoded (A) and cis-encoded (B) smallRNAs ¹⁷

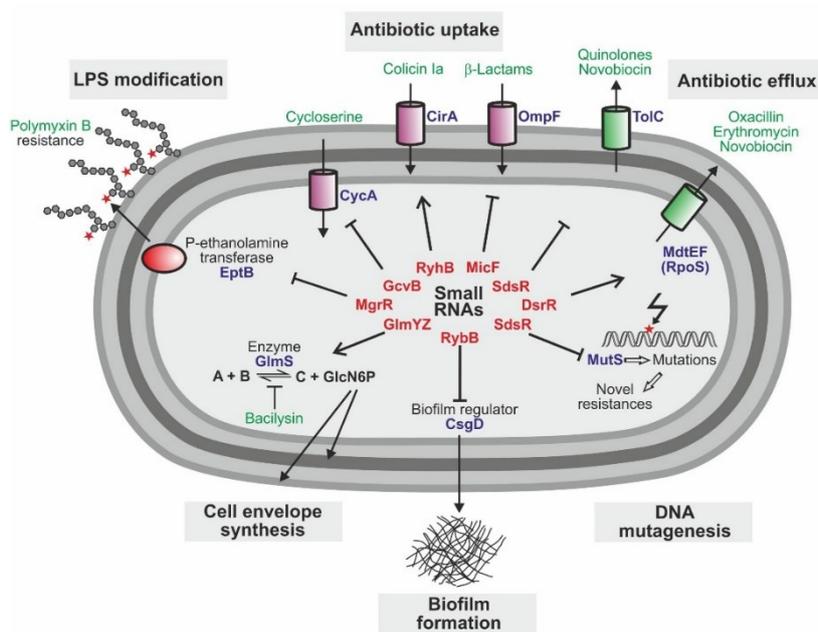


Fig. 4: *E. coli* Trans-encoded smallRNAs and their impact on antibiotic resistance and susceptibility. Trans-encoded smallRNA in *E. coli* are key players in the regulation of antibiotic resistance and are involved in antibiotic uptake, modification of the membrane composition, efflux pump function, production of enzymes, and biofilm formation. SmallRNAs are in red, target proteins in blue, antibiotics in green ¹⁹.

Table 2: bacterial smallRNA involved in pathogenesis mechanisms ²⁰

Bacteria	Diseases	smallRNA (nt)	Target
Gram -positive bacteria			
<i>Staphylococcus aureus</i>	Skin infections, bacteremia, endocarditis, osteomyelitis, toxin-mediated diseases, abscesses, nosocomial infections	RNAIII (514)	hla mRNA spa mRNA rot mRNA SA1000 mRNA
<i>Streptococcus pyogenes</i>	Pharyngitis, skin infections, acute rheumatic diseases, scarlet fever, necrotizing fasciitis, glomerulonephritis	FasX (300) Pel (459)	? ?
<i>Clostridium perfringens</i>	Food poisoning, wound infections, gas gangrene	VR RNA (400) VirX (400)	? ?
Gram -negative bacteria			
<i>Pseudomonas aeruginosa</i>	Burn and wound infections, endocarditis, cystitis, pneumonia in cystic fibrosis patients, septicaemia in immunocompromised patients	RsmY (120) RsmZ (119)	RsmA
<i>Vibrio cholerae</i>	Cholera	CsrB (417) CsrC (366) CsrD (351) Qrr1 (96) Qrr2 (108) Qrr3 (107) Qrr4 (107)	CsrA hapR mRNA
<i>Salmonella typhimurium</i>	Gastroenteritis, enterocolitis, septicaemia in immunocompromised patients	CsrB (350) tmRNA (363)	CsrA
<i>Chlamydia trachomatis</i>	Sexually transmitted genital infections, trachoma	lhtA (120)	hctA mRNA

1.3 Pathogenesis of TB

The respiratory tract is the main entry route MTB bacilli. Spread of the infection can occur via droplet secretions, coughing, or sneezing. About 3000 nuclei (5-10 μm in diameter) of infective droplets are emitted coughing and they can remain suspended in the air for several hours. Once inhaled, MTB overcomes the mucociliary barrier and reaches the alveoli. The probability that a healthy person becomes infected by MTB depends mainly on the aerial concentration of these particles, the duration of exposure, and the subject's immune defenses. The factors that can modify the TB illness fate are related both to the bacillus and to the host. Taking into consideration the statistics, about 1-2% of contacts immediately develop active TB while 1 in 3 will develop a latent form of infection. Among those, only 5-10% of latent infected will progress to an active form of TB and become infective, with a high risk in the first two years following the infection ²¹.

Macrophagic phagocytosis of small bacterial agglomerates (1-3 bacilli) will cause the first contact between MTB and the host innate immune response, and thus activate defense mechanisms. Early phases of the infection are important for the fate of TB when individual differences in the immune response are one of the determining factors in the progression of the disease and in its clinical course.

The interaction between MTB and host cell is highly complex and extremely intimate, and when the infection does not progress to active disease, mycobacteria are cleared or undergo a quiescent phase ²².

As mentioned, macrophages are the first immunological cells responding to MTB. In 90% of the cases, their first contact with MTB activates the immune system and the recruitment of lymphocytes, essential for the formation of a local lesion called granuloma ²³. In immunocompetent people, the infection is controlled by the immune system and the lesions may heal. Primary TB occurs in a person with insufficient immunity to confine and control bacilli in granulomas ²². When activated, T -cells migrate to the lungs and are involved in the granuloma formation together with activated macrophages, fibroblasts, and dendritic cells. From a histological point of view, MTB is circumscribed within a tissue structure consisting of epithelioid cells of macrophage derivation, monocytes, lymphocytes, and neutrophils (Fig. 5).

Granuloma represents an effective way of containment for the proliferation and spread of the pathogen. Within this hostile ambient, macrophages may kill mycobacteria and produce a necrotic central zone. MTB has adapted to survive and persist in this area. After 2-3 weeks this solid necrotic lesion will become caseous, and it's characterized by low levels of oxygen, low pH, and low nutrients. This will slow MTB metabolisms and activates promoter factors that push MTB to undergo a latency phase ²³. In most cases, the granuloma stops growing and heals, leaving a fibrotic and calcified lesion, within which the mycobacterium can be in a quiescent latent phase. The cell-mediated immune response of CD4⁺ T lymphocytes keeps infection under control. Whereas the granuloma

represents a containment measure for the host, it can be also beneficial to the pathogen, as exposure to antibiotics and additional immune cells recruitment is also limited.

In immunologically compromised individuals, both primary or a previously established infection can be reactivated and progress into the active form of the disease.

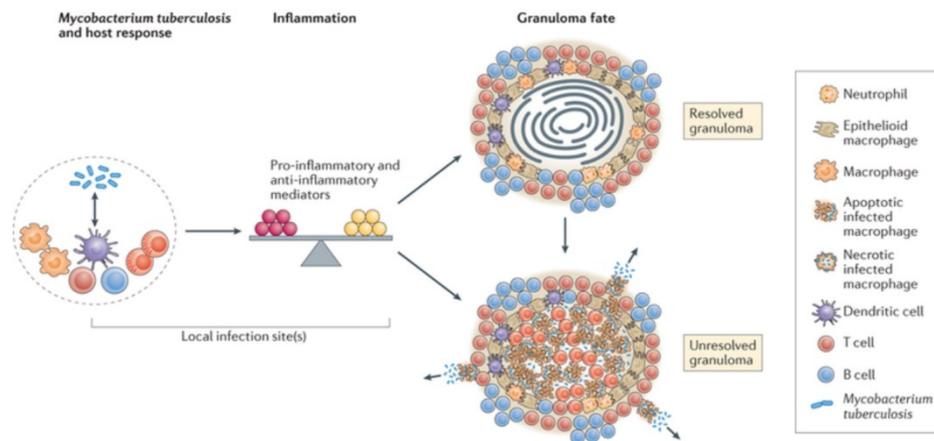


Fig. 5: Granuloma fate is influenced by a complex and dynamic exchange of host and bacterial features. This process is highly dynamic and iterative, with multiple components having pleiotropic, knock-on feedback effects on inflammation and the host-pathogen interaction ²⁴.

Cytokines are crucial for immune system activation and TB resolution, but further studies are necessary to completely understand the granuloma formation mechanism and the role of all immune system participant cells (Table 3). $TNF\alpha$ among others plays an important role in granuloma formation and contributes to the recruitment of myeloid cells. It is mainly produced by macrophages, and studies demonstrated that anti-TNF treatment reduces the number of macrophages recruited and thus the granuloma size, impairing the bacilli containment ²⁵. The balance between IL-12 and $IFN\gamma$ (Interferon γ) is important for the fate of the illness and for the activation of macrophagic killing functions. $IFN\gamma$ is mainly produced by $CD4^+$, $CD8^+$, NK when stimulated by dendritic cell's IL-12 and is essential for an efficient establishment of the granuloma, and for controlling bacterial

growth²⁶. Human population studies have correlated IL-12 production gene mutations to an improved predisposition to TB infection, and to show a severe form of the disease²⁷. During MTB infection the role of Type I IFN can be protective or detrimental for the host. Type I IFN activation during chronic phases of TB causes immunosuppression, inducing IL-10 production and favors bacteria survival²⁸.

The IL-1 cytokine family is composed of IL-1 α , IL-1 β , IL-18, and IL-33. IL-1 β production after MTB infection is a process that is tightly regulated. Mice model experiments demonstrated the importance of IL-1 signaling in controlling the infection. IL-1 α or β knock-out mice show a severe form of TB compared to the control²⁹.

Macrophage cytokines are crucial for the recruitment of neutrophils and adaptive immunity activation after MTB infection. Among others, macrophages produce pro-inflammatory cytokines such as type I IFN, TNF α , IL-1 β , IL-2, IL-18, IL-6, and anti-inflammatory cytokines IL-10 and Transforming Growth Factor (TGF)- β . They produce and secrete a huge range of chemokines for cells recruitment (CC, CXC, CX₃C) and growth factors as M-CSF (Macrophage colony-stimulating factor), GM-CSF (Granulocyte-Macrophage Colony-Stimulating Factor), and Erythropoietin³⁰. TNF α membrane expression through ADAM17 complex is essential for granuloma formation and is crucial for TB eradication while the secretion of soluble TNF α is important for catabolism with IL-6^{31,32}.

Table 3: Principal cytokines and chemokines and their role in TB ³³.

Cytokine	Receptor/Signal	Role in TB
TNF α	TNFR1, TNFR2 JNK, p38, NF κ B	Positive: Essential for survival following Mtb infection. Initiation of innate cytokine and chemokine response and phagocyte activation Negative: Mediator of tissue damage
IFN γ	IFNGR1, IFNGR2 JAK/STAT	Positive: Essential for survival following Mtb infection. Coordinates and maintains mononuclear inflammation. Expressed by antigen specific T cells Negative: Potentially pathogenic
IFN α /IFN β	IFNAR1, IFNAR2 JAK, TYK, ISG, ISRE	Positive: Required for initial recruitment of phagocytes to the lung Negative: Over expression of IFN α /IFN β results in recruitment of permissive phagocytes and regulation of T cell accumulation and function
IL-6	IL-6R, gp130 JAK, STAT3, MAPK	Positive: Potentiates early immunity – non essential unless a high dose infection.
IL-1 α /IL-1 β	IL-1R1, IL1RAcP MyD88, IRAK4, NF κ B	Positive: Essential for survival following Mtb infection. Induction of IL-17. Promotes PGE2 to limit Type I IFN
IL-18	IL-18Ra, IL-18R β MyD88, IRAK, NF κ B	Positive: May augment IFN γ – non-essential. Regulator of neutrophil/monocyte accumulation. of neutrophil and monocyte accumulation, optimal induction of IFN γ by T-cells
IL-12 p40, p35	IL-12R β 1, IL-12R β 2 JAK2, TYK2, STAT4	Positive: IL-12p40 and IL-12p35 essential for survival following Mtb infection. Mediate early T cell activation, polarization and survival. Negative: Over expression of IL-12p70 is toxic during Mtb infection.
IL-23 p40, p19	IL-23R, IL-12R β 1 JAK2, TYK2, STAT3	Positive: Required for IL-17 and IL-22 expression during Mtb infection. Non-essential in low dose challenge required for long term control. Negative: Mediates increased pathology during chronic challenge
IL-27 EB13, p28	IL-27Ra, gp130 JAK1/2, TYK2, STAT1/3	Positive: May control inflammation and reduce pathology Negative: Regulates protective immunity to Mtb infection by limiting the migration and survival of T cells at the inflamed site.
IL-35 p35, EB13	IL-12R β 2, gp130 STAT1/4	Positive? Regulate the availability of subunits of IL-12, IL-27 Negative? Potential immunoregulatory role.
IL-17A/F	IL-17RC, IL-17RA	Positive: Essential for survival following infection with some strains of Mtb. Induction and maintenance of chemokine gradients for T cell migration. Negative: Drives pathology via S100A8/A9 and neutrophils
IL-22	IL-22R1, IL-10R2 TYK2, JAK1, STAT3	Positive: Induces antimicrobial peptides and promotes epithelial repair, inhibits intracellular growth of Mtb in macrophages.

Chemokine	Receptor	Role in TB
CCL-3,-4,-5	CCR1	Positive: Upregulated during infection. Non-essential in mouse model
CCL-2,-7,-12	CCR2	Positive: Maximizes and organizes early macrophage and T cell accumulation in the lung Negative: Mediates recruitment of permissive phagocyte accumulation into the lung
CCL-17,-22	CCR4	Positive: Mediates optimal granuloma formation to mycobacterial antigen Negative? May limit T cell proliferation via T _{REG}
CCL-3,-4,-5	CCR5	Positive: Regulation of pulmonary infiltrates – non-essential. May mediate early dendritic cell accumulation in the lymph node. May augment macrophage Mtb killing via CCL5?
CCL-20	CCR6	Positive: Expression of CCR6 on T cells specific for Mtb antigens Negative? CCL-20 seen at high levels in active TB
CCL-19,-21,	CCR7	Positive: Mediates efficient migration of dendritic cells and Mtb-specific T cell activation.
CXCL-1,-2,-3,-5,-6,-7,-8	CXCR1 CXCR2	Positive: Expressed on neutrophils mediates accumulation Negative: Absence of CXCR2 or CXCL5 results in improved bacterial control and reduced neutrophil accumulation
CXCL-9,-10,-11	CXCR3	Positive: Required for optimal granuloma formation. Expressed on Mtb-specific T cells. Use of CXCL9-11 levels to indicate disease level? Required for early recruitment of T cells to lung
CXCL-13	CXCR5	Positive: Required for correct location of T cells within granulomas. Required for B cell follicle formation in Mtb infected lungs. Required for optimal protection.

1.4 MTB genomic diversity and pathogenicity

The genetic variability of MTB clinical isolates went neglected for decades; however, in the past 10-15 years the scientific community recognized it causes important consequences on the progression of infection. Even though the genomic diversity in MTBC is low compared with other bacteria, at least 44% of non-synonymous SNPs have been predicted to impact gene function, thus altering bacterial phenotype ³⁴. Drug stress and immune system response cause a selective pressure on the bacteria, which push the fixation of these mutations ^{35, 36}. Studies based on WGS techniques have recognized 7 main lineages of MTBC that can be classified in clades according to their proximity to their common phylogenetic ancestor. The first (monophyletic) clade, the so-called "modern", includes MTB lineages 2, 3, and 4 (which includes also the laboratory reference strain H37Rv). Clade 2, the "ancient", includes MTB lineages 1, 7, and *M. africanum* lineages 5, 6 and is paraphyletic not being part of the same phylogenetic group (Fig. 6) ³⁷. More recently, new strains have been identified and classified as a new Lineage 8 (Table 4).

Table 4: MTB lineage classification and spoligotype associated ¹¹

Evolutionary age (species)	Lineage name based on LSP/SNP ^a		Lineage and sublineage [RD associated]			Spoligotype family	
Ancient lineage (<i>M. tuberculosis</i>)	Indo-Oceanic lineage	1 [RD239]	11	1.1		EAI4 and EAI5	
					1.1.1		EAI4
					1.1.2	1.1.1.1	EAI5 and EAI3
					1.1.3		EAI6
					1.2		EAI2
					1.2.1		EAI1
					1.2.2		MANU ancestor and orphan profile
					2.1 (non-Beijing)	2.2.1 [RD181]	Beijing
					2.2 (Beijing) [RD105, RD207]	2.2.1 [RD150] 2.2.1.2 [RD142]	Beijing
					2.2.2		Beijing
Modern lineages (<i>M. tuberculosis</i>)	East-Asian lineage	2	2.1 (non-Beijing)			CAS except CAS1-Delhi	
						CAS1-Kili	
						CAS2	
						CAS	
						X2	
						X1	
						X3 and X1	
						T1 and H1	
						T1 and H1	
						H3 and H4	
	East African-Indian lineage	3 [RD750]	3.1			LAM7-TUR and T1	
						LAM7-TUR	
						LAM9	
						LAM3	
						LAM3	
						LAM9 and T5	
						LAM1	
						LAM11-ZWE, LAM9, LAM1, and LAM4	
						LAM11-ZWE	
						S	
Euro-American lineage	4	4.1	4.1.1 (X-type)	4.1.1.1 [RD183] 4.1.1.2 4.1.1.3 [RD193]	T1		
				4.1.2	4.1.2.1 (Haarlem) [RD182]	T1 and T2	
				4.2	4.2.1 (Ural) 4.2.2	H3, H4, and T1	
					4.2.2.1 (TUR) [RD182]	T2-Uganda	
				4.3 (LAM)	4.3.1 4.3.2	T2	
					4.3.2.1 [RD761]	T3	
					4.3.3 [RD115] 4.3.4 [RD174]	LAM10-CAM	
					4.3.4.1 4.3.4.2	T1 and T5	
				4.4	4.4.1 4.4.2	T1, T2, T3, T4 and T5	
				4.5 [RD122] 4.6	4.4.1.1 (S-type) 4.4.1.2	T1	
Ancient lineages (<i>M. africanum</i>)	West-Africa lineage 1	5 [RD711]	4.6.1 (Uganda) [RD724] 4.6.2 [RD726]	4.6.1.1			
				4.6.1.2			
				4.6.2.1			
				4.6.2.2 (Cameroon)			
				4.7			
				4.8 [RD219]			
				4.9 (H37Rv-like)			
				AFRL_2 and AFRL_3			
				AFRL_1			
Intermediary lineage (<i>M. tuberculosis</i>)	West-Africa lineage 2	6 [RD702]	7				

^aRegions of deletion (RD) are given in brackets and appear below the lineage/sublineage in which they are present.
^bLSP, large sequence polymorphism.

Phenotypic differences found among MTB lineages can be seen both in planktonic growth, gene expression, metabolism, and in *in-vivo/in-vitro* macrophagic infection, including in virulence, bacterial-host cell uptake, cytokine induction, and intracellular growth (Fig 6) ^{38, 39}. Studies also demonstrated that different MTB lineages have different probability rates to develop drug resistance ^{40, 41}.

Despite the progress made in our understanding of the global phylogenetic diversity of MTBC, much remains unknown with respect to both human- and animal-associated MTBC diversity ¹. Despite the number of studies concerning the origin of the MTBC and the proposed African origin, there are little data that explains why modern MTBC lineages are globally

dispersed ⁴². There is evidence of a correlation between virulence, transmission, survival to stress and lethality, and the different MTB lineages however, the molecular mechanism that controls this is still poorly understood ⁴³.

Ongoing studies are shedding light on the host-pathogen interactions and demonstrated that the host immune response is differentially induced by different MTB strains ^{44, 45}. In fact, ancient lineage strains induce a higher macrophagic response compared to the phylogenetic modern ones ⁴⁵. Modern lineages delay the immune activation reducing the early inflammatory response, thus providing the bacteria a selective advantage during the early phases of the infection ⁴⁴. A reduced host response (associated with high virulence) is supposed to be the key to MTB persistence and transmissibility. Indeed, it has been demonstrated that L2 and L4 lineages are responsible for most TB cases in the world ³⁷.

MTB lineages generally display a strong phylogeographical host adaptation, with lineages associated with geographical regions ⁴⁶. Ecologists distinguish between generalists and specialists depending on the size of a species' ecological niche. In infectious diseases, the niche of a pathogen is determined by host availability and the bacteria's capacity to survive (Fig. 6).

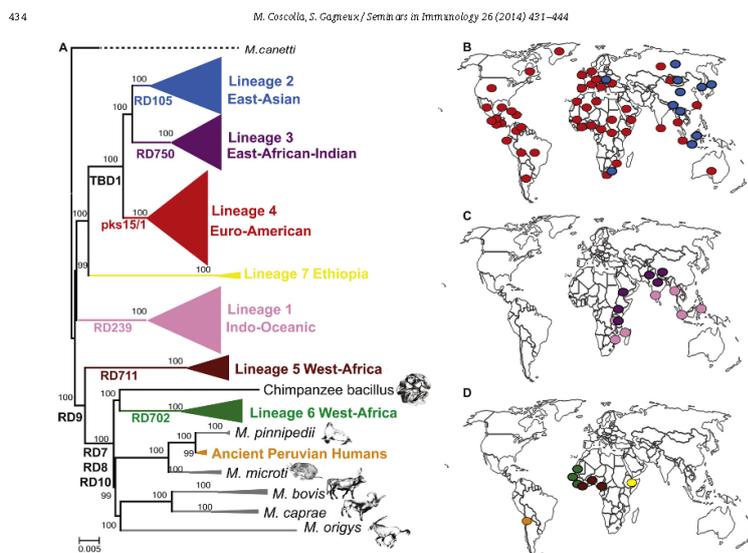


Fig. 6: World diffusion of MTBC lineages ³⁷

Lineage 1 (L1) is predominant in regions of the Indian Ocean, extending from Eastern Africa to Melanesia. Lineage 2 (L2) is mostly distributed, with a predominance in Eastern Eurasia and South East Asia. L3 is similarly distributed to L1 and is frequent among isolates reported from the Indian Ocean and in Northern Africa and across Southern Asia. L4 is well dispersed, with a predominance throughout Africa and Europe and the entire region of the Mediterranean. L5 and L6 are reported at low frequencies in Western and Northern Africa. L7 is limited to Ethiopia. Remarkably, L1–L4 are also reported in Northern Europe, where the majority of TB cases are due to recent immigration ⁴⁷.

M. africanum (L5, L6) was described for the first time in 1968 and it appears very similar to MTB *sensu stricto* ⁴⁸. However, *M. africanum* has been associated with a longer latency period and a lower virulence as an adaptation to low human population densities. *M. africanum* is almost exclusively found in Africa and represents the etiological agent of almost 40% of TB cases in some West African countries, suggesting a correlation between the pathogen and the host in selected regions ^{1, 49, 50, 51}. Host-adaptation has been associated, among other hypotheses, with the high-level concentration of vitamin B12 in African population plasma. Vitamin B12 plays an important role in the pathogenesis of TB and mutations in the cobalamin biosynthesis pathway in *M. africanum* could be the reason why *M. africanum* is difficult to find infecting non-African populations ⁵². *M. africanum* drug resistance and pathogenesis features are not clear. Its lower frequency of the insurgence of drug resistance, and its reduced virulence are nowadays under debate. At a clinical level, *M. africanum* has been associated with a slower progression to disease compared to MTB (as outlined for other ancient lineages), and it is more commonly found in people living with HIV and the elderly population. Compared to MTB infections, the clinical isolates of *M. africanum* were associated with lower bacterial burdens, lower signs of disease, and tissue pathology, even in hosts lacking IFN γ production ⁵³. Furthermore, several MTB's key virulence mechanisms have been shown to be affected in *M. africanum*. For

example, the PhoPR two-component system, essential for MTB virulence, for mycolic acid production, and for the secretion of ESAT-6 (Early Secretory Antigenic Target) antigen, shows a SNP (position 71 of PhoR protein) in ancient lineages (*M. africanum*) 5 and 6, and in animal-adapted strains, thus affecting PhoPR regulatory pathway⁵⁴. Of note, a single amino acid change in PhoP protein (position 219) in H37Ra highly attenuates H37Ra strain virulence⁵⁵. The control of *M. africanum* in mice lungs reflects the low immune cells recruitment and low induction of cytokines with low levels of IFN γ , IL-17, and TNF α . DosR expression, the regulon involved in the controls of the adaptation to oxygen limitation and linked to MTB virulence, is significantly lower in *M. africanum* compared to modern strains⁵⁶.

Differences are not observed only when comparing relatively (phylogenetically) distant ancient and modern lineages. Indeed, even between the monophyletic modern clade differences have been observed. Lineage 3, when compared to lineage 4, shows a higher anti-inflammatory phenotype, and Beijing strains (L2) commonly induce low levels of TNF α , IL-6, IL-10, and GRO- α compared to the laboratory strain H37Rv (which belongs to L4)⁵⁷. Moreover, L2 strains show a lower uptake and lower cytokine induction when compared to L4^{58, 59}. L2 has been also associated with treatment failure and disease relapse, with high HIV co-infection and it is also found at higher rates in TB meningitis compared with other lineages^{60, 61}. The “hypervirulence” of Lineage 2 seems to be related to phenolic glycolipids (PGL) production, which can modulate the immune system by interfering with the effector molecules of the host's innate immune response: in H37Rv the lack of PGL production is due to a deletion of 7pb in the *pks 15/1* gene which disables the synthesis of these lipids by increasing the immune response. A possible explanation for this correlation could be the presence of mutations in the *mut* gene, which is involved in repairing DNA damage. It is hypothesized that the sequential accumulation of mutations in this gene could cause an increase in the mutation frequency rate in Beijing strains which would confer better adaptability to

stress and the ability to acquire drug resistance, but this hypothesis finds contrasting evidence in the literature ⁶². Furthermore, the higher virulence rate is also caused by its ability to grow rapidly in human monocytes and to slow-down host immune response ⁶³ (Fig. 7).

It's also important to underline that there are several important differences between clinical strains and laboratory MTB strains. The canonical most used MTB strains in the laboratory are the so-called H37Rv, H37Ra, Erdman, BCG vaccine strain, CDC1551, and HN878 ⁶⁴. These are all modern strains, belonging to lineage 4 (H37Rv, H37Ra, CDC, Erdman) or lineage 2 (HN878). The principal limitation in working with laboratory strain is that the bacilli rapidly adapt to laboratory conditions and lead to genetic modification (such as the loss of PDIM wall component) and thus to potential artifacts. Moreover, even if synonymous mutations are supposed not to affect phenotype and protein activities, a synonymous SNP was found to alter the transcription start site (TSS) position of the *dosR* gene in Lineage 2, generating an increase in its expression ³⁹. *dosR* gene duplication events have also been observed in L2 strains, causing its upregulation ⁶⁵. A completely neglected topic concerns the effect of synonymous mutations on the post-transcriptional regulatory network linked to antisense transcripts, but pioneer studies in *M. bovis* already highlighted implications for how we define so-called "silent" mutations ⁶⁶.

Deletion events also influence bacteria phenotype. A deletion in the *pks* 15/1 gene in Lineage 4, reduces the phenolic glycolipid production decreasing host immune cell responses ⁶⁷. In Lineage 3, deletion in the *Rv1519* gene (annotated as conserved hypothetical protein) causes reduced production of IL-10 by the host ^{57,68}.

It is difficult to generalize and predict what will be the phenotype of strains, knowing only their lineage. Intra-lineage and sub-lineage differences are present among both modern and ancient strains. Even though phenotypic diversity (and immune specific response) has been associated with the different MTBC lineages, individual strains within these lineages exhibit a huge spectrum of phenotypes, suggesting that sub-

lineage should also be considered when studying host-pathogen interactions ^{44, 69}. Differences are present also when considering the inflammatory response to “modern” Beijing strains and “ancestral” Beijing strains, with a lower inflammatory response of the former ⁷⁰.

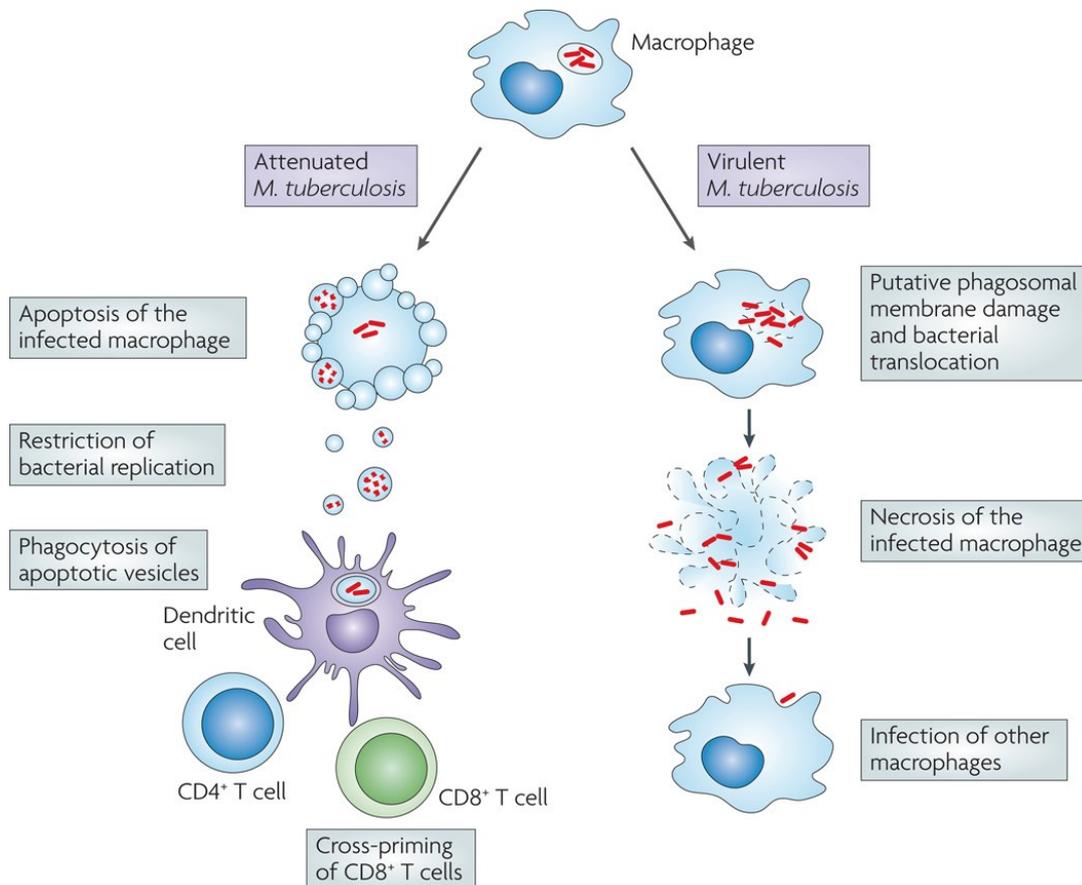


Fig. 7: Mycobacterium TB infects macrophages, survives, and replicates in the phagosome. Macrophages infected with attenuated strains of MTB undergo apoptosis. Apoptosis of infected macrophages provides an important link to adaptive immunity, as apoptotic vesicles containing bacterial antigens are taken up by dendritic cells, which presents antigens to naive T cells. By contrast, virulent MTB inhibits apoptosis and, instead, induces necrosis ⁷¹.

Studies on L4, investigating the global success of those strains, discovered that it is mainly a consequence of both biological and social phenomena. In a recent study on L4 strains, Stucki et al. discovered the presence of at

least 10 L4 sub-lineages, that can be further divided into specialists and generalists, with some sub-lineages showing an intermediate geographical distribution ⁶. In the sub-lineage analyzed in this study, specific T cell epitopes are conserved, but nonsynonymous variation may allow antigens to be differently and selectively recognized by T cell receptors of different human populations ⁶. Similarly, in 2021 new evidence allowed to define sub-lineages also in the ancient L5 emphasizing how much this field is evolving with the advent of WGS approaches ⁷².

It's important to remark that transmission patterns are complicated mechanisms that are not only linked to the intrinsic virulence of individual organisms (Lineages) but depend on several factors, most of them poorly understood, including the role and the variability of the host. For example, as shown in the study of Shanley et al., despite similar CFU levels seen in the selected MTB isolates, there appeared to be differences in lung pathology in infected guinea pigs ⁷³.

1.5 Macrophage phenotypic diversity and mycobacterial infection

Monocytes are important both for the activation of the inflammatory response and for the priming of innate immunity. The monocytes, once generated in the bone marrow, migrate to different tissues where they differentiate into specialized cells following exposure to various microenvironmental factors. Also, macrophages are highly adaptive cells, and their phenotype can change in a plastic way in responding to external stimuli, such as inflammation or pathogen invasion ¹².

The activation status of macrophages is called polarization. MØ (naïve, non-polarized) macrophages are activated by the secreted cytokines. According to the signal received, MØ macrophages are driven into a so-called classical M1 pro-inflammatory phenotype, or to a so-called non-classical M2 anti-inflammatory phenotype. The main difference between these cells is that in M2 macrophages the metabolism of arginine is shifted

to ornithine and polyamine, while in M1 cells it is shifted to NO and citrulline. Ornithine produced by M2 can promote cell proliferation and repair through the synthesis of collagen, fibrosis, and other tissue remodeling functions, while the NO produced by class M1 is an important molecular effector with microbicidal activity and inhibition of cell proliferation⁷⁴. The polarization states of macrophages have a huge impact on their function. It affects how macrophages can react to external signals, changing gene expression, membrane composition, receptor exposure, and cytokines production¹² (Fig. 8).

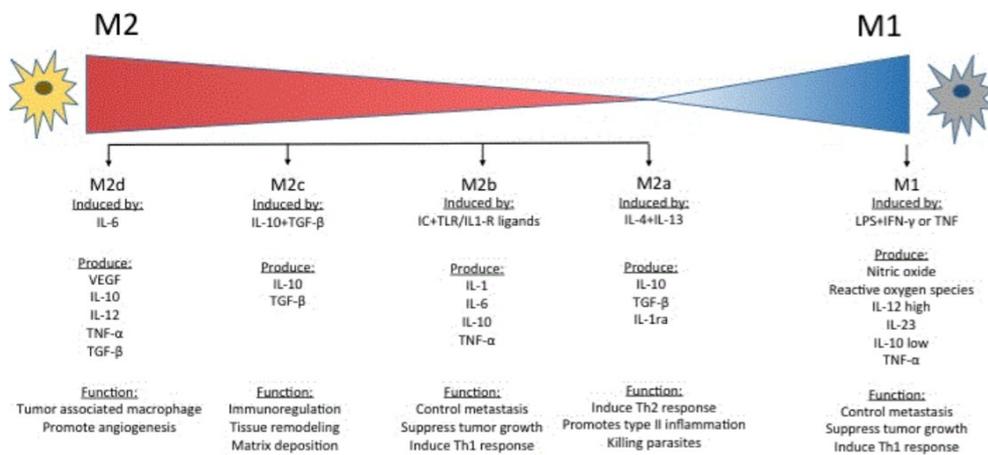


Fig. 8: Polarization status of macrophages and principal cytokine that induce the differentiation. The functions of different types of macrophages are highly dependent on the polarization status⁷⁴.

Th₁ cells signals are the main triggers for the M1 polarization, which enhances the macrophagic antigen presentation, the phagocytosis, the killing capacity, and the capacity to attract other leukocytes⁷⁵. On the other side, molecules produced by Th₂ cells are the main drivers for the M2 polarization, thus decreasing phagocytosis, inducing macrophagic fusion, increasing the anti-inflammatory activity and promoting the mannose receptor (CD206) synthesis⁷⁶.

In the lung, macrophages constitute a heterogeneous population distributed within two compartments: alveolar macrophages (AM) and interstitial macrophages. The former is mostly represented in the airways (bronchi and alveoli), the latter predominates in the parenchymal tissue. When MTB enters the lungs, AMs are the first line of defense that recruit bloodstream inactive M \emptyset (naïve, non-polarized) macrophages into the alveoli after infection. In this space, recruited M \emptyset macrophages can be polarized toward an M1 or M2 phenotype, depending upon the cytokines at the microenvironment. M2 macrophages are essential to prevent immunopathology, but they are more permissive to MTB infection, whereas M1 macrophages are able to control mycobacterial proliferation, but trigger an increased inflammatory response, increased lung damage, and high IFN γ production ⁷⁷. M2 permissiveness is mainly due to the high amount of CD206 that these cells have compared to M1 macrophages. MTB interaction with mannose receptors causes the activation of the phagocytic process and the induction of the production of anti-inflammatory cytokines, the reduction of the oxidative response, and the phagosome maturation ⁷⁸. Recent studies showed how M2 permissiveness is also linked to the different metabolic pathways activated. Indeed, M2 metabolism is up-regulated for fatty acid uptake and β -oxidation; intracellular MTB is known to use fatty acids and cholesterol from the host, suggesting that those bacteria in M2 macrophages possess a metabolic advantage (Fig. 9). On the contrary, MTB-infected M1 macrophages show a metabolism with high levels of glycolysis. In addition, M1 activated macrophages are able to produce high levels of ROS, NOS, and NADPH oxidases when compared to non-activated macrophages ⁷⁹. These molecules, besides their killing activity on bacteria, are involved in the enhancement of inflammatory activation and tissue damage. MTB has different enzymes involved in oxidative stress escaping such as the superoxide dismutase and KatG protein ⁸⁰.

Despite MTB macrophage infection have been reported to promote an M1 phenotype with high IFN γ production, the control of the first phases of the

infection is temporary and lasts a maximum of three weeks. Indeed, MTB is able to shift the initial M1 macrophagic polarization status to the more permissive anti-inflammatory M2 phenotype^{81, 82}. During this time, MTB produces exosomes rich in MTB lipids that increase the inflammatory response and recruit more neutrophils, dendritic cells, and additional macrophages, that will be also infected by MTB bacilli. MTB ManLAM and secreted ESAT-6 inhibit the interferon T cell pathways, and induce the production of IL-10, allowing the recruiting of anti-inflammatory, more permissive cells, the reduction of NO production, and increasing iron accessibility⁸³. This M1/M2 shift occurs in the granuloma when the acute infection progresses into chronic. Recent studies generated a predictive model of TB granuloma progression in time that was dependent on the M1 versus M2 ratio at the site of infection, thus providing further evidence of how critical the balance is between M1 and M2 phenotypes to the containment of the bacteria inside the granuloma, with a delay in M1 polarization that appears to be responsible for increased survival of the bacteria. M1 cells are predominant at early phases of the infection (2 to 4 months post-infection) while at later timepoints they became not protective and may be deleterious due to an excessive inflammation and tissue damage⁸⁴.

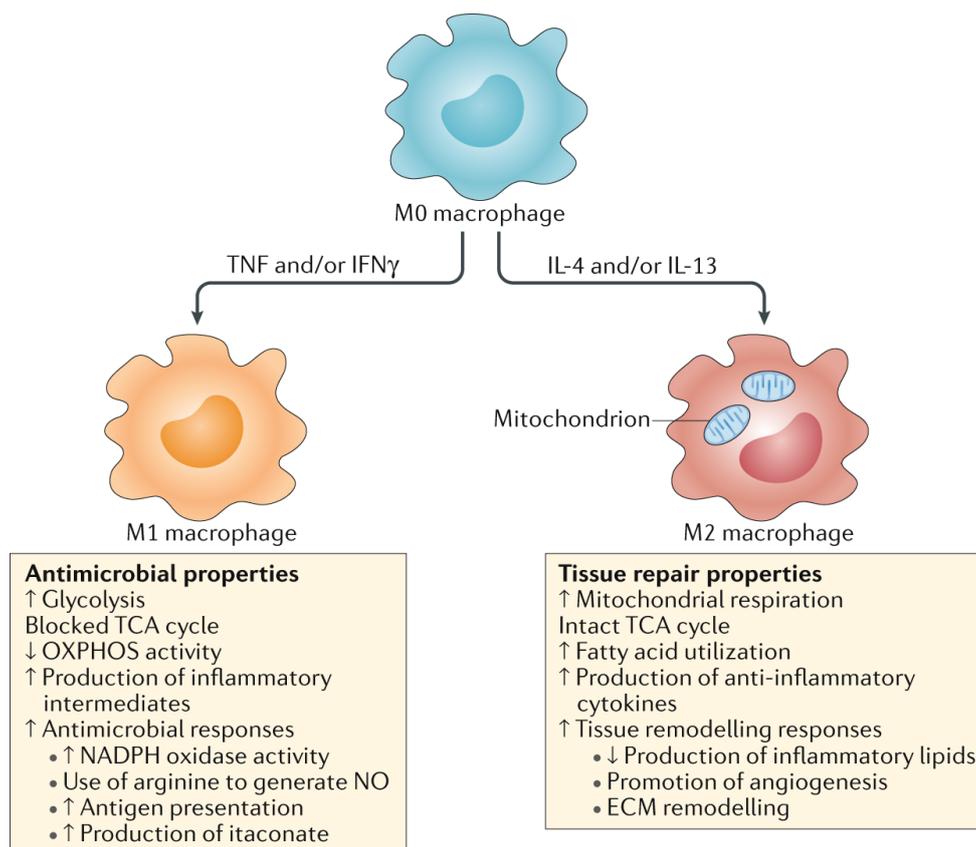


Fig. 9: Phenotypic characteristics of the M1-like and M2-like macrophage polarization states.

The M1-like and M2-like macrophage polarization states highlight the pro-inflammatory or anti-inflammatory phenotype of these cells by oversimplifying the extremely heterogeneous nature of macrophages in vivo. M0 macrophages are activated to an M1-like macrophage by exposure to IFN γ and/or tumor necrosis factor (TNF), or LPS. M2-like macrophages are induced through IL-4 and/or IL-13 exposure. M1-like macrophages are more glycolytically active and more prone to kill bacteria. By contrast, M2-like macrophages have increased mitochondrial respiration and enhanced fatty acid metabolism. ECM, extracellular matrix; NO, nitric oxide; OXPHOS, oxidative phosphorylation; TCA, tricarboxylic acid cycle ⁸⁵.

Macrophage polarization status should not be considered an on-off mechanism between pro- and anti-inflammatory phenotypes, but it often shows a spectrum of intermediate states even within the same cell

population¹³². Furthermore, it is not even fixed in time, because the initial polarization status is plastic and tends to shift from a state to another depending upon multiple external stimuli⁸⁶. As a consequence of this operational definition, M1 and M2 phenotypes do not reflect the huge spectrum of the macrophagic population. This seems especially appropriate for the macrophagic populations found in the lungs where AMs have been reported to present a hybrid pattern expressing both M1 and M2_a markers. AM are both crucial for first contact with pathogens and for maintaining homeostasis in the lungs. Given their double role, their activation is tightly regulated and balances both pro- and anti-inflammatory functions. Transcriptomic analysis showed that they express M1 typical proteins such as CD80, CD64, CD86, CD69, TLR2 and 4, CXC-chemokine ligand (CXCL) 9,10 and 11, and CC-chemokine ligand 5 (CCL5), but also typical M2 markers (CD163, CD206, MARCO, matrix metalloproteinase (MMP) 2, 7 and 9, the tyrosine-protein kinase MER, growth arrest-specific proteins, CD163, stabilin, arginase, and the adenosine A3 receptor)^{130, 131}. In healthy lungs, this phenotype remains stable even after exposure to environmental stimuli known to alter the alveolar environment, such as the pneumococcal colonization (Fig. 10). AM Pattern Recognition Receptors (PRRs) recognize the Pathogen-Associated

Molecular Patterns (PAMPs) and initiate the inflammatory process producing $\text{TNF}\alpha$, $\text{IL-1}\beta$, and $\text{IFN}\gamma$ ⁸⁸. At the same time, IL-10 and $\text{TGF}\beta$ and CD206 , act as anti-inflammatory signals, reducing the inflammation status of alveoli. The AM peculiar low capacity to act as antigen-presenting cells (APC), to produce low levels of oxidation agents such as ROI and NO, and makes them particularly favorable for MTB dissemination^{89, 90}. The presence of CD206 receptors on AMs further facilitates MTB cell enter and the scavenging of the immune response. Thus, the fate of MTB infection strongly depends on how MTB strains interact with AMs and how the resulting macrophagic signals activate or inhibit the recruitment of other immunity participants.

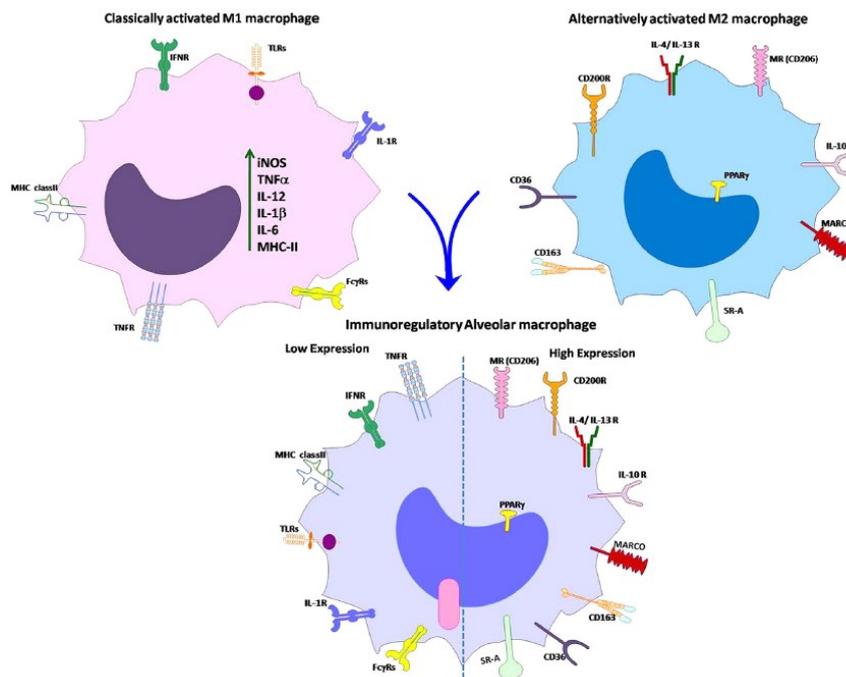


Fig. 10: Macrophage phenotypes and external stimuli that drive macrophagic differentiation. The figure reports the mainly expressed receptors found on those macrophage subtypes. AM expresses both M1 and M2 markers⁹¹.

1.6 MTB/macrophage interaction

The first interaction between MTB and macrophages mostly occurs when the PRRs of the host cell encounters the recognition of PAMPs on the bacteria's surface. This recognition is driven by a pattern of receptors such as Toll-Like Receptors (TLR), C-type Lectin Receptors, Fc Receptors (FcR), Scavenger Receptors (SR) ⁹. Activated PRRs give to the host cells a signal that leads to bacterial phagocytosis, and to the priming of cellular signals such as apoptosis, antigen processing-presentation, inflammasome activation, phagosome maturation, and autophagy ⁹². Among other important macrophagic surface proteins, involved in the bacterial recognition/phagocytosis, CLR/CTL (membrane-bound calcium-dependent receptors) play a crucial role: the well-known CD206 recognizes the MTB mannose molecules present in LAM and ManLAM surface glycolipids. Under normal conditions, bacteria are phagocytized by the phagosome, which is then fused with a lysosome to form the phagolysosome. This phagolysosome is acidified to low pH (5.2) and enzymes are activated to degrade bacteria.

When phagocytized into the macrophages, MTB bacilli are exposed to an array of stresses, including oxidative and nitrosative stresses, iron restriction, starvation, and low pH; MTB has evolved several strategies to survive within the macrophages and cope with such unfavorable conditions. Indeed, MTB is able to hijack the macrophagic response and to survive within it by acting at different levels (Table 5).

Table 5: Principal MTB escape mechanisms ⁹³

	MTB defence mechanisms	Definition
In the lungs	Recruitment of permissive monocytes	Virulent factor PDIM has been associated with the recruitment of low antimicrobial activity monocytes.
	Phagocytosis through specific receptors	The particular phagocytic receptors MTB engages may influence the ability of macrophage to control the infection (e.g. MR interaction with ManLAM, can impair phagosome maturation)
In the phagosome	Modulation of phagosome maturation	Prolonged retention of early endosomal markers and blocking of phagosomal maturation (e.g. PIM and ManLAM can impair the acquisition of PI3P and alter RAB protein dynamics.
	Permeabilization of the phagosome	EsxA and PDIM are among the virulence factor involved in the phagosomal escape mechanisms
	Phagosome perforation	ESX-1 secretes different factors, including EsxA EsxB, involved in the rupture of the phagosome.
	Phagocytosis through specific receptors	The particular phagocytic receptors MTB engages may influence the ability of macrophage to control the infection (e.g. MR interaction with ManLAM, can impair phagosome maturation)
	Blocking of phagosome maturation	Interference with RAB, PI3K and the vacuolar H(+)-ATPase. Promotion of phagosome-early endosome fusion. Expression of ROS detoxifying enzymes (e.g. SodA). Blocking of acidification
In the cytosol	Autophagy evasion	MTB infection reduce the LC3 recruitment. MTB impairs both autophagy initiation and the fusion of autophagosomes with lysosomes at Rab7 level.
Other defence mechanisms	Manipulation of the inflammasome	Triggering of cGAS-STING pathways deactivates the AIM2 inflammasome pathway reducing the production of pro-inflammatory cytokines.
	Apoptosis impairing	MTB reduces the efferocytosis process, increasing the necrosis levels and impairing apoptosis, by increasing the transcription of MTB anti-apoptotic genes (e.g. PknE, SigH, MPT64 and Rv3654c), thus limiting host ROS and apoptosis triggers factors

The ability of MTB to block the phagosome maturation and the consequent acidification of the compartment, together with its capacity to survive in low pH environments have been reported to be among the main successful strategies to avoid bacilli eradication. During normal phagocytosis, cell membrane movements phagocyte external debris of bacteria into phagosomes. Macrophages normally recruit RAB5 in early phagosomes, which is replaced with RAB7 in an event called Rab conversion ⁹⁴ (Fig. 11). At this level, Rab GTPases expressed on the phagosomal surface, attract the v-ATPases that are responsible for a first acidification of this compartment (pH 6). Then the early phagosomes fuse with lysosomes to further lower the internal pH (pH <5,5). This event is altered by MTB infection mainly by the activity of two MTB proteins (Kdk and PtpA) that interfere with its maturation by preventing the phagolysosome fusion event and its consequent acidification ^{95, 97}.

Moreover, the ability to interfere with the phagosome maturation of the MTB ManLAM has been associated with its capacity to block the Phosphatidylinositol 3-phosphate (PI3K)-calmodulin complex, and the further attraction of v-GTPases and V-ATPases, thus stabilizing phagosome internal pH around 6.2. In addition, the interaction between MTB and CD206 causes the induction of anti-inflammatory cytokines and the reduction of the oxidative response ⁹⁶.

When the phagolysosomal fusion occurs, MTB can survive in low pH environments thanks to its thick, low permeable membrane and securing enzymes, such as proteases MarP and RipA that allow the production of basic components, thus causing the neutralization of phagolysosome pH.

Escaping from phagosomes to the cytosol is the key mechanism that allow MTB to replicate in macrophages. The MTB genome encodes for 5 different ESX systems (ESX-1-ESX-5), a type VII secretion system that allows MTB to secrete soluble virulent factors ⁹⁸. *esx-1* is probably the most studied system and it encodes ESAT-6 (EsxA) and the 10 kDa Culture Filtrate Protein (CFP-10; EsxB), considered key virulence determinants of MTB and strong antigens for T-cells ⁹⁹. It has been shown that the ESX-1 secretion system together with the MTB membrane component PDIM enables phagolysosomal membrane disruption in macrophages, thus allowing MTB to reach the cytosol ²³⁻¹⁰⁰. Studies on BCG strains (which do not possess the ESX-1 complex) have confirmed the presence of only virulent MTB strains in the cytosol after 4 days from the initial infection in macrophages, as also confirmed by recent FRET-based experiments ¹⁰¹.

Once MTB is in the cytosol, host cells activate cGAS systems to sense bacterial DNA and activate STING signals and Type I IFN production ¹⁰². This leads to the expression of type I IFNs (α and β) and the subsequent release of IL-1 β and IL-18 after the activation of the inflammasome response ¹⁰³. The cytosolic shift of MTB also activates TBK-1, a DNA-responsive kinase, which is involved in the recruitment of LC3-II and the subsequent activation of the autophagic flux. During autophagy, double-membrane autophagosomes phagocyte cytosolic pathogens (xenophagy)

and deliver them to fuse with acidic lysosomes, resulting in the release of lysosomal hydrolases to digest the enclosed contents. During MTB infection, IFN γ increased production, activates macrophages, and induces autophagy for bacterial degradation¹⁸⁹. MTB has evolved several mechanisms to inhibit the autophagic response. H37Rv has been reported to induce IL-6 production, which selectively inhibits IFN γ induced autophagosome formation¹⁰⁴. In addition, MTB *eis* gene modulates autophagy in macrophages, by increasing the acetylation of the H3 histones of the IL-10 gene promoter region, thus increasing its expression. Furthermore, MTB inhibits RAB-7, an endosomal marker involved in autophagosome formation¹⁰⁵.

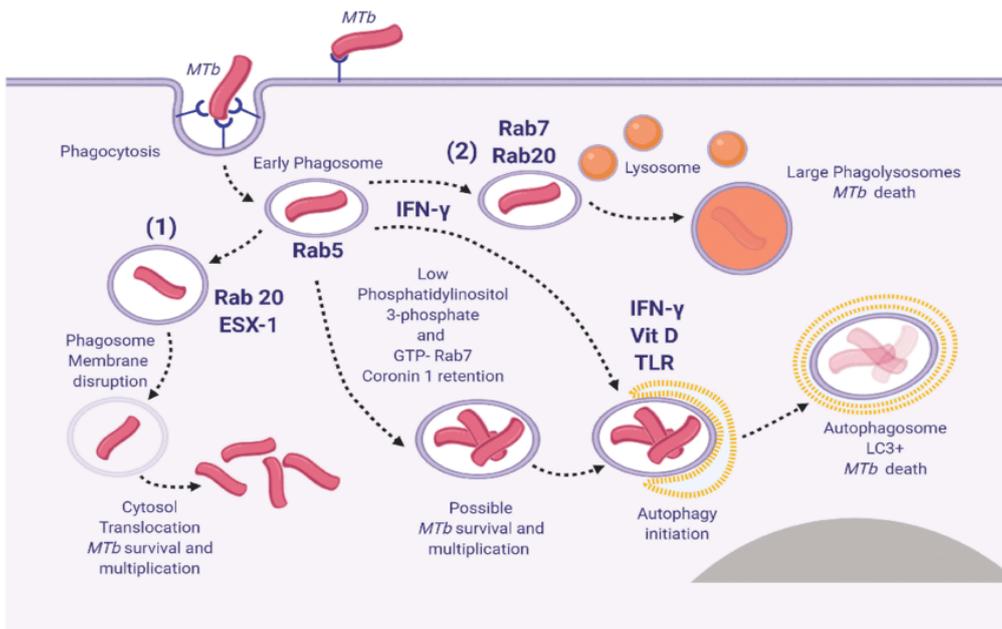


Fig. 11: MTB macrophage phagocytosis (1) MTB can prevent early phagosome maturation and by the action of Rab20-trafficking, ESX-1 destabilizes the phagosome membrane allowing bacteria to access the cytosol. (2) Some early phagosomes will undergo normal maturation, will fuse with the lysosomes and MTB will be killed. (3) Blocking of the early phagosome maturation (mainly by inhibiting PI3P generation) followed by MTB replication; (4) Delivery of the early endosomes or early-endosomes-to autolysosomes, where typically the activity of MTB it's controlled¹⁰⁶.

When unable to contain and eradicate MTB infection, engulfed macrophages die. MTB infected macrophages can undergo two forms of cell death (which may lead to drastic differences in the infection outcome): apoptosis or necrosis. Apoptosis is a process mediated by the caspase cascade, which results in the degradation of the cell and the formation of apoptotic vesicles. Apoptosis of MTB-infected cells is generally considered an advantage for the host because it eliminates pathogens, limits tissue inflammation, and promotes T-cell immunity activation ^{107, 108}. When an apoptotic macrophage dies, MTB is blocked into apoptotic bodies that are engulfed by activated macrophages and dendritic cells in a cascade process called efferocytosis. This facilitates the presentation of antigens to specific T-cells in order to activate the adaptive immunity. Necrotic death, on the other hand, is associated with the release of viable MTB bacilli for subsequent reinfection ¹⁰⁷. MTB generally kills infected cells via necrosis, which is characterized by the loss of macrophagic membrane integrity, and the release of cellular components in the extracellular space ¹⁰⁹. The role of MTB in cellular apoptosis is not clear. Several studies highlight that virulent MTB strains evolved several mechanisms to suppress host cell apoptosis and promote necrosis in order to replicate in the macrophages, and this effect is depending on both the virulence of the strains and on the multiplicity of the infection (MOI). MTB possesses several genes known to be involved in the apoptosis inhibition process (e.g., *nuoG* and *secA2*) ¹⁴⁶. Protein Phosphatase Mg²⁺/Mn²⁺-dependent 1A (PPM1A), has been identified as a key regulator of the innate antibacterial and antiviral response in macrophages and is targeted by MTB to prevent host macrophage apoptosis ¹⁰⁷. It is important to consider that the fate of an infected cell depends on many external factors and bacterial virulence only cannot explain the entire process. In general, virulent strains are associated with high levels of necrosis and apoptosis inhibition, however, there are studies that contradict this model, showing how even high virulent strains can induce a high level of apoptosis, even at low MOI; in fact, the *esxA*

gene, secreted by the ESX-1 system and encoding for the membranolytic protein EsxA, is considered pro-apoptotic ¹¹⁰.

1.7 Resistance to antimicrobials

Despite the introduction of an effective drug regimen (INH, RIF, PZA, and EMB), TB remains a major cause of morbidity and mortality globally. Drug resistance is due to genetic mutations (SNPs, deletions, and insertions), whereas drug tolerance is caused by epigenetic changes in gene expression and post-translational changes in drug-associated proteins.

WHO uses five categories to classify cases of drug-resistant TB: INH (INH)-resistant TB, RR-TB, MDR-TB, pre-XDR-TB, and XDR-TB (Table 6) ¹¹¹.

Table 6: WHO drug resistance definitions ¹¹¹

Drug resistance	Definition
Isoniazid-resistant-TB	Resistance to isoniazid
RR-TB	Resistance to rifampicin
MDR-TB	Resistance to both isoniazid and rifampicin
Pre-XDR-TB	TB caused by MTB strains that fulfil the definition of multidrug resistant and rifampicin-resistant TB (MDR/RR-TB) and which are also resistant to any fluoroquinolone.
XDR-TB	TB caused by MTB strains that fulfil the definition of MDR/RR-TB and which are also resistant to any fluoroquinolon and at least one additional Group A drug.

Anti-TB drugs are classified into three main categories. The first-line drugs are rifampicin (RIF / R), INH (INH / H), pyrazinamide (PZA / Z), ethambutol (EMB / E) (Table 7). They show a great activity against tuberculous bacilli and/or fewer side effects; they are the most used drugs and for this reason, the majority of resistance occurs against them.

Table 7: First line antitubercular drugs and their effects

Anti TB Drugs (First Line)	Isoniazid	Rifampicin	Ethambutol	Pyrazinamide
Main mechanism	Mycolic acid synthesis inhibition	RNA polymerase inhibition	Mycolic acid-Arabinosyltransferase inhibition	Mycolic acid inhibition
Effect	Bactericidal	Bactericidal	Bacteriostatic	Bactericidal

Drugs used for the treatment of MDR- can be divided into three groups ¹¹⁴: (Table 8)

- Group A: fluoroquinolones (FQ, levofloxacin and moxifloxacin), BDQ and linezolid (LZD)
- Group B: clofazimine (CFZ) and cycloserine or terizidone
- Group C: included all other medicines that can be used when a regimen cannot be composed with Group A and B agents.

Table 8: Grouping of medicines recommended for use in longer MDR-TB regimens

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Groups and steps	Medicine	Abbreviation
Group A: Include all three medicines	Levofloxacin <i>or</i> moxifloxacin	Lfx Mfx
	Bedaquiline ^{b,c}	Bdq
	Linezolid ^d	Lzd
Group B: Add one or both medicines	Clofazimine	Cfz
	Cycloserine <i>or</i> terizidone	Cs Trd
	Ethambutol	E
	Delamanid ^e	Dlm
	Pyrazinamide ^f	Z
Group C: Add to complete the regimen and when medicines from Groups A and B cannot be used	Imipenem–cilastatin <i>or</i> meropenem ^g	Ipm–Cln Mpm
	Amikacin (<i>or</i> streptomycin) ^h	Am (S)
	Ethionamide <i>or</i> prothionamide ⁱ	Eto Pto
	<i>P</i> -aminosalicylic acid ^j	PAS

The standard drug treatment regimen currently recommended by WHO, for drug-sensitive TB, consists of a 2-month intensive phase with RIF, INH, PZA, and EMB, followed by a 4-month phase with INH and RIF only. New regimens are now on use for MDR TB, in order to improve effectiveness and treatment adherence. In patients with confirmed RIF-susceptible, Isoniazid (INH)-resistant TB, treatment with RIF, EMB, PZA, and LEV is recommended for 6 months. A shorter BDQ-containing regimen of 9–12 months is recommended in patients with multidrug- or RIF-resistant TB. In multidrug TB all three Group A drugs and at least one Group B drug should be included to ensure the treatment to be effective. If only one or two Group A agents are used, both Group B agents are to be included. If the regimen cannot be composed with agents from Groups A and B alone, Group C agents are added (Table 8).

Genetic resistance to anti-tuberculous drugs develops thanks to spontaneous mutations at a variable frequency (10^{-6} to 10^{-8}) for the different genes involved (Table 9)¹¹². Once resistance mutation has occurred, the consequent selection is caused by selective pressure linked to an inappropriate regimen. An increased treatment failure and death is associated with the most resistant forms of TB ¹¹³.

Table 9: List of genes reported to be associated with phenotypical resistance to the listed TB-drugs ¹¹⁵

Drug	Tier 1	Tier 2
INH	<i>ahpC, inhA, katG</i>	<i>mshA, ndh, Rv1258c, Rv2752c</i>
RIF	<i>rpoB</i>	<i>rpoA, rpoC, Rv2752c</i>
EMB	<i>embA, embB, embC</i>	<i>embR, ubiA</i>
PZA	<i>pncA, clpC1, panD</i>	<i>Rv1258c, PPE35, Rv3236c</i>
FQ	<i>gyrA, gyrB</i>	None
BDQ	<i>pepQ, Rv0678, mmpL5, mmpS5, atpE</i>	<i>Rv1979c</i>
LZD	<i>rplC, rrl</i>	None
CFZ	<i>pepQ, Rv0678, mmpL5, mmpS5</i>	<i>Rv1979c</i>
DLM	<i>fgd1, ddn, fbiA, fbiB, fbiC, Rv2983</i>	None
AMK	<i>rrs, eis, whiB7</i>	<i>whiB6, ccsA, fprA, aftB</i>
STM	<i>rrs, rpsL, gid, whiB7, Rv1258c</i>	<i>whiB6</i>
ETO	<i>inhA, ethA</i>	<i>ethR, mshA, Rv3083, ndh</i>
KAN	<i>rrs, eis, whiB7</i>	None
CAP	<i>rrs, tlyA</i>	<i>whiB6, ccsA, fprA, aftB</i>

Tier 1: gene considered most probably to contain resistance mutations.

Tier 2: genes with a reasonable pre-test probability of containing resistance

INH: Isoniazid; RIF: Rifampicin; EMB: Etambutol; PZA: Pyrazinamide; FQ:

Fluoroquinolon; BDQ: Bedaquilin; LZD: Linezolid; CFZ: Clofazimin; DLM:

Delamanid; AMK: Amikacin; STM: Streptomycin; ETO: Ethionamide; KAN:

Kanamycin; CAP: Capreomicin

RIF is a drug discovered in 1967 that has become the key drug for TB treatment due to its high activity against MTB. RIF is active both on replicating and dormant mycobacteria. RIF potent antibiotic activity is due to its ability to inhibit bacterial RNA polymerase (RNAP) and thus cause cell death. Bacterial RNAP is an essential holoenzyme, structurally very similar to the homologous enzyme in Eukaryotes and Archaea despite the low sequence identity. Over 95% of the mutations conferring RIF resistance are grouped into three main sites: RIF clusters I, II, and III, but most are found in an 81bp region of cluster I called RRDR (RIF resistance determining region) ¹¹⁶. The most frequent replacements are Ser450Leu, His445Tyr, as the cytosine to thymine transitions are very common and are particularly favored in MTB because of its high G/C content and due to exposure to ROS (Reactive Oxygen Species), produced by the host's immune system during infection ¹¹⁷. Mutations in essential genes cause a decrease in

cellular fitness, resulting in a slower rate of growth and lower virulence. In RIF-resistant strains of MTB, it has been found that compensatory mutations in *rpoA* and *rpoC* (coding for the α and β 'subunits of RNA polymerase) may be present which do not cause drug resistance but restore the original fitness by compensating the *rpoB* mutation ¹¹⁸.

INH is a prodrug capable of interfering with the biosynthesis of MTB bacterial cell wall by altering the synthesis of mycolic acids. It is active only on growing bacteria and, once it has entered the cell, it needs to be activated to its anionic form by a catalase-peroxidase KatG encoded by the *katG* (*Rv1908c*) gene. The active form reacts with NAD⁺ to form an INH-NAD adduct capable of binding the active site of the NADH-dependent enoyl-ACP reductase (InhA), a type II fatty acid synthetase enzyme (FAS II) that reduces the monounsaturated acyl ACP to acyl ACP ¹¹⁹. Resistance to INH is associated with mutations in *katG* (50-95% of which 94% is Ser315Thr) and *inhA* (8-43%). Mutations in *ndh* gene, encoding for an NADH dehydrogenase, are associated with resistance to both INH and ethionamide. mutations in *ndh* gene cause defects in the oxidation of NADH to NAD thus accumulating NADH in the bacteria. High levels of NADH inhibits the binding of the INH-NAD adduct to InhA enzyme, impairing its activity ¹⁹.

EMB is a bacteriostatic drug that inhibits the proliferation of actively replicating bacteria and is used in combination with INH and RIF for standard TB treatments. It interferes with the transfer of arabinogalactan in the synthesis of the bacterial wall. The genes involved in drug resistance to EMB are part of the *emb* gene cluster, which includes 3 contiguous genes, *embA*, *embB* and *embC*. The first two encode for an arabinosyl transferase involved in the arabinosilation of arabinogalactans, while *embC* is involved in the arabinosylation of lipoarabinomannans. Most resistance-conferring mutations fall into a hot spot region called the ERDR (Ethambutol Resistance Determining Region) of *embB*, of which the most frequent is at codon 306 ¹²⁰.

Unlike any other anti-TB antibiotics, PZA does not exhibit activity against growing bacteria at physiological pH; however, it is an indispensable drug both for sensitive strains and for the treatment of MDR cases ¹²¹. It can act on latent state bacteria and only in acid pH environments, typically found within granulomas. PZA is an analog of nicotinic acid, it is a prodrug that needs activation in its active form (POA, pyrazinoic acid) by a pyrazinamidase encoded by the *pncA* gene ¹²². PZA passively enters the cell and is converted into POA which is secreted by a weak efflux pump. Here, in an acidic environment, it can re-enter and accumulate in the cell, where it destroys the membrane energy potential by interfering with the production of energy. PZA acts only in acidic compartments such as phagolysosomes, or in low pH environments that are formed in some areas of the lungs during the early stages of infection ¹²³. The mutations that confer resistance can be found throughout the *pncA* gene and in the 82 base pairs upstream of the 5' region of the gene. Mutations associated with resistance to PZA have also been recently described in the *rpsA* and *panD* genes, but their contribution to PZA resistance is poorly known ¹²⁴.

FQs inhibit type II topoisomerase (DNA gyrase) by binding to the enzyme and inhibiting DNA supercoiling. MTB has only topoisomerase II, unlike other bacteria that have topoisomerase II and IV. Resistance to FQs has been associated with the presence of mutations in a QRDR (Quinolone Resistance Determining Region) hot spot region in the *gyrA* gene and with mutations in the *gyrB* gene ¹²⁵.

Delamanid (DLM) is a nitro-dihydro-imidazo-oxazole derivative that inhibits mycolic acid synthesis. Its antibacterial activity is specific for mycobacteria, inhibiting methoxy- and keto-mycolic acid synthesis through the F420 coenzyme and generating nitrous oxide. Mycobacterial resistance to DLM may develop due to a mutation in one of the five coenzyme F420 genes necessary for the activation of DLM. Mutations in the genes of the F420 signaling pathway of the MTBC (*dnn*, *fgd1*, *fbiA*, *fbiB*, *fbiC*, and *fbiD*), can lead to DLM resistance ¹²⁶.

Bedaquiline (BDQ) is the first new antitubercular agent approved by the U.S. Food and Drug Administration (FDA) since 1970's. It possesses a potent bactericidal activity as an inhibitor of the ATP synthase proton pump. The main mutations that give resistance to BDQ are on the C subunit of ATP synthase encoded by the *atpE* (*Rv1305*) gene. In addition, mutations in the transcriptional regulator *Rv0687* gene were frequently associated with an increase of the BDQ MIC (Minimal Inhibitory Concentration) due to an upregulation of the MmpL5/MmpS5 efflux pump expression, which also leads to cross-resistance to CFZ¹²⁷.

The mechanism of antimicrobial action of CFZ is not entirely clear. In *Mycobacterium* TB, CFZ appears to act as a prodrug, which is reduced by the NADH complex causing the reduction of the available cellular ATP. CFZ is thought to compete with menaquinone (MK-4), a factor involved in the electron transfer chain¹²⁸.

LZD is an antibiotic in the oxazolidinone class. It inhibits protein synthesis by binding the bacterial 23S ribosomal RNA subunit and preventing the formation of the 70S ribosomal unit. Drug resistance for LZD usually occurs by SNPs in *rrl* and *rplC* genes. Mutations in *rplC* are associated with high LZD MIC values, while *rrl* mutations correlate with lower LZD MICs¹²⁹.

Aim of the PhD project: role of the genetic diversity of MTB and its involvement on macrophage infection and on drug tolerance

Tuberculosis (TB) remains a major cause of morbidity and mortality worldwide and new vaccines candidates have not shown advantages compared to the Bacillus Calmette-Guerin (BCG) vaccine. *Mycobacterium tuberculosis* (MTB) evolved several strategies to survive within host cells and the metabolism of the bacterium is closely joined with the intraphagosomal milieu and to the immunological state of the host ¹⁴¹. In addition, the genetic variability of MTB clinical isolates, and drug resistance development, have important consequences on the outcome of infections and on diagnostic approaches ^{43, 142}. In fact, as outlined strain-dependent variations (MTB lineages) have been associated with key pathogenic aspects ^{143, 144}, thus contributing to the wide spectrum of TB clinical presentations ^{23, 145}.

So far, the research in the field focused on either the host variability or pathogen variability in a mutual and exclusive way, and anyhow these topics remain in their infancy in TB. In this PhD project, we aimed at better understanding the role of the genetic diversity of MTB in the pathogenesis and in drug tolerance. In particular, for the pathogenesis, we hypothesized to design an experimental plan which brought together macrophage phenotypic plasticity and MTB genetic variability to advance our knowledge in the host-pathogen interactions. We analyzed the acidification of the phagosomes, the autophagic flux, the induction of apoptosis, and cytokine production in classically (M1) and alternatively (M2) activated THP-1 macrophage-like cells challenged with different MTB clinical isolates representative of ancient and modern phylogenetic lineages.

For a better understanding of the role of the genetic diversity of MTB in drug resistance mechanisms, we decided to focus our attention on the neglected topic of smallRNAs. In our laboratory, we consolidated the evidence available on the smallRNA candidates. This was the starting point for addressing the role of lineage-specific mutations mapping in smallRNAs

in this project. The analysis identified a mutation-specific for ancient lineages and mapping in the promoter region of the sRNA *ncRv0842c*, cis-encoded to the *Rv0842* gene, coding for a putative efflux pump reported to be involved in RIF resistance development. Accordingly, we characterized the role of *ncRv0842c* during RIF challenge to provide insights on the different mechanisms of drug tolerance among phylogenetically different strains of MTB.

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Chapter 2

Interplay between macrophage heterogeneity and *Mycobacterium tuberculosis* complex phylogenetic lineages during infection

Abstract

Host heterogeneity and bacterial genotypic background are key players in determining the outcome of infection, but their interplay remains largely uncharacterized. To reduce this knowledge gap, we considered cytokines production, phagolysosomal acidification, and mycobacterial survival during infection with either modern or ancient MTB lineages in classically and alternatively activated macrophages.

THP-1-derived macrophages either polarized toward M1 or M2 subtypes were infected with: H37Rv, H37Ra, Beijing (lineage L2) *M. africanum* (lineage L6). Infection was carried out at MOI 10:1. Live single-cell confocal microscopy was used to analyze phagolysosomal acidification at 6h and 24h post-infection (p.i.), autophagy and apoptosis at 24h p.i. Supernatants were collected at 4, 24, and 48h p.i. for measuring 20 cytokines/chemokines levels by Bio-Plex Multiplex-Immunoassay. After 48h p.i. cells were lysed for CFU count on 7H10 agar plates.

M2 infected cells induce the production of M1 specific marker, indicating an M1 shift at early phases. Variability among cytokine and chemokine pattern production appears to be higher in M1 cells, where the different MTB categories induce different host responses. In general, ancient/specialist strains induce a lower production of pro-inflammatory cytokine compared to modern/generalist strains.

Confocal microscopy showed that in M1 cells acidification is delayed by modern strains, whereas at earlier time points ancient virulent strains are not blocking acidification. At 24h p.i. none of the strains is blocking phagosomal acidification, but ancient strains displayed lower levels of acidification compared to modern strains. In M2 cells at 24h p.i. modern strains blocked acidification, whereas ancient strains were less efficient in

blocking the acidification of phagosomes. CFU data show a faster reduction of the bacterial load in L6 when compared to the modern ones. H37Ra CFUs are lower than H37Rv strains in both M1 and M2 cell types.

Our data highlight the role played in the infection by both the macrophage's activation status and the pathogen's features.

Introduction

Despite our increasing understanding of the pathogenesis of tuberculosis (TB), several gaps remain to meet the challenge of eradicating the disease.

Mycobacterium tuberculosis (MTB) survives in macrophages that are crucial immune system components for the formation of the granuloma, but also provide a survival niche from which the bacteria may disseminate. While in the macrophages, MTB is exposed to an array of stresses, including oxidative and nitrosative stresses, iron restriction, starvation, and low pH. MTB can block the phagosome maturation and the consequent acidification of the compartments, it can modulate the apoptosis and it's able to inhibit ROS production ¹.

MTB has been considered a unique clone and the laboratory strain H37Rv has been largely used as a reference strain for research studies. However, the MTB phylogeny is composed of at least 7 different lineages and H37Rv is not representative of the genetic heterogeneity of this pathogen. This genotypic variation is associated with variation in transmission capacity and in the propensity to acquire drug resistance. This means that regional variation in the prevalence of specific lineages or sub-lineages can have consequences for the observed epidemiology of TB ². More broadly, the different distribution of the lineages also reflects the heterogeneity in the host-pathogen co-evolution and adaptation. Within the MTB complex (MTBC) it is possible to distinguish phylogenetically modern lineages (L2, L3, L4), that are considered to be more virulent, generalist (e.g. capable of causing disease in many different human populations), most geographically widespread, and ancient lineages (L1, L5, L6, L7), which are less virulent and that are more prone to infect populations in restricted geographical areas of the globe. Modern lineages 2 (Beijing), 3 (CAS/Delhi) and 4 (Haarlem, H37Rv) possess a large deletion known as TbD1 and are associated with major TB epidemics, and high transmission rate compared to Lineage 1 (Indo-oceanic), 5 and 6 (*M. africanum*). Modern MTB strains,

in general, induce low early inflammatory response than ancient lineages^{3, 4}.

Despite this microbe-centric classification, evaluating the virulent features of a microorganism is strongly related and dependent on the host system⁵. In this context, human heterogeneity at various levels should be taken into consideration. Besides genetic variability among individuals, human alveolar macrophages have been reported to show a mixed, plastic M1/M2 macrophagic phenotype^{6, 7}. However, only a few studies kept into account the polarization status of the host's macrophages in *in vitro/ex vivo* infection models⁸.

In this work, we studied early host-pathogen interaction between MTB and human macrophages, taking into consideration both host macrophage phenotypic variability (resembling alveolar macrophages plasticity) and MTB genetic diversity considering major phylogenetical differences. In particular, we focused our attention on phagolysosomal acidification, autophagy (self-degradative process), apoptosis (programmed cell death), and necrosis (cell injury) in classically (M1) or alternatively (M2) activated THP-1-derived macrophage-like cells. A single-cell approach was considered to better appreciate the heterogeneity of the fate of the host-pathogen interaction. Cytokines induction, macrophagic and bacterial survival were also considered to investigate the virulence features of the MTB lineages considered.

Materials and Methods

- ***M. tuberculosis* strains selection and classification**

Clinical isolates representative of main ancient (L1, L6) and modern lineages (L2, L4) and previously characterized were kindly provided by Prof. Stefan Niemann (Research Center Borstel, Borstel, Germany) ⁹. *M. tuberculosis* H37Rv NCTC 7416, *M. tuberculosis* H37Ra NCTC 7417, and *M. bovis* BCG NCTC 5692 were also included (Table 1).

Strain lineages were then classified and grouped according to seven categories considering the capacity to cause outbreaks and to develop drug resistance and/or association with worse clinical outcomes based on literature data (Table 1) ¹⁰⁻²¹:

- ✓ **Basic:** grouping is simplified by considering ancient lineages less virulent, and modern lineages more virulent
- ✓ **Disease:** grouping is based on the ability of the strain to cause disease in an immunocompetent host
- ✓ **Phylogenesis:** grouping is defined according to the phylogeny of the lineages
- ✓ **Virulence, absolute:** grouping is based on virulence features
- ✓ **Virulence, relative:** grouping is based on virulence features in comparison with the H37Rv reference strain
- ✓ **Geographical distribution and host adaptation:** grouping is based on the definitions provided in Stucki et al related to the niche width of MTB strains ¹⁶.
- ✓ **Laboratory adaptation:** grouping is based on the laboratory selection of the strain ²²

- **Sequencing**

Whole-genome sequencing was performed by Illumina technology. Briefly, DNA libraries were generated using Nextera XT Library Prep kit (Illumina, San Diego, USA) according to the manufacturer's protocol, and libraries

were run on MiniSeq or NextSeq devices (Illumina, San Diego, USA). Variant calling for genetic analysis of the strains was performed according to the MTBseq pipeline (reference genome: *M. tuberculosis* H37Rv Genbank NC_000962.3)²³.

- **Electroporation**

All mycobacterial strains were engineered with the pSMT3::mCherry plasmid (Ex/Em 587-610 nm). The pSMT3::mCherry plasmid was a kind gift from Prof. Giovanni Delogu (Università Cattolica del Sacro Cuore, Rome, Italy). Briefly, bacteria were cultured in liquid 7H9 medium + Albumin Dextrose Catalase Oleic Acid (OADC) supplement (Becton Dickinson New Jersey, USA), 0.05% Tween[®] 80 (Sigma Aldrich, St. Luis, MO, USA) until OD₆₀₀ 0.4-0.6. Bacteria were centrifuged and washed twice with 10% glycerol, to eliminate salt and contaminants, in order to achieve the electro-competence, and resuspended in 10% glycerol for frozen stockage. Competent cells were electroporated (2500V, 25µF, 200 ohms) with 2µg of salt-free plasmid and selected on 50 µg/mL Hygromycin (Roche, Basel, Switzerland) 7H10 agar plates (Becton Dickinson, New Jersey, USA)²⁴.

For macrophage infection, the strains were thawed on selective Middlebrook 7H10 agar medium supplemented with 10% OADC supplement, 0.05% Tween[®] 80. and 50µg/mL Hygromycin. A single colony was picked up and used for inoculating Middlebrook 7H9 liquid medium supplemented with 10% Middlebrook OADC supplement, 0.05% Tween[®] 80. and 50µg/mL hygromycin and incubated at 37 °C until OD₆₀₀ ~0.3. Bacterial suspensions were centrifuged at 17000 g for 5 minutes, washed with sterile PBS, and resuspended in complete RPMI (cRPMI) medium w/o antibiotics, de-clumped by 10 passages through a 21-gauge needle, and diluted in cRPMI immediately before the infection.

Table 1: MTB strains considered in this study and classification used for the analysis of the results. Assay performed: y (yes) n (no)

Strain	Alias	Lineage Nomenclature	Lineage type	Grouping							Assay performed					
				Basic	Disease	Phylogenesis	Virulence_abs	Virulence_ref	Geo_host_adaptation	Lab adaptation	Phagosomal acidification	Autophagy	Apoptosis	Cytokine production	CFU	Cell Survival
<i>M. africanum</i> 5684/02	Afri 1	6	ancient	Less/ancient	Causing	Ancient	Less	Less	Specialist	Clinical	y	y	y	y	y	y
<i>M. africanum</i> 10514/01	Afri 2	6	ancient	Less/ancient	Causing	Ancient	Less	Less	Specialist	Clinical	y	y	y	y	y	y
<i>M. bovis</i> BCG NCTC 5692	BCG	bovis	animal	BCG	Non-causing	Animal	Less	Less	Other	Laboratory	y	y	n	n	n	y
<i>M. tuberculosis</i> 12594/02	Beijing 1	2	modern	More/modern	Causing	Modern	More	More	Generalist	Clinical	y	y	y	y	y	y
<i>M. tuberculosis</i> 1934/03	Beijing 2	2	modern	More/modern	Causing	Modern	More	More	Generalist	Clinical	y	y	y	y	y	y
<i>M. tuberculosis</i> CDC1551	CDC	4	modern	More/modern	Causing	Modern	More	More	Intermediate	Clinical	y	n	n	n	n	y
<i>M. tuberculosis</i> 1797/03	EAI 1	1	ancient	Less/ancient	Causing	Ancient	Less	Less	Specialist	Clinical	y	n	n	n	n	n
<i>M. tuberculosis</i> 4850/03	EAI 2	1	ancient	Less/ancient	Causing	Ancient	Less	Less	Specialist	Clinical	y	n	n	n	n	n
<i>M. tuberculosis</i> H37Ra NCTC 7417	H37Ra	4	modern	H37Ra	Non-causing	Modern	Less	Less	Other	Laboratory	y	y	n	y	y	y
<i>M. tuberculosis</i> H37Rv NTCT 7416	H37Rv	4	modern	H37Rv	Causing	Modern	More	H37Rv	Generalist	Laboratory	y	y	y	y	y	y

- **Cells preparation and infection for confocal microscopy** ²⁵

THP-1 (ATCC® TIB-202™) cells were differentiated in cRPMI (with L-glutamine 2mM, Hepes 10mM, Non-essential amino acids 100nM, Sodium Pyruvate 1mM, Sigma Aldrich, Missouri, USA) supplemented with 10% FBS (Fetal Bovin Serum, Euroclone, Italy), antibiotics (penicillin G 100 U/ml and streptomycin sulfate 100 U/ml, Sigma Aldrich Missouri, USA), and 100 nM of phorbol 12-myristate 13-acetate (PMA, Sigma Aldrich Missouri, USA) for 3 days in Eppendorf Cell Imaging 96-wells plates (55000 cells/well) bottom glass 170µm thick (Eppendorf, Germany) ²⁶. Type I collagen (Sigma Aldrich, Missouri, USA) was used for cell adhesion. Given the lack of standard protocols for recapitulating the alveolar macrophage *in vitro* cell lines, we opted to consider the two extremes of the dynamic state of activation of the macrophage. Three days after removal of the PMA, macrophage-like cells were polarized in cRPMI medium without antibiotics for 16 h with either lipopolysaccharide (50ng/mL, Sigma Aldrich Missouri, USA) to induce an M1-like phenotype or human IL-4 (20 ng/mL, E. coli-derived human IL-4 protein (R&Dsystems, Minnesota, USA) to induce an M2a-like phenotype prior to the infection (from here on “M2”) ²⁷. Effective polarization of cells was evaluated by checking the expression of specific markers for M1 (TNF α , IL-1 β , CD80, IL-6), and M2 (CD206, IL-10) (data not shown). THP-1 derived macrophages were infected with mycobacteria at a theoretical multiplicity of infection (MOI) 10:1. Infection was carried out by centrifugation (300 g x 5 min, room temperature) followed by two hours of incubation at 37 °C CO₂ 5.0%. Macrophage-like cells were then washed twice and supplemented with cRPMI without antibiotics. cRPMI without phenol red in case the infection was set up for imaging readouts.

- **Live imaging microscopy**

Phagolysosomal acidification: macrophages were stained with Hoechst 33342 NucBlue™ Live ReadyProbes™ Reagent (Ex/Em 360-460 nm, ThermoFisher, Massachusetts, USA) for nuclei, and with LysoSensor™ Green DND-189 (Ex/Em 443-505 nm, pK_a 5.2, ThermoFisher) for monitoring

phagolysosome acidification according to manufacturer's instructions. Indeed, the LysoSensor dye is an acidotropic probe that accumulates in acidic organelles as the result of protonation. This protonation also relieves the fluorescence quenching of the dye by its weakly basic side chain, resulting in an increase in fluorescence intensity. Thus, the LysoSensor Green exhibits a pH-dependent increase in fluorescence intensity upon acidification only, with very low background in the cytosol as it is almost nonfluorescent except when inside acidic compartments. All images have been acquired and analyzed using the same settings for allowing quantitative comparison of LysoSensor intensities (Supplementary Phagolysosomal acidification). Imaging experiments were performed at 6- and 24-hours post-infection (p.i.). For each strain, 6 images (2 positions for each well out of 3 wells per strain) were acquired as z-stacks (0.89 μm x 7 stacks) in at least two independent experiments.

Induction of autophagosomes: macrophages were stained with Hoechst 33342 for nuclei (Ex/Em 450-461 nm), and with CYTO-ID[®] Green (Ex/Em 499-548 nm) from the CYTO-ID[®] autophagy detection kit 2.0 (Enzo Biochem, Farmingdale, NYS, USA) according to manufacturer's instructions for monitoring induction of autophagosomes. The dye exhibits bright fluorescence upon incorporation into pre-autophagosomes, autophagosomes, and auto-phagolysosomes, whereas it shows limited staining of lysosomes²⁸. Imaging experiments were performed at 24 h p.i. As control of autophagy induction, macrophages were treated with 500nM of rapamycin and 50 μM of chloroquine overnight. For each strain, 8 images (2 positions for each well out of 4 wells per strain) were acquired as z-stacks (1.48 μm x 4 stacks) in at least two independent experiments.

Induction of apoptosis and necrosis: macrophages were stained with CytoCalcein Violet for cytosol (Ex/Em 405-450 nm, a marker for living cells cytoplasm), with Apopxin Green (Ex/Em 490-525 nm, apoptotic marker), and 7-AAD (Ex/Em 546/657, necrotic marker) according to the

Apoptosis/Necrosis assay kit (Abcam, Cambridge, UK) for monitoring apoptosis/necrosis at 24 h p.i. Apopxin targets the hallmark of apoptosis-exposed phosphatidylserine. Loss of plasma membrane integrity represents a straightforward approach to demonstrate late-stage apoptosis and necrosis, and it is detected by the ability of the membrane-impermeable 7-AAD to label the nucleus. As control of apoptosis, macrophages were treated with 5 µg/mL cyclohexamide and 2ng/mL TNFα overnight. For each strain, 4 images (2 positions for each well out of 2 wells per strain) were acquired as z-stacks (1.48 µm x 4 stacks) in at least two independent experiments.

All experiments were carried out using a Leica TCS SP5 confocal microscope equipped with an incubator chamber allowing to stably maintain cells at 37 °C in 5% CO₂. Unless otherwise specified, images were acquired as 16-bit, 1024×1024 resolution, TIFF files using a 63x, 1.40 NA immersion oil objective. A 405 nm UV diode laser was used for Hoechst 33342, a 458 nm Argon laser was used for LysoSensor, a 488 nm laser was used for CYTO-ID and Apopxin, a 561 nm laser was used for dsRed and 7-AAD.

- Immunofluorescence

THP-1 PMA derived macrophages were plated on 96well plates as described above and infected with MTB strains at MOI 10:1. After 24h post-infection cells were stained with LysoSensor Green and Hoechst in order to measure lysosomal acidification. After the image acquisition, well positions were saved for the subsequent immunofluorescence analysis on the same selected cells. Cells were fixed with 3,7% Paraformaldehyde for 10 minutes, permeabilized with an Hepes-Triton-X-100 buffer (Sigma Aldrich, Missouri, USA), and stained with antibodies. Targeted human protein: IFNγ, TNFα, MIP-1β, and MCP-1. Primary and secondary antibodies have been used at a final concentration of 1 µg/ml (Table 2).

For IFNγ and TNFα cells were analyzed at confocal microscopy using the pre-saved positions in order to compare acidification and cytokine production within the same cells.

Table 2: Primary and secondary antibodies for ImmunoFluorescence assay

Protein target (human)	Primary antibody			Secondary antibody		
IFN-gamma	IFN gamma Monoclonal Antibody (25718)	Mouse/IgG2a	Unconjugated	Donkey anti-Mouse IgG (H+L) Highly Cross-Adsorbed Secondary Antibody	Donkey/IgG	Alexa Fluor Plus 647 # A32787
CCL4 (MIP-1 beta)	CCL4 Polyclonal Antibody	Rabbit/IgG	Unconjugated	Goat anti-Rabbit IgG (H+L) Highly Cross-Adsorbed Secondary Antibody	Goat/IgG	Alexa Fluor Plus 488 # A32731
TNF-alpha	TNF alpha Monoclonal Antibody (28401)	Mouse/IgG1	Unconjugated	Donkey anti-Mouse IgG (H+L) Highly Cross-Adsorbed Secondary Antibody	Donkey/IgG	Alexa Fluor Plus 647 # A32787
MCP-1	MCP-1 Polyclonal Antibody	Rabbit/IgG	Unconjugated	Goat anti-Rabbit IgG (H+L) Highly Cross-Adsorbed Secondary Antibody	Goat/IgG	Alexa Fluor Plus 488 # A32731

- **Images and data analysis**

Images were processed for single-cell analysis with Fiji software ²⁹. Stacks have been analyzed by applying the maximum intensity projection method. As a general rule, at least 5 infected plus 5 non-infected cells (unless not available) were considered for each acquisition. Effective cell infection was visually inspected by the use of orthogonal views.

Phagolysosomal acidification: Single-cells were manually segmented. Acidified phagolysosomes and mycobacteria were selected by thresholding the 458 nm and the 561 nm channels, respectively. LysoSensor signal was acquired as mean signal intensity in cells region of interest (ROI). Acidified phagolysosomes and mycobacteria counts were also acquired for each single cell ROI. To analyze the spatial relationship between bacteria and lysosomes (colocalization) we wrote a custom pipeline in Matlab. Briefly, the confocal stacks are maximal projected and the individual cells are segmented manually. Next, the cell nuclei and the bacteria are automatically segmented, and the average LysoSensor intensity around the segmented bacteria L_{bac} is calculated (in a region of 1 pixel in width). Such intensity is compared to the average LysoSensor intensity in the cell $\langle L \rangle$, to provide an inclusion index $I_i = (L_{bac} - \langle L \rangle) / \langle L \rangle$. Such index would be higher than zero when bacteria are located inside lysosomes and lower than zero when bacteria are excluded from lysosomal compartments. Random positioning of bacteria would provide $I_i = 0$. All images have been acquired and analyzed using the settings as reported in Supplementary Phagolysosomal acidification.

Induction of autophagosomes: Single-cells were manually segmented. Autophagosomes and mycobacteria were selected by thresholding the 488 nm and the 561 nm channel, respectively. CYTO-ID signal was acquired as mean signal intensity in cells region of interest (ROI). mCherry mean signal across the cells ROI has been used as a proxy for mycobacterial burden. The autophagic pathway was analyzed by considering both the number and the size of autophagosomes ³⁰. The number of autophagosomes per cell was calculated using MatLab. All images have been acquired and analyzed using the settings as reported in Supplementary Autophagy.

Induction of apoptosis and necrosis: Single cells were manually segmented. CytoCalcein Violet, and Apopxin Green were acquired as mean signal intensity in cells region of interest (ROI). Signal from 7-AAD in the nuclei was never observed at the timepoints considered, thus analysis on the necrosis was not performed. Apoptotic cells have been identified using FlowClust (ver. 3.9), a Bioconductor R package implementing a robust model-based clustering approach based on multivariate t mixture models with the Box-Cox transformation for gating cell populations (rule of identifying outliers: 83% quantile) ³¹. All images have been acquired and analyzed using the settings as reported in Supplementary Apoptosis. High mycobacterial burden was considered when dsRed signal was found to be equal or above the central value of the distribution of all dsRed signals in infected cells. Signals below this value were defined as low mycobacterial burden.

Statistical analysis: Linear mixed-effects (LME) models were employed to evaluate differences among strains in the three main fields of investigation (phagolysosomal acidification, autophagic pathway, and apoptosis) ^{32, 33}. Uninfected control THP-1 macrophages (M1 or M2) were used for normalization. The logarithmic transformation or the Ordered Quantile normalization transformation of the outcome were considered to satisfy underlying model assumptions. Zero-inflated Mixed-effects Poisson

models were applied to properly analyze count data with excess zeros ³⁴. These procedures were employed to properly account for the dependence structure among observations, by including nested random-effect terms, thus considering in the model unobservable sources of heterogeneity among experimental units. Post-hoc analysis was performed considering all the pairwise comparisons. Linear mixed-effects models were also applied to properly evaluate the relationship between two numeric variables. Analyses were performed using R statistical software ³⁵.

- **Cells preparation and infection for CFUs and cytokine immunoassay** ³⁶

For CFU and cytokine analyses we used the same infection protocol as described for confocal microscopy, except for the fact that infections were carried out in 6well (Nunc™ Cell-Cultured Treated 1,25 x 10⁶ cells/well) instead of 96well plates. Bacteria used for these experiments are indicated in Table 1, and included modern strains H37Rv NCTC 7416, H37Ra NCTC 7417, 2x L2, 2x L6. Macrophage-like cells were then washed twice and supplemented with cRPMI without antibiotics. Supernatants were collected at 4, 24 and 48 hours post-infection and analyzed with Bio-Plex Pro™ Human Cytokine 20-plex Assay (Bio-Rad, California, USA) according to the manufacturer's instructions. Cytokines and chemokines included in the customized panel were selected in order to cover the entire spectrum of most commonly activated macrophagic molecules in responding to MTB infection. Pro-inflammatory cytokines: IL-1 β , IL-6, TNF α , IL-15; IFN γ ; IL-18; Anti-inflammatory cytokines: IL-1ra, IL-10; Chemokines: IP-10 (cxcl10-Interferon gamma-induced protein 10), MIP-1 β (CCL4-Macrophage Inflammatory Protein), IL-8 (CXCL8), MCP-1 (CCL2-monocyte chemoattractant protein-1), RANTES (CCL5), SDF-1 α (CXCL12), GRO- α (CXCL1), MIP-1 α (CCL3-Macrophage Inflammatory Protein); Growth factors: VEGF (Vascular endothelial growth factor), PDGF-bb (Platelet-derived growth-stimulating factor), G-CSF (Granulocyte colony-stimulating factor), FGF basic.

Four independent experimental replicates were considered. Linear mixed-effects (LME) models were employed to evaluate differences among strains. Logarithmic and Ordered Quantile normalization transformations of the outcome were considered to satisfy underlying model assumptions. These procedures were employed to properly account for the dependence structure among observations, by including nested random-effect terms, thus considering in the model unobservable sources of heterogeneity among experimental units^{32, 33}. Post-hoc analysis after LME was performed considering all the pairwise comparisons. Analyses were performed using R statistical software.

For mycobacterial survival, cells were lysed with sterile water. Supernatants and cell lysate were serially diluted and plated on 7H10 agar plate in order to count MTB CFUs at 4h, 24h, and 48h post-infection.

- **Macrophage survival**

THP-1 were plated and differentiated in 96well plates as described above (M1 or M2) and infected with MTB strains at MOI 5:1. MTB strains used for this experiment include the laboratory strains H37Rv, H37Ra, *M. bovis* BCG NCTC 5692, CDC 1551 and clinical strains 2x L2, 2x L6. After 48h and 5 days post-infection, cells viability was assessed by the use of the crystal violet assay as previously described³⁷. Briefly, cells were fixed with cold (-20°C) methanol (Sigma Aldrich, Missouri, USA) and stained with crystal violet. This experiment relies on the detachment of adherent cells from cell culture plates during cell death where only live, adherent cells are fixed and stained with Crystal violet. After a wash step, the Crystal violet dye is solubilized with lysis buffer (0.1M sodium citrate, 50% Ethanol, pH 4,2) and measured by absorbance at 570 nm using the BioTek™ Gen5™ software (Agilent). The percentage of cell death compared to the uninfected control was considered.

Results

We report here main findings using the grouping geographical-host (geo-host), as it somehow encompasses different features (phylogeny, distribution, host adaptation); however, it should be kept in mind that this grouping would lose to pick-up relevant differences between laboratory and clinical isolates and would not allow gaining details in terms of direct comparison with H37Rv, for example. For this reason, the same analyses described here for the group geo-host adaptation have been performed for all the grouping proposed in Table 1 and reported as Supplementary.

- **Phagolysosomal acidification**

The LysoSensor Green has been used to evaluate the level of acidification of phagolysosomes of THP-1 M1 and M2 macrophages infected with H37Rv, H37Ra, BCG, CDC, Afri, EAI and Beijing at 6 and 24 h p.i. (Fig. 1). A total of 7384 single cells (4098 at 6 h p.i.; 3286 at 24 h p.i.) were considered for the analysis (on average, for each strain tested, M1: 176 not infected cells + 172 infected cells; M2: 180 not infected cells + 174 infected cells) (Supplementary Phagolysosomal acidification). Data were normalized on THP-1 macrophages not infected control and strains were grouped as previously reported in Table 1.

At 6 h p.i. in M1 macrophages, only categories intermediate and other displayed blocking of phagolysosomal acidification, and specialist strains showed the highest number of acidified macrophages (50% vs <25% of all the other categories). A bystander effect in non-infected M1 macrophages was observed for specialist strains at 6 h p.i, despite it was not found statistically significant (Fig. 2). At 24 h p.i. none of the categories is blocking the acidification of the phagolysosomes. However, despite an increase in the acidification of the phagolysosomes was observed, the number of macrophages displaying such acidification was found generally low (<25%) (Fig. 3).

Phagolysosomal acidification

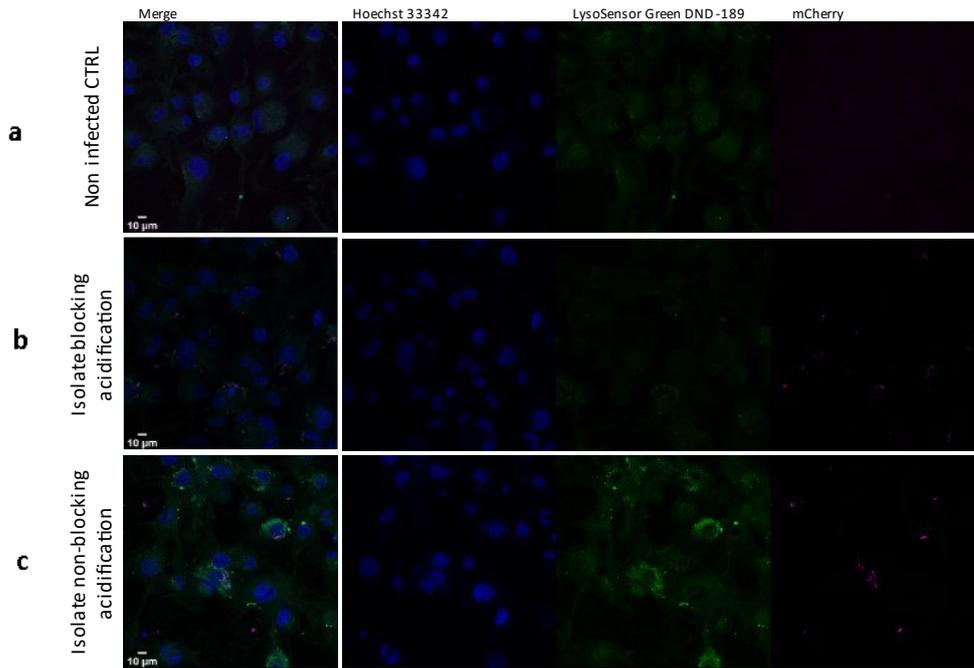


Fig. 1: THP-1 cells non-infected (a); infected with acidification-blocking bacteria (b); infected with non-acidification-blocking bacteria (c). The fluorescence images were acquired after staining with LysoSensor (Green) and Hoechst (Blue). Bacteria are reported in red. The reported images are illustrative of the pipeline adopted

M1 – 6 h p.i.

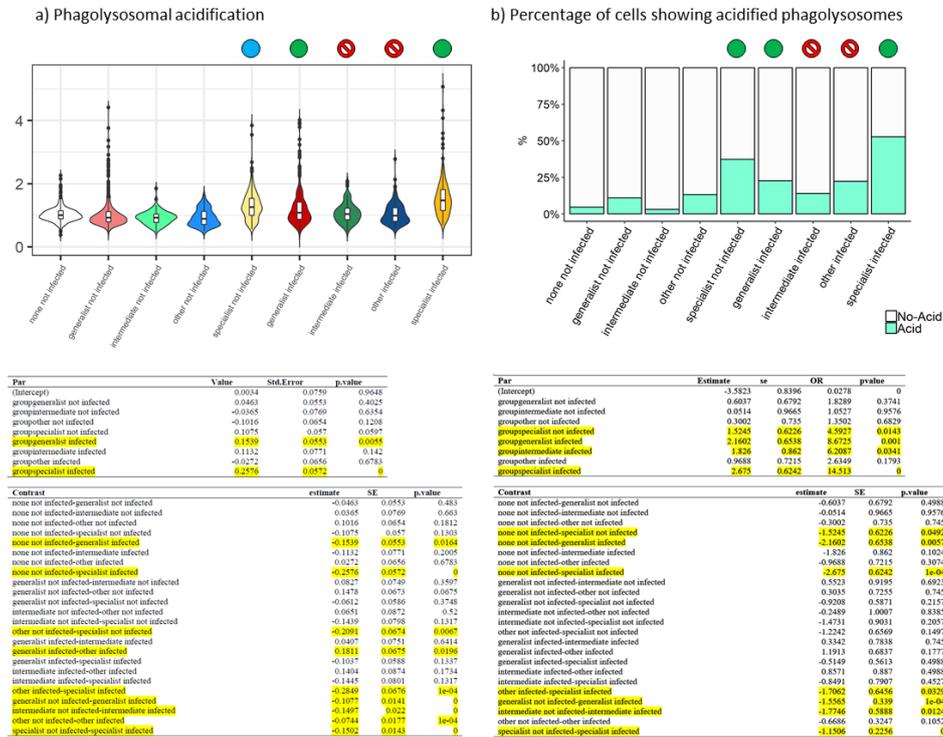


Fig. 2 : Phagosomal acidification at 6h p.i. in M1 cells with statistical reports

M1 – 24 h p.i.

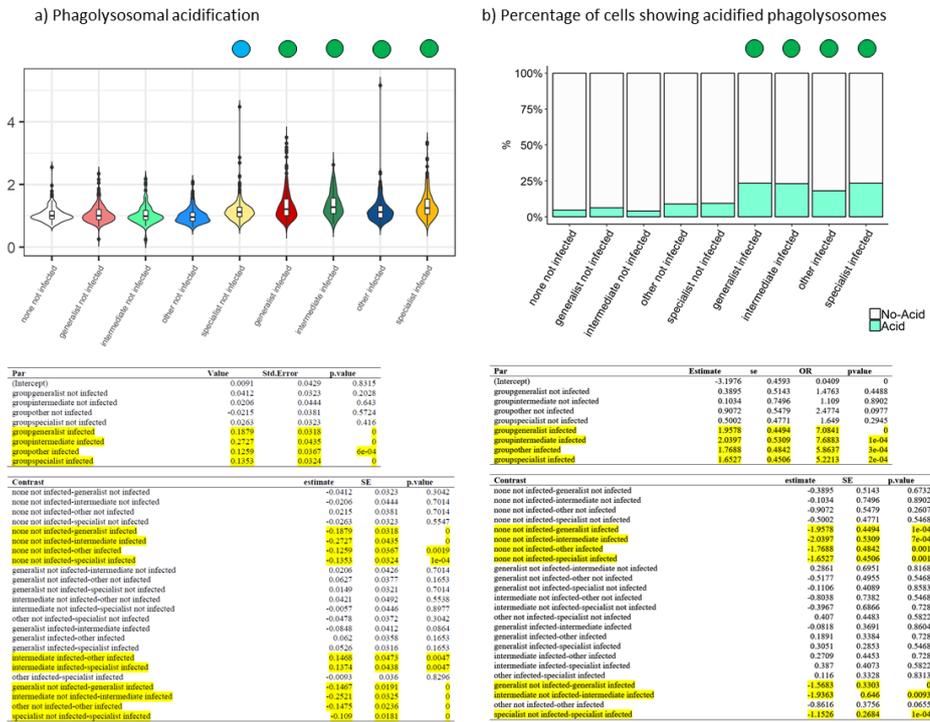


Fig. 3 : Phagosomal acidification at 24h p.i. in M1 cells with statistical reports

In M2 macrophages, none of the category is blocking the acidification of the phagolysosomes at 6 h p.i. and between 25% (other) and 50% (specialist and generalist) of macrophages showed acidification. A bystander effect in non-infected M2 macrophages was observed for specialist strains at 6 h p.i., with approx. 40% of bystander cells showed acidification (Fig. 4). As the time of infection progresses, generalist and intermediate categories are blocking the acidification of the phagolysosomes. The number of acidified macrophages also decreased below 25% except for the category other (50%). A bystander effect in non-infected M2 macrophages was observed for specialist strains at 24 h p.i., despite it was not found statistically significant (Fig. 5).

M2 – 6 h p.i.

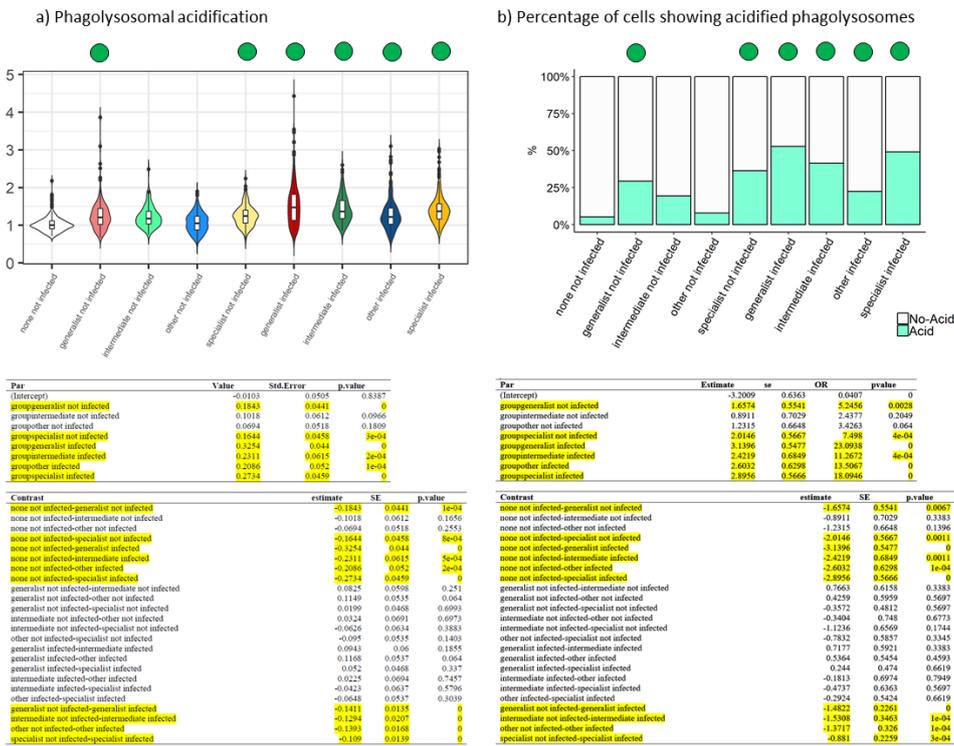


Fig. 4 : Phagosomal acidification at 6h p.i. in M2 cells with statistical reports

M2 – 24 h p.i.

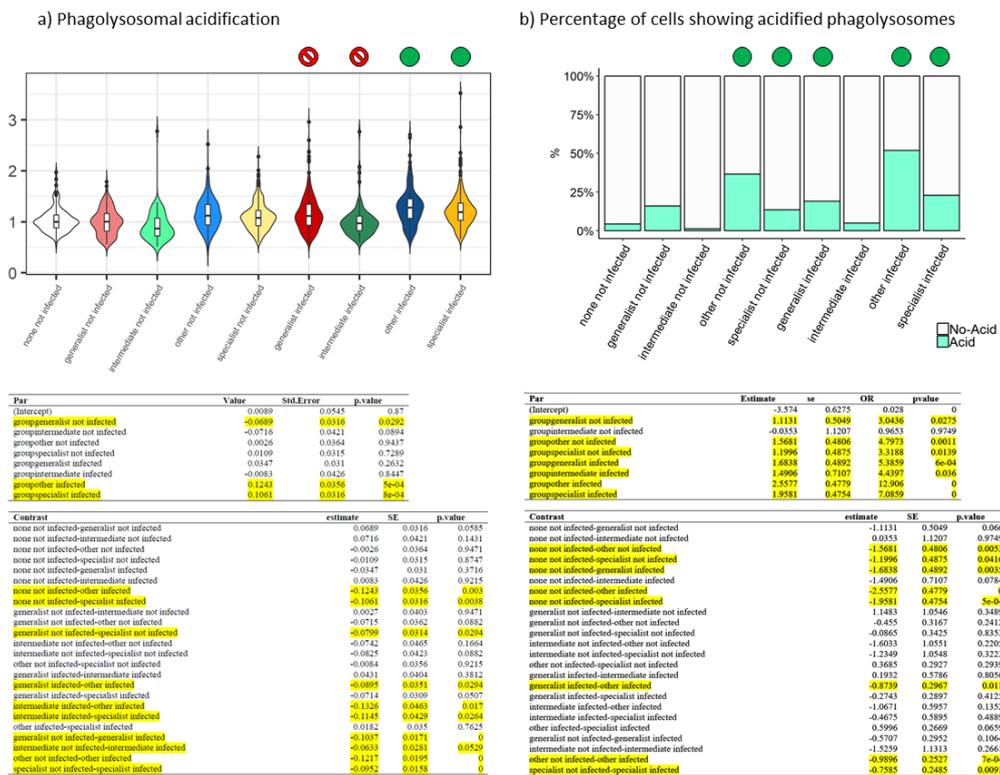


Fig. 5 : Phagosomal acidification at 24h p.i. in M2 cells with statistical reports

In terms of the number of mycobacteria per cell (MOI), in M1 macrophages no statistically relevant differences in effective MOI were observed at 6 h p.i. Colocalization with acidified compartments was similar among all the groups (Fig. 6). At 24h p.i. specialist strains showed lower MOI values compared to the other groups. Colocalization with acidified compartments was found lower for specialist and intermediate strains (Fig. 7).

M1 – 6 h p.i.

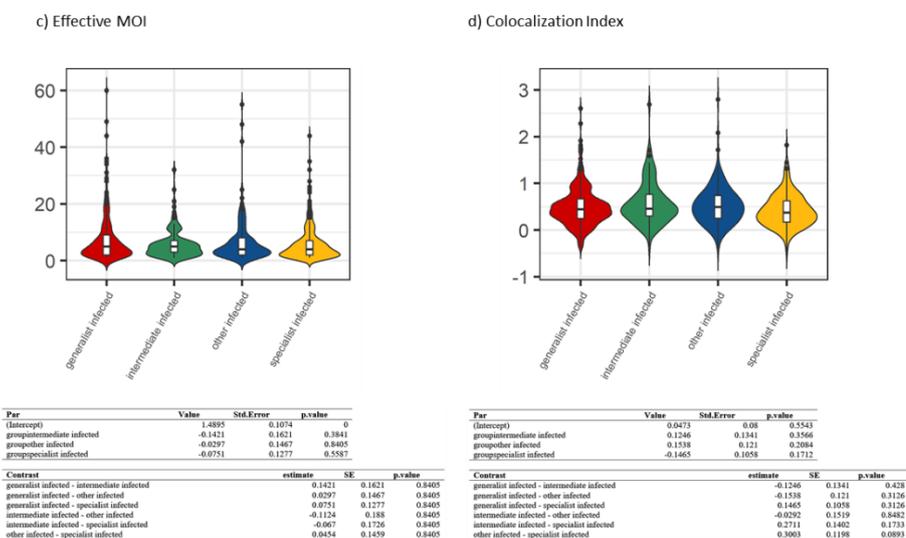


Fig. 6: MTB colocalization with acidified compartments at 6h p.i in M1 cells and statistical reports

M1 – 24 h p.i.

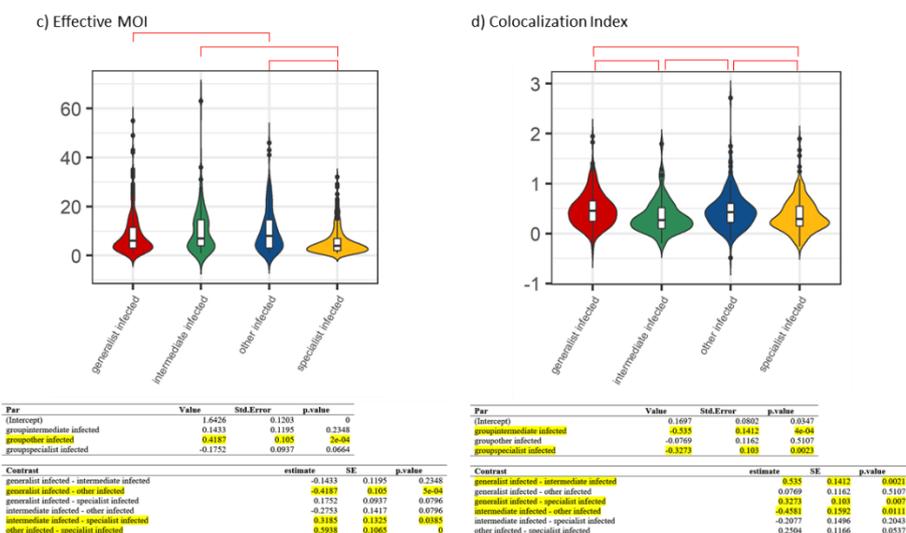


Fig. 7: MTB colocalization with acidified compartments at 24h p.i in M1 cells and statistical reports

In M2 macrophages, specialist strains showed lower MOI values compared to the other groups at 6 h p.i.. Colocalization with acidified compartments was found lower for specialist and intermediate strains (Fig. 8). No statistically relevant differences in effective MOI at 24 h p.i. were observed, even though generalist strains displayed a higher distribution of MOI values. Colocalization with acidified compartments was similar among all the groups (Fig. 9).

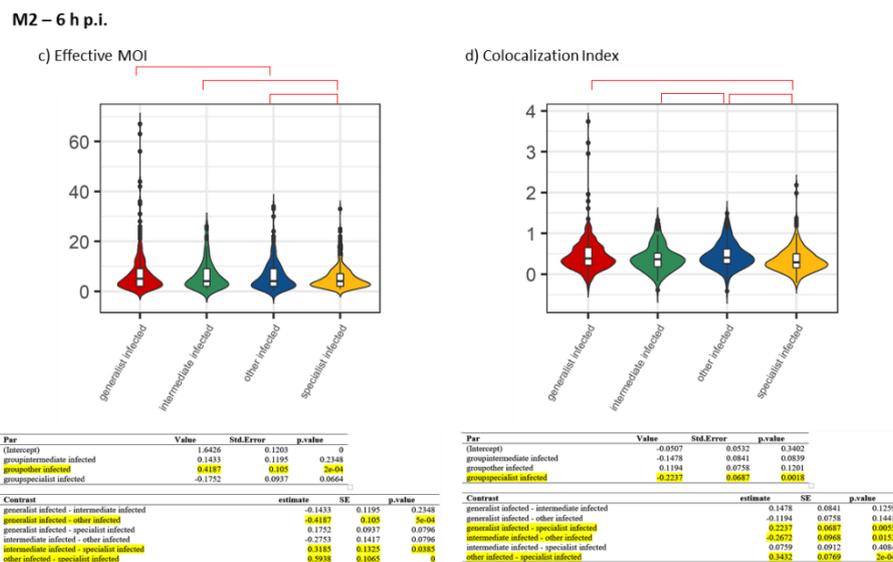


Fig. 8: MTB colocalization with acidified compartments at 6h p.i in M2 cells and statistical reports

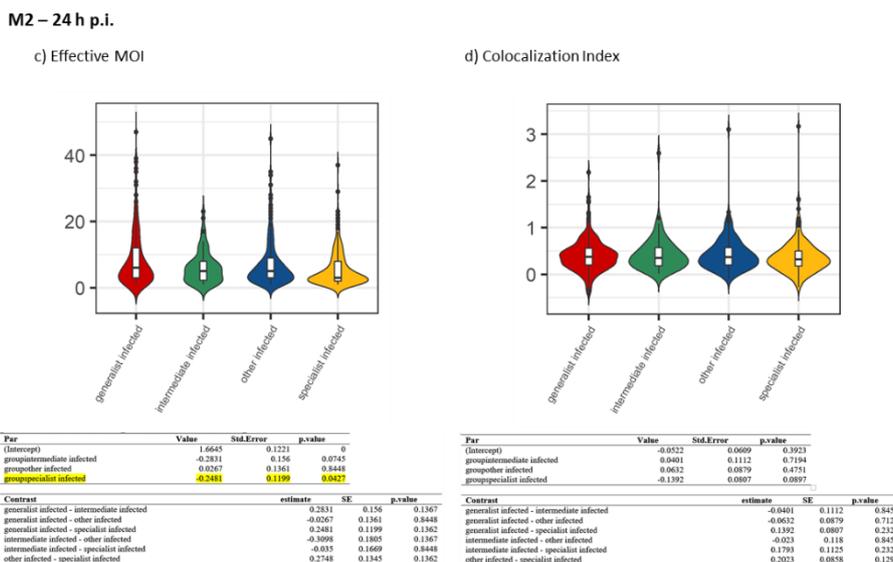


Fig. 9: MTB colocalization with acidified compartments at 24h p.i in M2 cells and statistical reports

No correlation between MOI and the colocalization index was found (Supplementary Phagolysosomal acidification).

- **Autophagic flux**

We evaluated the autophagic flux in THP-1 M1 and M2 macrophages infected with H37Rv, H37Ra, L6, and L2 at 24 h p.i. by the use of the CYTO-ID kit (Fig. 10). A total of 1960 single cells were considered for the analysis (on average, for each strain tested, M1: 64 not infected cells + 67 infected cells; M2: 60 not infected cells + 63 infected cells) (Supplementary Autophagy). The size and the number of autophagosomes were considered, and strains were grouped as previously reported in Table 1.

In M1 macrophages, only the group "other" showed some degree of autophagy; indeed, whereas also generalist and specialist groups are showing an increase in the number of autophagosomes, their size does not show statistically increased size, thus suggesting poor maturation and progression in the autophagic flux (Fig. 11).

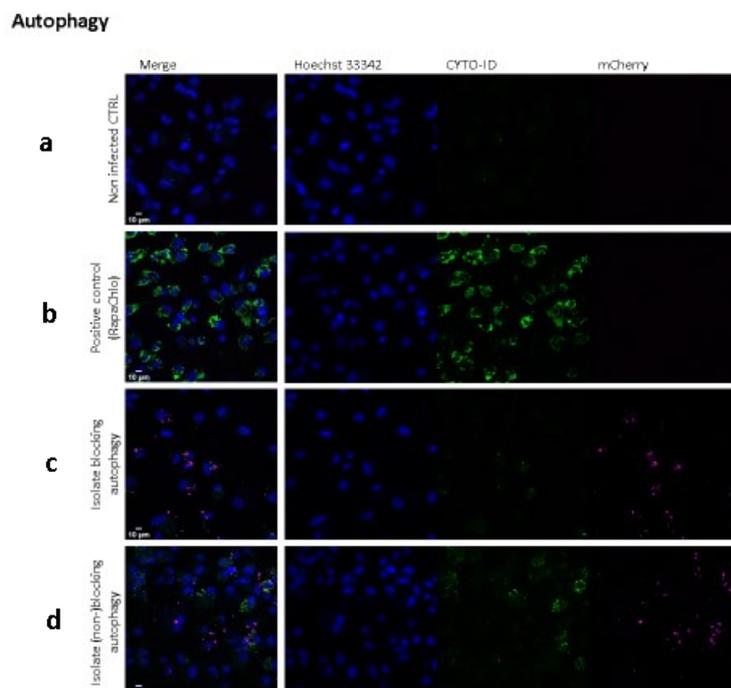


Fig. 10: THP-1 cells non-infected (a); treated with autophagy inducer (b); infected with autophagy-blocking bacteria (c); infected with autophagy non-blocking bacteria (d). The fluorescence images were acquired after staining with CYTO-ID (Green) and Hoechst (Blue). Bacteria are reported in red. The reported images are illustrative of the pipeline adopted

M1 – 24 h p.i.

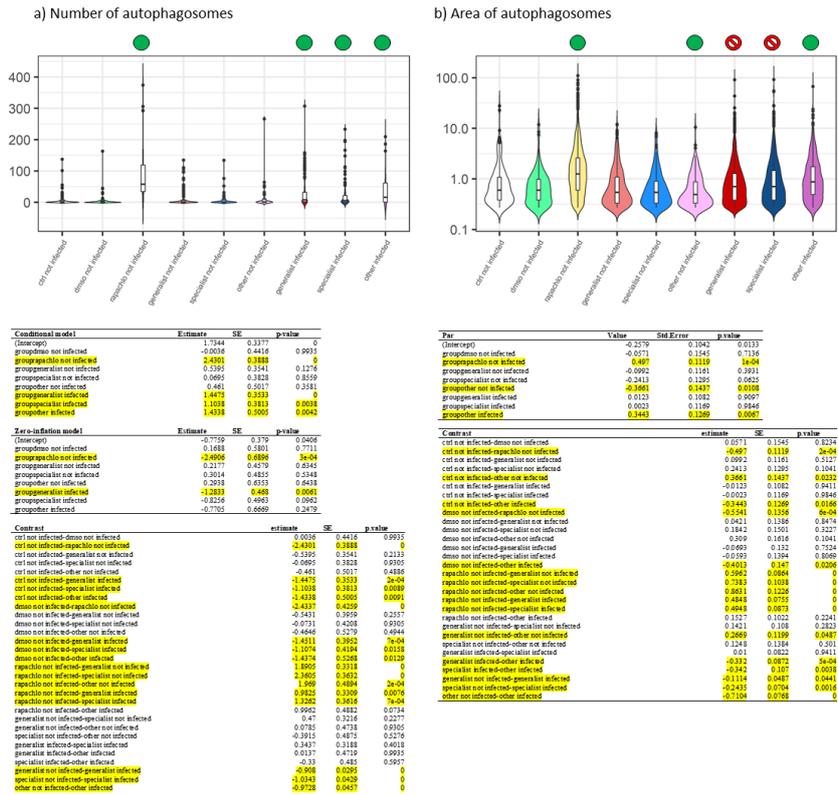


Fig. 11: Autophagic flux at 24h p.i. in M1 cells with statistical reports

In M2 macrophages, despite a certain increase in the number of autophagosomes, their size does not show statistically increased size, thus suggesting poor maturation and progression in the autophagic flux. None of the group is inducing autophagy (Fig. 12). No correlation between MOI and autophagy was found (Supplementary Autophagy).

M2 – 24 h p.i.

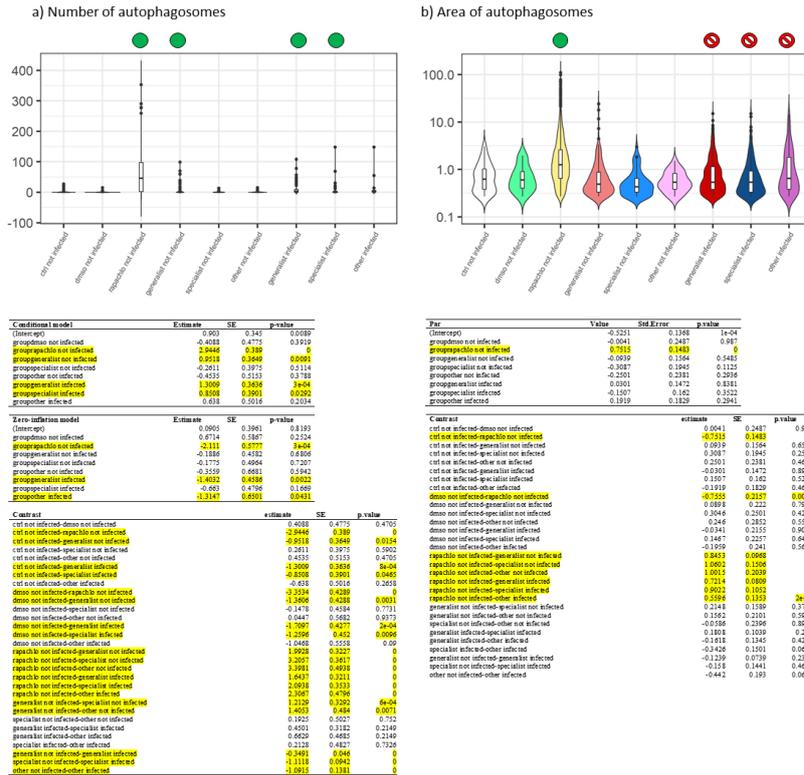


Fig. 12: Autophagic flux at 24h p.i. in M2 cells with statistical reports

• Apoptosis and necrosis

We evaluated the induction of apoptosis and necrosis THP-1 M1 and M2 macrophages infected with H37Rv, H37Ra, L6, and L2 at 24 h p.i. (Fig 13). A total of 2470 single cells were considered for the analysis (on average, for each strain tested, M1: 98 not infected cells + 75 infected cells; M2: 104 not infected cells + 79 infected cells) (Supplementary Apoptosis). Despite the marker for necrosis 7-AAD had a spectrum overlapping to one of mCherry mycobacteria (7-AAD Ex/Em 546/657; mCherry Ex/Em 587-610 nm) and both have been imaged using the 561 laser, the morphology of the bacteria and of the necrotic cells was completely different and easily distinguishable (Supplementary Apoptosis). However, in our testing conditions (24 h p.i.) necrosis was never observed, thus necrosis was not further considered.

In M1 macrophages, bystander non-infected cells of generalist and other strains showed increased apoptosis (40%). Despite not statistically

significant, also macrophages infected with generalist strains showed a similar increase in the percentage of apoptosis. Stratification of the data by mycobacterial burden showed higher apoptotic levels in bystander non-infected cells or in cells infected with lower mycobacterial load (Fig. 14).

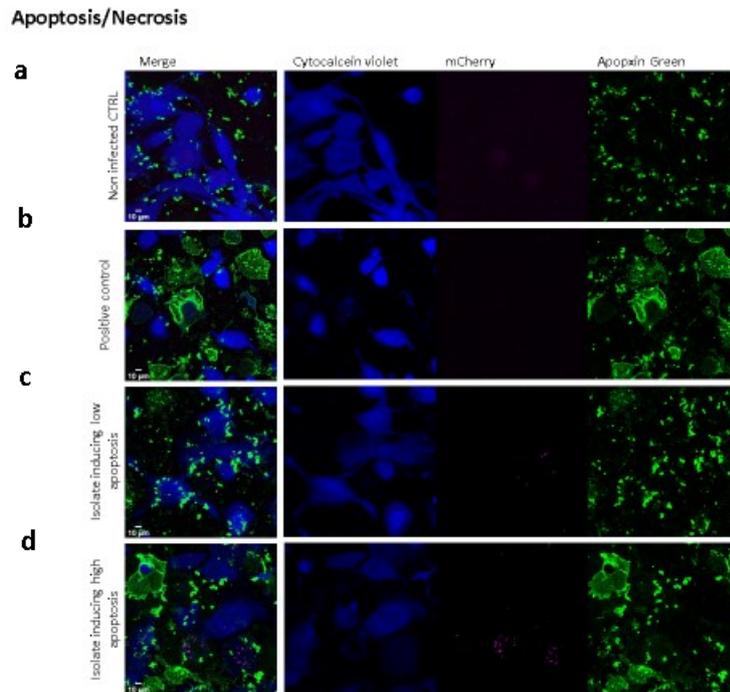


Fig. 13: THP-1 cells non-infected (a); treated with apoptosis inducer (b); infected with apoptosis-blocking bacteria (c); infected with apoptosis non-blocking bacteria (d). The fluorescence images were acquired after staining with CytoCalcein (Green) and Hoechst (Blue). Bacteria are reported in red. The reported images are illustrative of the pipeline adopted

M1 – 24 h p.i.

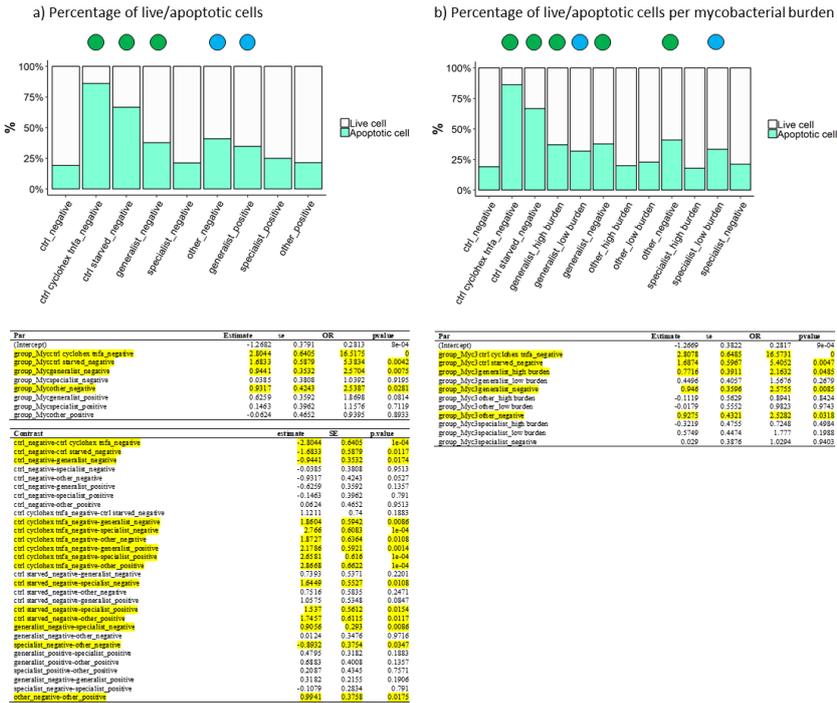


Fig. 14: Apoptosis levels at 24h p.i. in M1 cells with statistical reports

In M2 macrophages, only bystander non-infected cells of other strains showed a slight, statistically not significant increase in apoptosis (30%) (Fig. 15).

M2 – 24 h p.i.

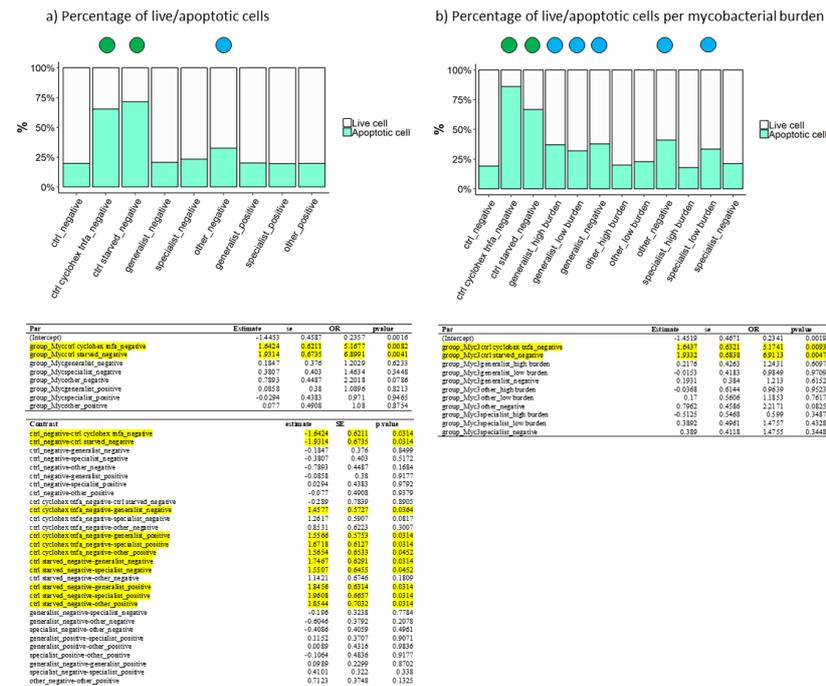


Fig. 15 Apoptosis levels at 24h p.i. in M2 cells with statistical reports

- **Cytokine production**

Cytokines induction analysis was performed on macrophages infected with H37Rv, H37Ra, L2, or L6 MTB strains. Uninfected controls were used as calibrators for analysis. For consistency with the previous analyses, strains grouped according to seven categories above-mentioned (Table 1), even though given the reduced number of strains tested some categories are in this case redundant. The first important result we obtained is about the M1/M2 cytokine-chemokine pattern production. In general, in M2 cells the infection with different MTB categories produces a more homogeneous cytokine pattern compared to infected M1 cells, where the differences among strains are more pronounced. Among the analytes considered, we found IL-1 β , IL-6, TNF α , IL-15 pro-inflammatory cytokines to be the most modulated by MTB infection, when compared to the uninfected control cells. In fact, with few exceptions (IL-1 β , IL-18) in M2 cells, different MTB categories do not show statistical differences. In M1 cells most of the differences between generalist and specialist strains, are found both in pro-inflammatory cytokines (IL-1 β , IL-15, IL-18), in anti-inflammatory cytokines (IL-1ra), and in growth factors (FGF, G-CSF). In general, laboratory strains were found to induce higher levels of pro-inflammatory cytokines compared to clinical isolates. Detailed statistical analysis is reported in Supplementary Cytokines.

Pro-inflammatory cytokines: in both M1 and M2 macrophages, specialist strains induced a significantly lower level of IL-1 β compared to generalist strains at all time points considered. Expression patterns of this cytokine are similar between M1 and M2 infected macrophages (Fig. 16). Main differences in IL-1 β production were found when comparing L6 and L2 infected cells at 4h p.i. (in M2 cells) and at 24h p.i. (M1 cells), where induction levels of L6 strains are similar to the non-infected control. In general, specialists induce a minor production of this cytokine in both cell types considered. IL-18 production is low, with similar production in M1 and M2 cells. Specialist strains induce a lower production of this cytokine

compared to generalist ones. IL-15 induction is statistically higher in M1 cells when infected with generalist strains.

No statistical differences were found comparing IL-6 production in specialists vs generalists strains, but IL-6 levels observed in M2 macrophages are lower than M1.

As for IL-6, TNF α shows no statistical differences comparing specialists vs generalists strains, and is mostly induced in laboratory strains in M2 cells.

IL-15, IFN γ , IL-18 are poorly released in response to MTB infection in both M1 and in M2 cells at the time points considered. IFN γ production is negligible at the time points considered with low production both in M1 and M2 infected cells. Immunofluorescence and single-cell analysis on TNF α and IFN γ showed similar findings (Supplementary Cytokine). We performed a correlation analysis between these cytokines levels and (i) bacillary load or (ii) phagolysosomal acidification; despite a few correlations could be found, they were most likely strain-specific rather than providing evidence for more general assumptions (Supplementary Cytokine). In all cytokine considered, both in M1 and M2 cells, laboratory strains induce a higher production of pro-inflammatory cytokine, compared to clinical ones.

Anti-inflammatory cytokines: most of the differences among MTB categories are found in M1 infected cells. IL-1ra is highly induced compared to the control, especially in generalist strains, which are statistically different compared to specialists in M1 cells at all time points considered (Fig. 16). In M2 cells, laboratory strains induce a high level of this cytokine at 24h p.i. IL-10 is slightly induced by MTB infection; Specialists vs Generalists show no differences in their expression pattern. Contrarily from pro-inflammatory cytokines, anti-inflammatory production, in laboratory strains infected cells is similar to the clinical ones. Statistical differences are found only in the IL-1ra secretion, in M2 cells at 24h p.i., and in the IL-10 production at 4h p.i. in M1 cells and at 24h p.i. in M2 cells.

Chemokines: lower differences are found in the chemokine measurements, with M1 and M2 infected cells that show similar production patterns (Fig. 17). No statistical differences are found in the induction of IL-8, MCP-1 and RANTES.

MIP-1 β production confirms what we observed in the immunofluorescence assay, where the level in M1 cells infected with generalist strains is statistically higher than specialist strains at 24h p.i.

Same confirm for MCP-1 production, who despite the high induction compared to the non infected control, show no statistical differences among categories considered.

Considering the differences among clinical and laboratory infected cells, once again laboratory strains induce a higher production of the selected chemokine (MIP-1 α/β , SDF-1 α , GRO α). Immunofluorescence and single-cell analysis on MCP-1 and MIP-1 β showed similar findings (Supplementary Cytokine).

Growth factors: similar results are found considering growth factor production. In general, specialist strains induce a lower secretion of those molecules, especially in M1 cells (Fig. 18). Major differences are found in the G-CSF and in the FGF basic production (M1 at 4 and 24h p.i, - M2 at 48h p.i.). Laboratory strains induce a higher induction of growth factors FGF basic, G-CSF, and PDGF-bb compared to clinical strains.

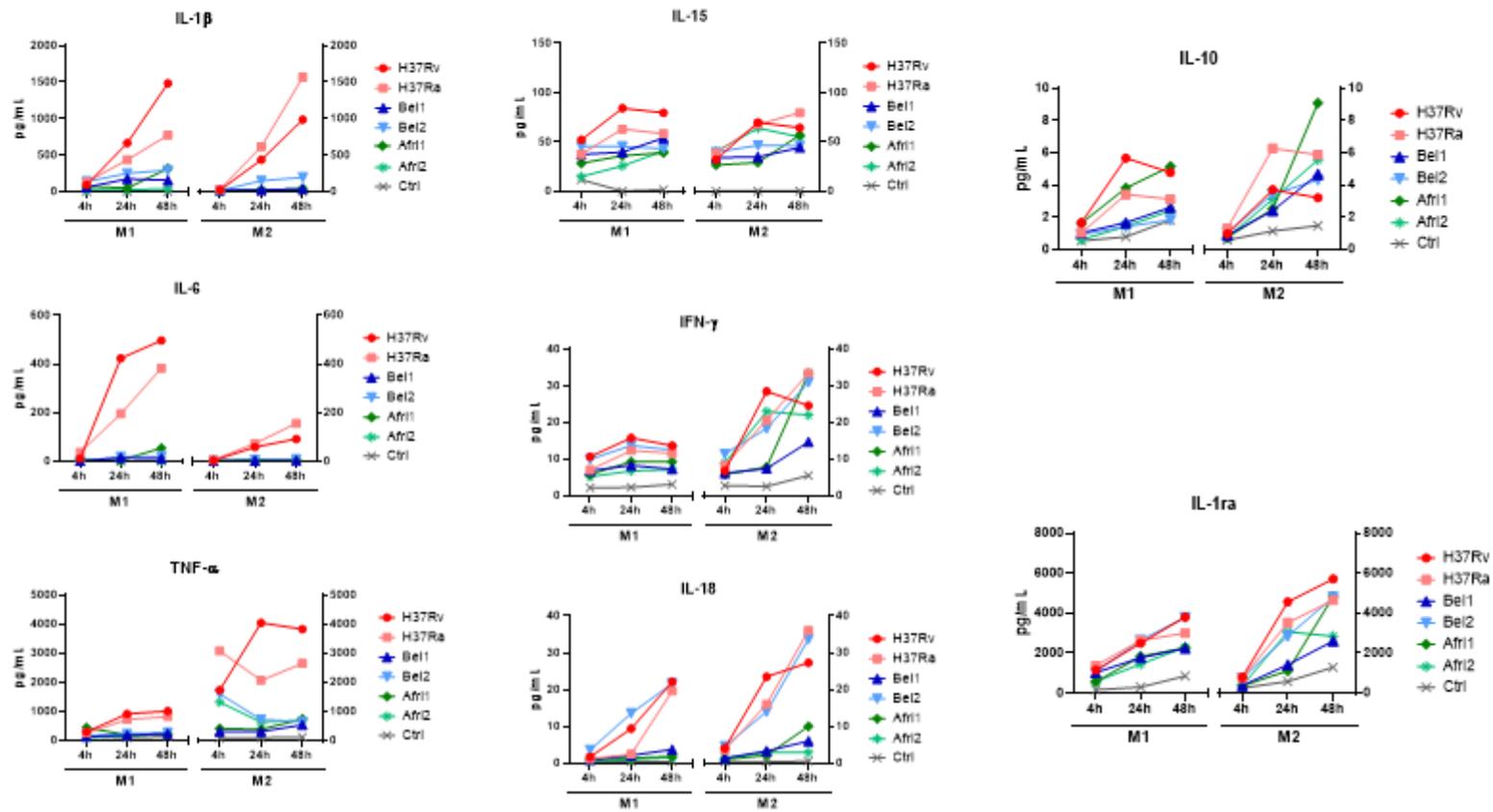


Fig. 16: Pro and anti-inflammatory cytokine results. MTB strains used: H37Rv, H37Ra, L2 (Bei1, Bei2), L6 (Afr1, Afr2)

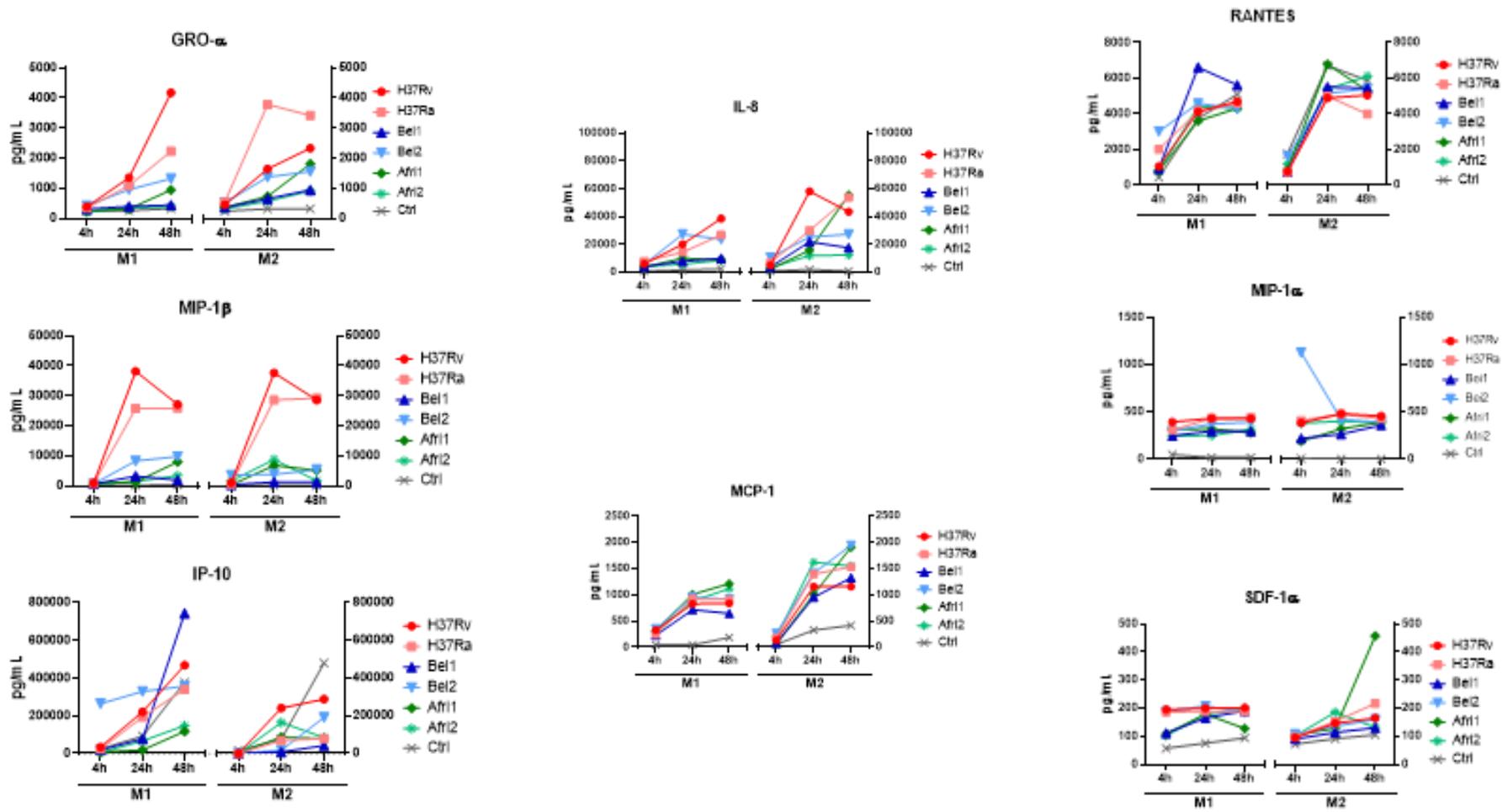


Fig. 17: Chemokine results. MTB strains used: H37Rv, H37Ra, L2 (Bei1, Bei2), L6 (Afri1, Afri2)

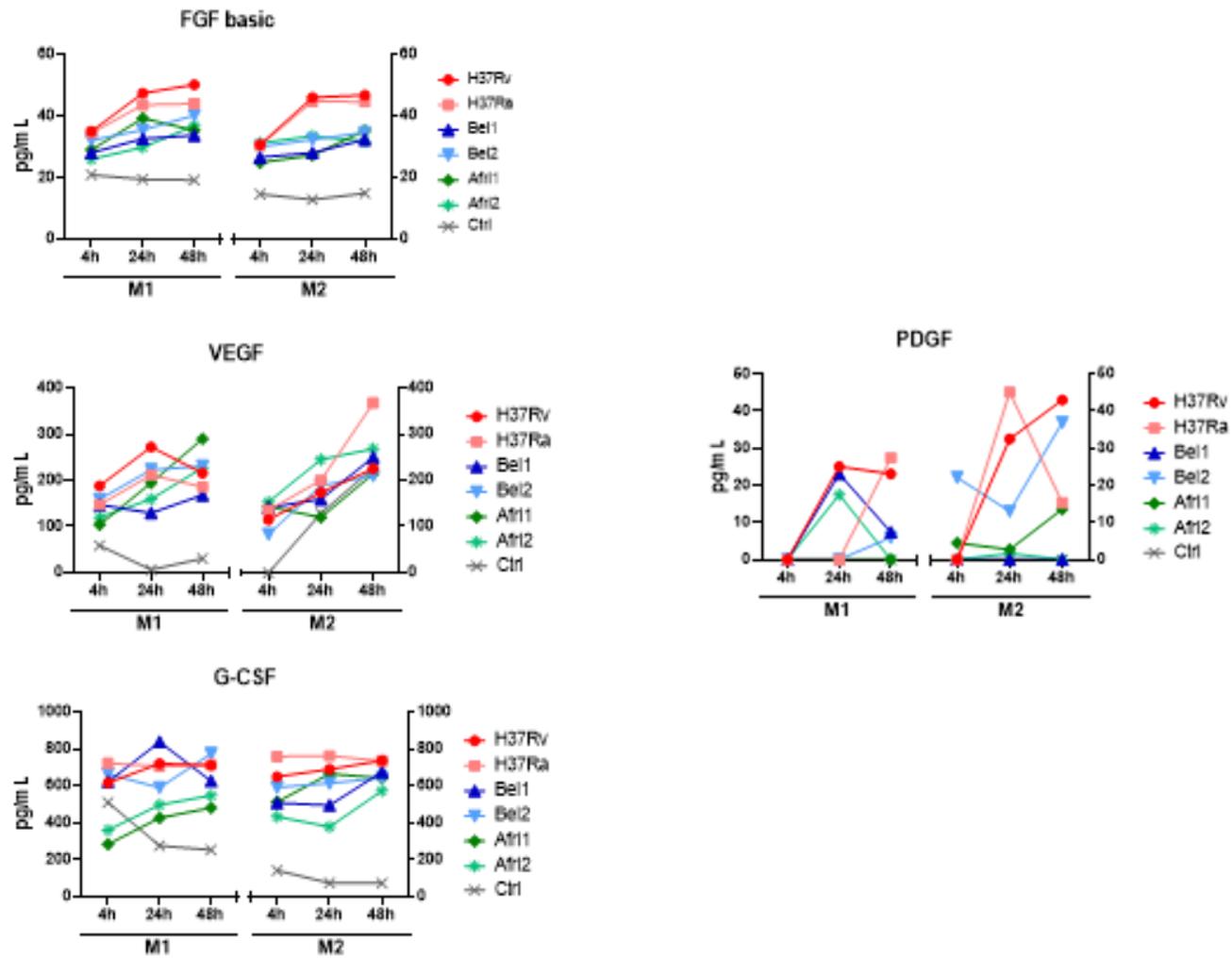


Fig. 18: Growth factors results. MTB strains used: H37Rv, H37Ra, L2 (Bei1, Bei2), L6 (Afri1, Afri2)

- **Cell survival**

We first evaluated the amount of healthy cells after the cytokine polarization of THP-1 (T0), before MTB infection. Results show how cytokine M1 induction with LPS causes a loss of about 20% of the THP-1 derived macrophages, while treatment with IL-4 (M2 polarization) causes the loss of about 11% of the cells. Whereas different MTB strains are associated with different levels of M1 macrophages death, M2 macrophages display less heterogeneity in survival after MTB challenge.

In M1 macrophages, at 48 h p.i. more virulent strains induce a reduction of live cells of about 50% compared to the 20% induced less virulent strains. As the infection progresses, at 5 days p.i. we could no longer appreciate differences among the categories considered.

In M2 macrophages, at 48h p.i. the level of cell death (about 20%) is lower compared to M1 cells, with no statistical differences among categories. This result is in line with apoptosis levels found for those categories. At 5 days p.i. MTB infection causes an increased M2 cell death (about 60%), compared to M1(about 40%). More virulent/generalist strains induce higher cell loss compared to less virulent/specialist ones (Fig. 19).

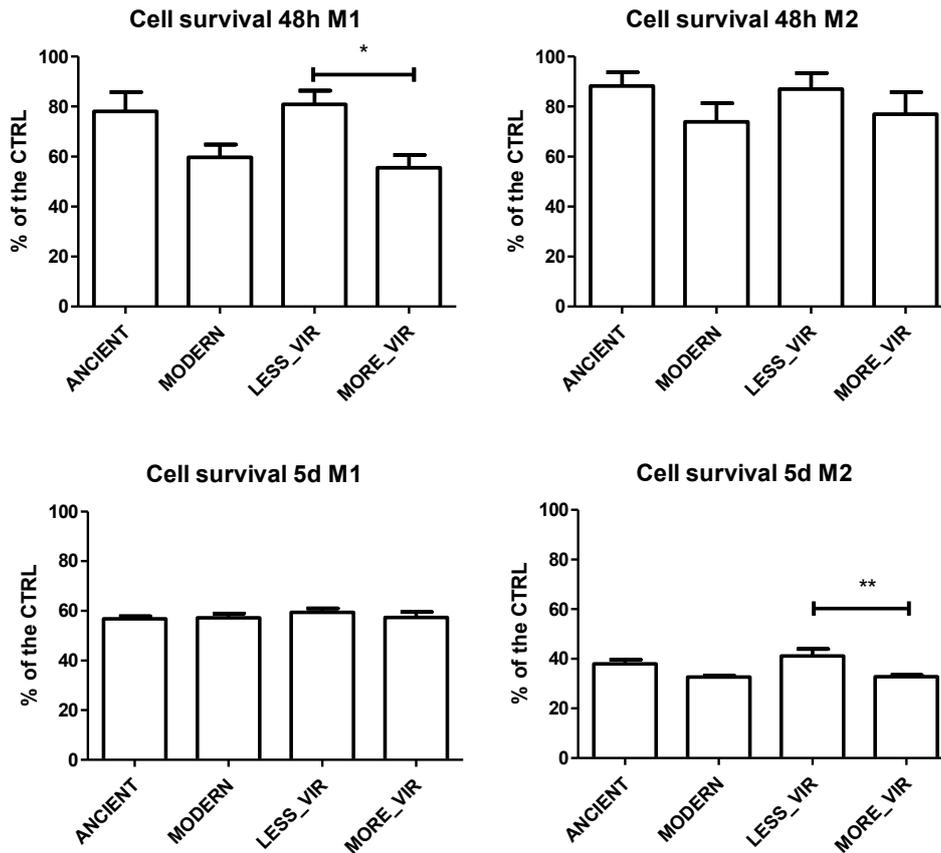


Fig. 19: Cell survival at 48h and 5days post-infection.

- **CFU**

CFU counts were performed on the selected MTB strains to study both the macrophagic entry (CFU at 4h p.i.) and the survival capacity (CFU at 48h p.i.). At 4h p.i. data were normalized on the CFU of the inoculum. The number of bacteria that effectively are phagocytized by the macrophages is below 10% of the inoculum in all the MTB strains selected. Differences are present considering M1/M2 cell phenotype: entry of bacteria is increased in M2 cells in all MTB strains selected. Higher CFU levels are found in H37Ra infected cells, while H37Rv and L6 show a similar CFU level. L2 uptake resulted to be very low compared to all other strains, with CFU values close to 1% of the inoculum in M1 cells and to 3% in M2 cells. At 48h p.i. CFU were normalized on the CFU at 4h p.i. (representing the number of bacteria actually internalized by the macrophages). H37Rv and H37Ra show

similar CFU counts, while clinical strains appear to grow more in infected macrophages. L2 strains show a higher number of CFU, despite the initial lower uptake, with larger growth in M1 cells (Fig. 20).

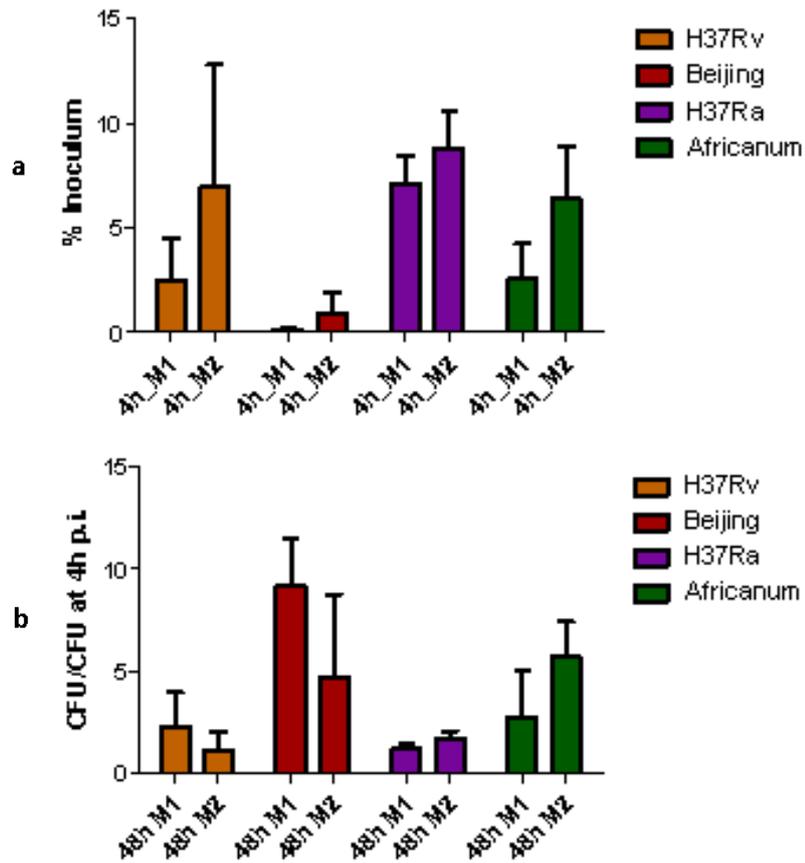


Fig. 20: a) CFU results at 4h p.i. (% of the inoculum) and b) at 48h p.i. (normalized on CFU at 4h) Beijing: Generalist; Africanum: Specialist

Discussion

How MTB survives within the macrophages is a key point in the pathogenesis and virulence of MTB. Early views of pathogenicity and virulence were pathogen-centered and were based on the assumption that these characteristics were intrinsic properties of bacteria, focusing mainly on the capacity of some microbes to cause disease in healthy hosts³⁸. The definition of bacterial virulence has changed over time, and it's now considered as a relative capacity to cause damage, both at cellular or at organ/tissue level. In this view, virulence is a property of the pathogen, but it is modulated by host susceptibility and resistance^{39,40}.

Despite their central role in MTBC infection, the phenotype of the macrophages is an underestimated component of *in vitro/ex vivo* infection models. Alveolar macrophages have specifically evolved for the alveolar environment where they exert their role against invasive organisms but also prevent inflammation and tissue damage. Alveolar macrophages exhibit an intermediate phenotype between pro-inflammatory, classically activated (M1), and anti-inflammatory, alternatively activated (M2) macrophages, and express both M1 and M2 markers⁴¹. As a point of note, despite the presence of both M1/M2 marker sets, they are skewed in favor of the M2 phenotype, with high CD206 and low toll-like receptors⁴². Alveolar macrophages generate a non-canonical NRF2-driven transcriptional response to Mycobacterium TB, which was recently demonstrated in an *in-vivo* mouse model. NRF-2 has been associated with anti-inflammatory responses, and it may create an M2-like phenotype that inhibits IL-1 and IL-6 production⁴³.

On the other end, the genetic diversity of MTBC has been increasingly recognized as another important player in determining the fate of the host-pathogen interaction⁴⁴. The interaction between immune cells and pathogens is complex, and at the single-cell level, their dynamics culminate in heterogeneous outcomes of infection.

In this work, we studied early host-pathogen interaction between MTB and human macrophages, taking into consideration both host macrophage phenotypic variability (resembling alveolar macrophages plasticity) and MTB genetic diversity considering major phylogenetical differences. In particular, we focused our attention on phagolysosomal acidification, autophagy, apoptosis. Cytokines induction, macrophage and bacterial survival were also considered. In terms of differentiation, functions, and reactions to external stimuli, THP-1 generated macrophages have been described to behave similarly to human macrophages ⁴⁵. The adoption of a single-cell analytical approach allowed to better appreciate the heterogeneity of the host-pathogen interaction for some of the above-mentioned analyses.

Despite previous studies, focusing mainly on the laboratory strain *M. tuberculosis* H37Rv, reported that mycobacteria arrest the maturation of their phagosomes and avoid further acidification of the bactericidal compartment, there is evidence that MTB can reside in acidic phagosomes (thus surviving low pH stress) ⁴⁶. The biogenesis of mycobacterial phagosomes is a complex and dynamic process. As a result, various features such as vacuolar proton-ATPase exclusion, synthesis of antacid compounds, selective prevention of fusion with active lysosomes, interference with phagosome integrity and repair, and acidic resistance all contribute to the MTB escape from the immune system mechanisms ^{47–49}. Concerning phagosomal acidification, in M1 macrophages acidification is delayed by more virulent, modern, generalist isolates, whereas at earlier time points less virulent, ancient, specialist strains are not blocking acidification. At later time points, none of the strains is blocking phagosomal acidification, but less virulent, ancient, specialist strains displayed lower levels of acidification compared to more virulent, modern, generalist/intermediate strains. In addition, more virulent, modern, generalist/intermediate isolates displayed higher colocalization with

LysoSensor positive regions, thus suggesting higher colocalization with regions at lower pH.

Macrophages classically activated prior to ingestion of mycobacteria are able to acidify the phagosomal compartments^{50, 51}. The higher level of acidification together with the higher colocalization of more virulent, modern, generalist isolates with acidified compartments suggest they are better at tolerating the acidic environment. In support of this view, L2 strains constitutively overexpress the DosR regulon, a regulon playing a pivotal role in protecting mycobacteria from acidification during intramacrophagic infection^{52 - 54}. Similarly, L2 strains harbor a lineage-specific mutation in KefB (*Rv3236c* gene), a potassium/proton antiporter relevant for arresting phagosomal acidification^{54, 55}. Diversely, isolates belonging L5 and L6 (*M. africanum*) harbor mutations negatively affecting the two-component virulence regulatory system PhoP/PhoR, a regulon involved in the adaptive response to acidic pH⁵⁷. Recently, evidences correlate the levels of sulfolipid-1 (SL-1) and lysosomal delivery: strains showing lower SL-1 production display a reduction in the delivery to lysosomes⁵⁸. Interestingly, L5 and L6 lineages have been reported an impaired SL-1 production likely due to the mutations affecting the PhoP/PhoR regulatory system whereas L1 possesses a lineage-specific mutation Ala43pro in the *papA2* gene abrogating cell wall SL-1^{59, 60}. Despite recent findings that found no statistically significant differences in survival of modern and L1 isolates in acidic pH in vitro, the transcriptional regulatory network of ancient and modern lineages is obviously distinct, with crucial implications for stress response⁶¹.

The picture observed in alternatively activated macrophages (M2) was different. Acidification is observed at early time points with clinical isolates, but not with laboratory strains. However, at 24 h p.i. more virulent, modern, generalist/intermediate strains blocked acidification, whereas less virulent, ancient, specialist strains were less efficient in blocking the acidification of phagolysosomes. Contrary to what observed in M1 macrophages, in alternatively activated macrophages no differences in

terms of colocalization of mycobacteria within acidified compartments were observed. The acidification level and the number of acidified organelles were generally lower compared to classically activated macrophages. Similarly, the Colocalization Index was also generally lower in M2 macrophages.

These macrophages are able to mount a weak response barely able to counteract less virulent, ancient, specialist strains at the time points considered, while more virulent, modern, generalist/intermediate strains quickly hijack macrophage antimicrobial defenses. The acidification of phagosomes depends on the organism phagocytosed and the specific receptor starting the signaling pathway. The mechanisms besides this sharper difference in coping with phagosomal acidification observed only in M2 macrophages require further elucidation; however, we noticed that ancient, animal, specialist strains harbor a mutation (Thr171Ala) in the phosphatidylinositol 3-phosphate phosphatase SapM (*Rv3310* gene), an enzyme playing a crucial role in blocking phagosome maturation in M2 and unpolarized macrophages only ⁶². It's possible that this mutation, in combination with the genetic background, explains why ancient isolates are less effective at preventing phagosome acidification in alternatively activated macrophages.

The differences in response timing between M1 and M2 macrophages are consistent with prior findings. Indeed, when phagosomes from activated or M1 macrophages were compared to those from resting or M2 macrophages, they displayed a delayed acidification kinetics ⁶³⁻⁶⁵.

To be noted that laboratory strains showed higher capacity to block phagolysosomal acidification compared to clinical isolates. This highlights the fact that infection models based only on reference laboratory strains are not always able to completely recapitulate the pathogenesis of tuberculosis. Irrespectively from the macrophagic phenotype, ancient, specialist isolates showed lower bacillary load compared to modern, generalist isolates. At early time points p.i. there was an increase in the number of M1 macrophages showing phagosomal acidification in presence

of less virulent, ancient, specialist strains; however, after this initial response, the number of such cells was reduced. More virulent, modern, generalist isolates did not cause an increase in the number of responsive cells. Overall, at 24 h p.i. only a relatively small amount of M1 macrophages challenged with mycobacteria (15-25%) displayed phagosomal acidification. Limited bystander effect in non-infected macrophages was observed. Similarly to classically activated macrophages, at early time points p.i. there was an increase in the number of M2 macrophages showing phagosomal acidification, but after 24 h only animal lineages and laboratory/non-causing strains maintained a high percentage of “acidified” cells, whereas either less virulent, ancient, specialist and more virulent, modern, generalist isolates displayed low or very low number of responsive cells (<10%). In M2 macrophages a consistent bystander effect in non-infected cells was observed.

The use of a single-cell analytical approach highlighted the heterogeneity of the macrophagic response to *M. tuberculosis* infection. Whereas a general increase in the phagosomal acidification could be observed, our data suggest this increment is mainly due to a relatively small proportion of cells rather than being a population-average response (at least when considering macrophages infected with either less virulent, ancient, specialist and more virulent, modern, generalist isolates). Furthermore, the differences in acidification time could be also due to fluctuations in the levels of *M. tuberculosis* damaging (virulence) factors as already hypothesized⁶⁶.

A bystander effect on non-infected macrophages was previously observed mainly during studies on apoptosis, and this was found to be induced by less virulent strains⁶⁷⁻⁷⁰. For the first time, we report a similar effect in terms of acidification. Mycobacterial protein and lipid antigens or exosomes derived from infected cells can reach non-infected bystander cells, a phenomenon modulating the response of immune cells and reported to affect subsequent IFN γ activation of macrophages⁷¹. Notably, the bystander increase of acidification was found in M2 macrophages and

was mainly driven by less virulent, ancient, specialist strains MTB is able to evade macrophagic clearance due to autophagy⁷². Our data did not show relevant differences among less virulent, ancient, specialist, and more virulent, modern, generalist strains as either the categories were able to block the formation of autophagosomes. Previous findings showed similar results, despite highlighting a difference in the mechanism used by different genotypes to hinder autophagy^{72,73}.

Other parameters we evaluated were induction of apoptosis and necrosis. At the time point considered (24 h p.i.) we could not observe necrosis, thus we limited our analysis to apoptosis. Apoptosis has been considered to have a debatable role with groups supporting a protective effect for the host and contributing to infection control, whereas others suggest a detrimental effect increasing mycobacterial dissemination⁷⁴⁻⁷⁷. Apoptosis was observed only in bystander non-infected cells of more virulent, modern and generalist isolates. Induction of apoptosis by virulent strains seems contradictory to its protective role against MTB. However, previous findings highlighted that apoptosis signals at day 1 p.i. progressed to necrosis at later timepoints; furthermore, exposure of phosphatidylserine is observed in different forms of programmed cell death (namely apoptosis, necroptosis and ferroptosis)⁷⁸. Apoptosis data are in line with what we observed with the cell survival assay, where in M1 macrophages, more virulent strains induce a major cells death rate compared to less virulent strains, while in M2 cells we found lower induction of apoptosis, with no statistical differences among MTB categories.

Data on human and mice models show how the principal protection against TB depends on the activation of the Th1 response, and the subsequent production of pro-inflammatory cytokines such as IFN γ and TNF α , which are involved in the granuloma formation. The role of macrophages in phagocytosing bacteria and presenting antigens is crucial for the T cell priming and in controlling MTB infection. Because their

expression often links with efficient immune responses, type-1 inflammatory cytokines are critical in the protection against MTB, and genetic deficiencies of these factors lead to improved TB susceptibility. MTB infection can drive macrophages toward M1 in the early phase, and shift their status into M2 phenotypes in intermediate and late phases. This strategy is thought to be put in place for an attempt by the host to halter the pathophysiology caused by MTB or a bacterial strategy to shield from immune attack⁷⁹⁻⁸¹.

In our study we decided to focus our attention on the cytokine/chemokine production at the early phases of the interaction between MTB different strains and human macrophages.

Cytokine, chemokine and growth factors selected in this study are the main produced elements involved in the macrophagic early response to MTB infection. In general, MTB appears to strongly modulate the production of pro-inflammatory cytokines and to induce an M1 shift in the pre-activated M2 cells, increasing pro-inflammatory production. The interleukin 1 and type 1 interferon (type 1 IFN) signaling pathways are well studied in mice but poorly understood in human models. They have been shown to play opposing roles in tuberculosis. Type-1 IFNs contribute to pathogenesis through weakening of host resistance to MTB while IL-1 β is required for host control of infection. We found IL-1 β production highly increased in time, especially in the laboratory strains H37Rv and H37Ra⁸². Differences are also found between L2 and L6 strains, with the latter that induce the lowest IL-1 β production. Those results are in line with the results coming from the single cell level experiment, where virulent strains induce a higher delay in the acidification of M1 cells at early timepoints, while in M2 we saw a blocking of the acidification at later timepoints (unlike less virulent strains). Of notice, the induction of TNF α and IFN γ appears to be higher in M2 cells, showing a more rapid shift to M1 polarization, particularly in the laboratory strains. It is important to remark that in literature it is reported that cell death caused by MTB strains, is non-directly correlated with IL-1 β secretion⁸³.

IL-6 has been associated to host protection by the expression of early immune response, and low levels of IL-6 are associated with lung lesions and cavitary TB. Our data show that M2 cells are less capable of expressing IL-6 following MTB infection in all strains considered, thus suggesting a lower capacity to inhibit damages caused by MTB. In M1 cells, clinical strains are able to retard IL-6 production, thus increasing cellular damage, compared to the laboratory strains⁸⁴. IL-18 is a potent immunoregulatory cytokine with different immunological properties. It regulates the mechanisms of both innate and adaptive immunity and plays a key role in host defense, mostly regulating IFN γ release. In our study, we observed poor secretion of this cytokine, indicating immune suppression of the infected cells from MTB strains. Once again, higher levels are found in laboratory strains and in one of the two L6 strains selected. IL-15 secretion is slowed by MTB infection since the amount measured in both M1 and M2 cells is low. IL-15 participate to the activation and proliferation of T and natural killer (NK) cells. High concentrations of this cytokine favor TNF α , IL-1, and IL-6 production, whereas very low concentrations favor IL-10 production, thus increasing MTB survival. All strains considered inhibit the IL-15 production, with statistical differences among the categories selected at early timepoints.

Concerning anti-inflammatory cytokines, IL-10 production is suppressed by MTB infection, even in the M2 cells, confirming an M1 shift, while the IL-1ra, antagonist of IL-1 β , is highly produced. The induction of IL-1ra is reported to play an important role in the establishment of the infection. Low levels of this cytokine are correlated with better survival in MTB-infected mice.

The expression of chemokine and growth factors in MTB infection plays important role in the recruitment of other immunity cells, implicated in the formation of granuloma and the containment of TB. Expression of those factors has been seen after 12 days post-infection in mice and is not modulated after the first contact with the pathogen⁸⁵. In our study, most of the chemokines selected are not modulated by MTB at the time point

considered and show low differences among MTB categories selected, while growth factors (VEGF, G-CSF, FGF) resulted to show differences in M1 cells.

Another expected, but important result we obtained, is about the differences we found in the cytokine production of cells infected with clinical or laboratory strains. Those strains appear to increase pro-inflammatory cytokines, far more than clinical strains, which seem to delay the immune system reducing the inflammatory response.

Finally, we also considered the strains' capacity to replicate in the host and kill macrophages as an extreme measure of virulence (intended as "capacity to cause damage" as previously defined). Our data show how there is not a direct correlation between cytokine induction and the killing capacity of MTB strains. Indeed, despite low differences in pro-inflammatory signalling secretion between ancient/specialist/less virulent and modern/generalist/more virulent strains, the latter are more able to induce cell death and to replicate within macrophages. L2 strains showed a lower macrophagic uptake, but an increased capacity to replicate in macrophages and to kill the host ⁸⁶. These findings are confirmed by literature data. Our single-cell based study further contribute to show how important are both the intrinsic virulence capacity of MTB strains and the activation status of the receiving host. In M1 macrophages, higher phagolysosomal acidification and colocalization of modern/generalist/more virulent strains would be expected to correlate with improved infection control, whereas the outcome is, as mentioned, lower macrophagic survival. The strategy adopted by MTB in M2 macrophages is somehow completely different, with modern/generalist/more virulent strains escaping acidified compartments. Our study further contribute to better understand the role of the genetic diversity of MTBC strains in the pathogenesis of TB and in relationship with defined host conditions. Future studies should be directed toward the identification of individual factors responsible for lineage-specific behaviours.

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Chapter 3

ncRv0842c*, a smallRNA regulating the efflux pump *Rv0842* involved in rifampicin tolerance in *Mycobacterium tuberculosis

Abstract

Regulatory pathways mediated by smallRNAs (sRNAs) in *Mycobacterium tuberculosis* (MTB) and their role in stress response are still poorly understood. We identified the smallRNA *ncRv0842c* cis-encoded to the *Rv0842* gene, a putative efflux pump putatively involved in response to rifampicin (RIF). The aim of our study was to characterize the role of *ncRv0842c* during RIF challenge in different MTB lineages.

First, we looked at lineage-specific mutations mapping in smallRNAs. We then selected *ncRv0842c* for further characterization. qPCR was performed on different MTB strains belonging to ancient (L1, L5) and modern (L2, L4) lineages to assess the expression of *ncRv0842c* and *Rv0842* in both basal and RIF-induced stress. H37Rv strain was used as a reference. Overexpression of *ncRv0842c* was achieved by the use of the pMV261 plasmid as a vector. RIF Minimum Inhibitory Concentration (MIC) was determined by microplate AlamarBlue assay (MABA). The effect of *ncRv0842c* overexpression was compared to the one obtained by the use of carbonyl cyanide 3-chlorophenylhydrazone (CCCP).

WGS analysis revealed a synonymous mutation in *Rv0842* (L45L) specific for ancient lineages abrogating the -10 promoter region of the cis-encoded smallRNA *ncRv0842c*. Basal expression analysis showed a strong down-regulation of the smallRNA *ncRv0842c* in L5 compared to H37Rv. During RIF challenge, the efflux pump was up-regulated whereas the sRNA was down-regulated in H37Rv, while the smallRNA expression was consistently not affected by RIF stress in L5. MABA assay performed on the *ncRv0842c* overexpressing mutant in H37Rv and L4 clinical isolates showed a 1-dilution reduction of the RIF MIC in comparison with their own control strain

(empty pMV261). Similarly, a 1-dilution reduction of the MIC was observed when control strains were treated with CCCP. The mutant strains belonging to L1 and L5 did not show MIC changes for RIF despite the overexpression of the smallRNA.

ncRv0842c is a negative regulator of the efflux pump *Rv0842* and is downregulated during RIF stress. Our data showed that synonymous mutations may have an important effect on regulatory pathways. In addition, the characterization of lineage-specific mutations involving the smallRNA post-transcriptional regulatory network would provide a better understanding of stress response in MTB.

Introduction

Tuberculosis, caused by *M. tuberculosis* (MTB) complex species, globally is the top infectious cause of death after COVID-19; an estimated 9.9 million people developed and 1.5 million died from the disease in 2020. Drug resistance poses a serious threat to ongoing efforts in controlling the epidemic.

The World Health Organization (WHO) framework for ending the global TB epidemic relies in part on the early identification of drug-resistant cases in order to implement timely and appropriate treatment and infection control strategies. Global progress has been made in the rapid detection of drug resistance as a result of improvement of genotype-phenotype association studies and broad implementation of WHO-endorsed assays ¹. However, despite the identification of genetic markers with high predictive values for the detection of drug resistance, our capacity to diagnose resistant cases for many drugs is below the target-product-profiles described by the WHO to support the End TB Strategy ². One underlining concept behind this diagnostic gap is likely the fact that whereas the main drivers of resistance are often only a few key mutations, a larger proportion of mutations contribute to resistance by their cumulative (especially when non-essential genes are involved in the mechanisms of drug resistance) and or regulatory effects.

Another relevant topic drawing attention in the field is related to better understanding the role of genetic diversity within the MTB complex in both intrinsic and acquired drug resistance. This complexity has not always been considered in the past.

Like other pathogenic bacteria, MTB has to adapt to rapidly changing conditions and thus has evolved a plethora of sensory systems that activate or repress the expression of genes in response to shifting environmental and host signals ³. In particular, small regulatory RNAs (smallRNAs) have recently emerged as major regulators of adaptive responses and regulate essential pathways in bacteria to respond quickly to their environment ⁴.

Despite an increasing number of smallRNAs have been identified in MTB, their potential role in drug resistance has not been explored, and much more attention has been paid to their potential role in virulence and stress response ⁵.

We propose an improved approach to achieve a better understanding of MTB drug resistance focusing on smallRNAs and keeping into account the genetic background of the strains.

Methods:

- **MTB strains**

For initial assessment 20 modern MTB (eight L4 strains, nine L2, four L3) and two ancient MTB strains (L5) were selected in order to study the baseline expression of the candidate_1080 (renamed *ncRv0842c*) smallRNA both in exponential and stationary growth phases. Strains selected included MDR, non-MDR, clade-related and sporadic isolates.

For further characterization of *ncRv0842c* nine phylogenetically modern clinical strains (L3 and L4) were selected from our laboratory collection, according to the RIF (RIF) DST results (four RIF-sensible strains and five with Increased MIC - iM strains, with RIF-MIC close to the RIF breakpoint). As control, the laboratory strain H37Rv (NTCC7416) and two ancient MTB lineage-reference RIF -sensible strains (L1 and L5), selected from ITM (Anversa) collection, were included in the study (Table 1) ⁶. In addition, strains were checked for kanamycin susceptibility to allow the overexpression of the smallRNA *ncRv0842c* by the use of the pMV261 plasmid harboring a kanamycin resistance cassette.

Table 1: MTB strain selected for this study, and their RIF MIC ($\mu\text{g}/\text{mL}$)

Strain (<i>rpoB</i> mutation)	Lineage	MABA MIC	RIF DST
<i>BG 3009</i>	L4	0.03 $\mu\text{g}/\text{mL}$	S
<i>CCM123</i>	L4	0.06 $\mu\text{g}/\text{mL}$	S
<i>CCM321</i>	L4	0.06 $\mu\text{g}/\text{mL}$	S
<i>CCM190</i>	L4	0.06 $\mu\text{g}/\text{mL}$	S
<i>R246 (Asp453Tyr; Thr444Ser)</i>	L4	0.5 $\mu\text{g}/\text{mL}$	iM
<i>R399 (Leu430Pro)</i>	L3	0.25 $\mu\text{g}/\text{mL}$	iM
<i>R453 (Leu452Pro)</i>	L3	0.125 $\mu\text{g}/\text{mL}$	iM
<i>R749 (Leu452Pro)</i>	L3	0.25 $\mu\text{g}/\text{mL}$	iM
<i>R743 (Leu450Pro)</i>	L3	0.25 $\mu\text{g}/\text{mL}$	iM
<i>L1-082</i>	L1	0.03 $\mu\text{g}/\text{mL}$	S
<i>L5-097</i>	L5	0.015 $\mu\text{g}/\text{mL}$	S

For mutant strains overexpressing the smallRNA, competent cells were prepared by washing three times a well-growth bacterial suspension in sterilized ice-cold PBS with 10% glycerol. When required, 40 μl of competent cells were incubated in ice with 2.5 μg of pMV261 (MOCK control) or pMV*ncRv0842c* for 1 minute; then, the suspension was moved into a 0.2 cm electroporation cuvette and electroporated with the MicroPulser (Bio-Rad, California, USA) apparatus at 2.5 kV, 25 μF , and 200 Ω . Electroporated bacteria were collected from the cuvette, diluted with 1 ml of new complete 7H9 media without antibiotics and let recovered for 24 h before being plated on 7H10 agar plates complemented with kanamycin (50 $\mu\text{g}/\text{ml}$) to obtain transformed colonies. After 21 days, at least 3 colonies for each plate were selected, checked for the presence of the plasmids by performing end-point PCR. Transformed colonies were grown in the presence of kanamycin to allow plasmid retention.

- **Plasmids**

pMVnc0842c for constitutive overexpression of the smallRNA was prepared by cloning the smallRNA *ncRv0842c in-frame* with P_{hsp60} promoter in the plasmid backbone pMV261 (low copy, KAN-R). To avoid the addition of unrequired nucleotides to the 5' region of the smallRNA, the UTR region of the promoter was truncated, and the smallRNA was cloned directly after the Transcription Start Site of the promoter (TSS). An artificial terminator was added to the 3' region of the smallRNA as previously described ⁷.

- **Culture media and bacterial culture**

Complete Middlebrook 7H9 (Becton, Dickinson, Franklin Lakes California USA) liquid culture medium was prepared accordingly to manufacturer instruction, and enriched with 10% of OADC (Oleic Albumin Dextrose Catalase, BBL™ Middlebrook) supplement and 0.05% Tween80 (Sigma-Aldrich, Missouri, USA).

Middlebrook 7H10 plates (Becton, Dickinson, Franklin Lakes California USA) were prepared accordingly to manufacturer instruction, enriched with 10% of OADC, and poured in sterile 10 cm non-treated culture plate (20 ml per plate).

The MTB laboratory reference strain H37Rv (NTCC7416) and the clinical isolates selected for the study were grown in 7H9 at 37 °C using vented flasks (Corning®, Sigma Aldrich, Missouri, USA). Experiments were performed with bacteria in log-Phase (OD₆₀₀ 0.2 to 0.5). Selective 7H9 media for plasmid selection were added with 50µg/mL of kanamycin (Sigma Aldrich, Missouri, USA), and/or RIF (Sigma Aldrich, Missouri, USA) at sub-inhibitory concentrations.

- **Whole genome sequencing**

Genomic DNA of the selected strains was extracted as described in and sequenced for Whole Genome Sequencing (WGS) analysis using Illumina technology as previously described ⁸. Briefly, the library was constructed

using a Nextera XT Library Preparation Kit (Illumina, California, USA) and was sequenced on Illumina HiSeq 2500 platform. FASTQ data obtained were analyzed on the MTBseq bioinformatic tool specific for MTB⁹.

- **Genetic variability in MTB smallRNAs**

Previous work in our laboratory described nearly 2000 smallRNA candidates¹⁴. Further studies based on RNAseq and literature evidence allowed to identify 756 “likely true” smallRNAs (data not shown)¹⁵. WGS data from 1811 isolates were collected for variants identification. Strains were divided into ancient lineages and modern lineages. Regions belonging to smallRNA candidates were selected for further analysis. SNPs presence was compared between ancient and modern lineages, selecting mutations present in one of the two categories in at least 95% of the isolates.

- **Northern blot**

The existence of the predicted smallRNA *ncRv0842c* was confirmed by Northern Blot on exponential and stationary phase H37Rv RNA. Northern blot analysis was conducted using ³²P labeled, customized RNA probes, complementary to the smallRNA sequence. RNA probes were obtained transcribing the DNA oligo RNA polymerase T7. X-ray film was developed with a Phosphorimager (Typhoon Amersham Biosciences Corp, California USA).

- **Sub inhibitory RIF stress**

RIF-induced stress was assessed at sub-inhibitory concentrations (25%, 50%, 90% RIF-MIC) on H37Rv and on L5 strains. Bacterial total RNA was extracted after 30m, 6h, and 24h post-treatment using the mirVana™ miRNA Isolation Kit (ThermoFisher, Massachusetts, USA) according to manufacturer instructions. Non-treated bacteria were used as a control. Briefly, the MTB cultures were grown in 30mL in complete 7H9 with RIF. To obtain the complete lysis of the wax and lipid-rich MTB membrane, the samples were homogenated in a TissueLyser (Qiagen, Hilden, Germany)

with zirconia beads (Zirconia/Silica Bead, 0.1 mm Diameter, BioSpec Oklahoma, USA). An Acid-Phenol:Chloroform extraction was performed following the mirVana kit protocol. This assay is specific for the extraction of total bacterial RNA, and has a high yield on preserving smallRNA.

The *TURBO DNA-free* Kit (Invitrogen, Massachusetts, USA) was used to remove DNA carry-over from RNA samples. Briefly, 5 μ l of 10x DNase buffer and 1 μ l of DNase were added to 44 μ l of RNA and incubated 25 minutes at 37 °C in a water bath. Then, to block the DNase reaction, the solution was moved in ice and added with 50 μ l of Acid-Phenol:Chloroform, vortex, and centrifuge at 13000 rpm for 5'. 40 μ l of the aqueous solution was supplemented with 0.1 Vol of 3M Sodium acetate and 3 Vol ice-cold 100% Ethanol and precipitated overnight at -20°C. The day after, the solutions were centrifuged at 13000 rpm, 4° C for 15'; the pellets were washed with 1 V of ice-cold 75% Ethanol, resuspended in Nuclease free water, and quantified by Bioanalyzer 2100 RNA 6000 nanochip (Agilent, California, USA).

- **Real-Time PCR**

Extracted RNA (2 ng) was retrotranscribed with the High-Capacity cDNA Reverse Transcription (RT) Kit (Applied Biosystems, Massachusetts, USA) following the manufacturer's instruction.

Quantitative Real-Time (qRT-PCR) was carried out in an Applied Biosystems™ 7500 Real-Time PCR System (ThermoFisher, Massachusetts, USA) using a SYBR® Green-based chemistry. Briefly, experiments were performed in 96-well plates (ThermoFisher, Massachusetts, USA), the reaction mix was composed of 12.5 μ l of 2X Universal SYBR™ Green PCR Master Mix (Applied technology), 100 nM of both forward and reverse primers (10 μ M mother solution), 5 μ l of first-strand cDNA and nuclease-free water to the final volume of 25 μ l. The thermocycler was set with the following parameters: 95 °C for 5 min followed by 40 cycles of denaturation (95° for 30 s), annealing (58° for 30 s) and elongation (72° for 30 s). To

ensure the reaction specificity, the melting curves were assessed for each primer setup experiment.

The Δ CT value was calculated using the SigA gene as endogenous control. H37Rv was used as calibrator for the experiments.

- **MGIT TPP**

The BACTEC MGIT 960 (Becton Dickinson, New Jersey, USA), culture system was used to calculate the Time To Positivity (TTP) of H37Rv strain carrying the overexpressing plasmid pMVncRv0842c. The mock H37Rv strain was used as a control. Briefly, 800 μ l of OADC, 200 μ l of bacterial suspension (OD_{600} 0.01) and 8 μ l or 16 μ l of 0.562 mg/ml RIF (respectively 90% and 2x 90% RIF-MIC) were added to a 7 ml 7H9 MGIT tube. The difference between the TTP from the mock strain and the overexpressing strain was calculated to obtain the Δ TTP (in hours, h).

- **Minimum inhibitory concentration (MIC)**

Microplate Alamar Blue assay (MABA) was used to determine the MIC of RIF and/or Carbonyl cyanide m-chlorophenyl-hydrazone (CCCP) for the selected strains. The drug concentrations tested ranged from 4 μ g/mL to 0.0007 μ g/mL for RIF and from 10 μ g/mL to 0.02 μ g/mL for CCCP. For each experiment, the selected drug was pre-diluted at 4x higher concentration in 7H9 enriched with OADC and KAN w/o Tween80 (7H9_MABA), and subsequently, 9 two-fold serial dilutions were performed in 7H9_MABA directly in the 96-well round-bottom tissue culture plates with a low evaporation lid (Becton Dickinson, New Jersey, USA).

Bacteria growth at Log phase were diluted in 7H9_MABA at OD_{600} 0.011 and inoculated on the previously prepared plates and incubated for 1 week at 37 °C.

To determine the MIC, the cell viability reagent AlamarBlue (Sigma-Aldrich, Missouri, USA) was added to each well; AlamarBlue is a stabilized solution containing the cell-permeable blue dye known as resazurin. Viable cells

(cells with active metabolism) are able to reduce the resazurin to resorufin, a pink dye; this colorimetric change can be easily detected.

The MIC is defined as the lower drug concentration able to sufficiently decrease the cell viability to prevent the colorimetric switch ¹⁰.

- **Drugs Combination Assay**

The combination assay between RIF and CCCP was performed with a constant concentration of CCCP (90% MIC determined as previously described). For each experiment, RIF was pre-diluted at 4x higher concentration in 7H9_MABA enriched with 2x 90% MIC of CCCP (7H9_MABA_CCCP); 9 twofold serial dilutions were carried out in 7H9_MABA_CCCP directly in the 96-well round-bottom tissue culture plates with a low evaporation lid (Becton Dickinson, New Jersey, USA). Bacteria growth at Log phase were diluted in 7H9_MABA at OD₆₀₀ 0.011 and inoculated on the previously prepared plates and incubated for 1 week at 37 °C.

The experiments were carried out as previously described only on the Mock control strains and the obtained MICs were compared with the MIC obtained for the corresponding *ncRv0842c* overexpressing strains in absence of CCCP.

Results

Sequencing analysis

To characterize the strains selected for smallRNA expression, whole-genome sequencing (WGS) was performed using Illumina technology. Sequencing produced results with a mean coverage >30 and the genomes were analyzed using the MTBseq software. Variant calls were mapped on genomic regions matching with smallRNA candidates, including 3' and 5' flanking regions (+/- 100 nucleotides) to avoid the omission of mutations in UTRs. The comparison between 85 ancient and 1726 modern lineages produced a list of 64 unique SNPs on 64 different smallRNA candidates: 22 SNPs were present only in ancient lineages while 42 in modern lineages. Among the ancient specific lineages, only 7 candidates harbor a SNP involving the coding region of the AS gene, while the remaining candidates involved UTRs or intergenic regions (Table 2). Interestingly, among the SNPs affecting AS smallRNAs we found two unique mutations, specific for ancient lineages and found to be synonymous on the CDS strand (Candidate_1221 and Candidate_1080). We decided to further investigate the role of the synonymous mutation of Candidate_1080.

This synonymous mutation falls in the *Rv0842* efflux pump gene at position 45 (ctA→ctG Leu45Leu), and coincides with a Tag→Cag mutation on the opposite (candidate_1080, renamed as *ncRv0842c*) strand, affecting its -10 promoter region (Fig. 3). Ancient-lineage specificity for this SNP was confirmed by cross-checking WGS data at the Leibniz-Zentrum database (Borstel, Germany).

Table 2: smallRNA candidates specific for ancient MTB strains from the WGS analysis. Among smallRNA considered, the candidate_1080 (renamed *ncRv0842c* according to the proposed standard nomenclature) was selected for this study¹¹. *ncRv0842c* has an ancient specific SNP affecting its -10 promoter region. AS: antisense; UTR: Untranslated region; IG: Intergenic; CDS: Coding Sequence.

Variant (coordinates)	Candidate smallRNA (Strand)	smallRNA type	SNP location	Codon change (on the AS gene)
1639643t>c	candidate_1221 (-)	AS	CDS <i>Rv1453</i>	cgt-cgc / Arg-Arg
1639643t>c	candidate_2064 (+)	AS	3' UTR <i>Rv1454c</i>	/
1906336a>g	candidate_2074 (+)	Whitin UTRs	3' UTR <i>Rv1681</i> / 5' UTR <i>Rv1682</i>	/
1931470c>t	candidate_2075 (+)	AS	5' UTR <i>Rv1704c</i>	/
2039884t>g	candidate_1296 (-)	AS	CDS <i>Rv1800</i>	tgt-tgg / Cys-Trp
2052687g>c	candidate_1298 (-)	AS	3' UTR <i>Rv1809</i>	/
2447539t>c	candidate_1366 (-)	Whitin UTRs	5' UTR <i>Rv2185c</i> / 3' UTR <i>Rv2186c</i>	/
3007238t>c	candidate_1863 (-)	Whitin UTRs	5' UTR <i>Rv2689c</i> / 3' UTR <i>Rv2689c</i>	/
3027606c>g	candidate_1461 (-)	AS	CDS <i>Rv2714</i>	ctg-ctc / Leu-Leu
3369869c>a	candidate_2123 (+)	AS	5' UTR <i>Rv3010c</i> / 3' UTR <i>Rv3011c</i>	/
3415332a>g	candidate_1507 (-)	AS	3' UTR <i>Rv3053c</i>	/
4200220t>c	candidate_886 (+)	AS	5' UTR <i>Rv3753c</i>	/
2097144g>c	candidate_1310 (-)	AS	CDS <i>Rv1847</i>	gtg-ctg / Val-Leu
2097144g>c	candidate_2081 (+)	AS	5' UTR <i>Rv1846c</i>	/
2019236t>g	candidate_1282 (-)	AS	CDS <i>Rv1782</i>	gtc-ctt / Val-Pro
1496964g>t	candidate_2059 (+)	AS	3' UTR <i>Rv1329c</i>	/
1203824t>c	candidate_1129 (-)	AS	CDS <i>Rv1078</i>	cag-tag / Gln-Stop
1040706t>g	candidate_135 (+)	AS	CDS <i>Rv0932c</i>	tcg-gcg / Ser-Ala
938246a>g	candidate_1080 (-)	AS	CDS <i>Rv0842</i>	cta-ctg / Leu-Leu
1287112t>c	candidate_161 (+)	IG	IG	/
139756t>c	candidate_668 (+)	IG	IG	/
3302683c>t	candidate_442 (+)	IG	IG	/

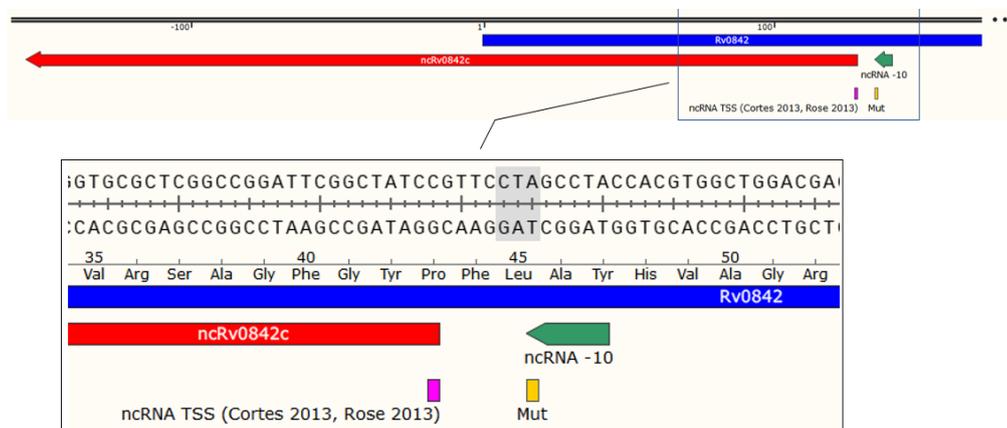


Fig. 3: WGS analysis show a synonymous mutation in *Rv0842* (L45L) specific for the ancient lineages (L1, L5, L6) abrogating the -10 promoter region of the cis-encoded smallRNA *ncRv0842c*

Northern blot

Northern blot was performed to confirm the existence of the predicted smallRNA *ncRv0842c*. The predicted size of the smallRNA candidate *ncRv0842c* is 283bp; according to its coordinates, 937956 – 938239 (*strand -*), it resulted to overlap for 127bp the gene *Rv0842*, coding for an efflux pump member of the major facilitator superfamily (MFS).

The 127 bp overlap entails that the TSS and the promoter region of the smallRNA candidates are antisense to the coding region of the gene.

To avoid unreliable results, RNA was extracted from a planktonic culture in both exponential and stationary growth phases and the Northern blot revealed a defined band, raging around 250bp, in both tested conditions even if the band obtained for the Log-phase extract shows a higher intensity.

The other bands shown in the image (Fig. 4) have lower signal intensity, thus indicating that they are either products of RNA maturation or degradation of the *ncRv0842c*.

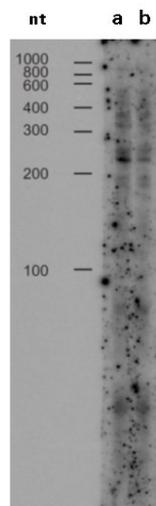


Fig. 4: Northern blot analysis revealed the presence of a specific band at the expected length (about 250) at exponential (a) and at stationary (b) growth phase. Multiple bands indicate putative maturation fragments of the smallRNA.

qRT-PCR

The expression of the smallRNA *ncRv0842c* was measured using a q-RT PCR and the *sigA* gene was used as an endogenous control to normalize the results obtained.

The pilot experiments were run on RNA extracted from a planktonic culture in both exponential and stationary growth phases and in the strains belonging to the different lineages of MTB and in *M. africanum*. Results were normalized on the laboratory strain H37Rv. qRT-PCR showed that the basal expression of *ncRv0842c* in the *M. africanum* strains considered is greatly lower in comparison with H37Rv, in both exponential (-16 fold change; *P-value* 0.046) and stationary phase (-32 fold change; *P-value* 0.013) (Fig. 5). Drug resistance background nor epidemiological features (sporadic vs clade-related isolates) did not show to affect the baseline expression of the smallRNA (data not shown).

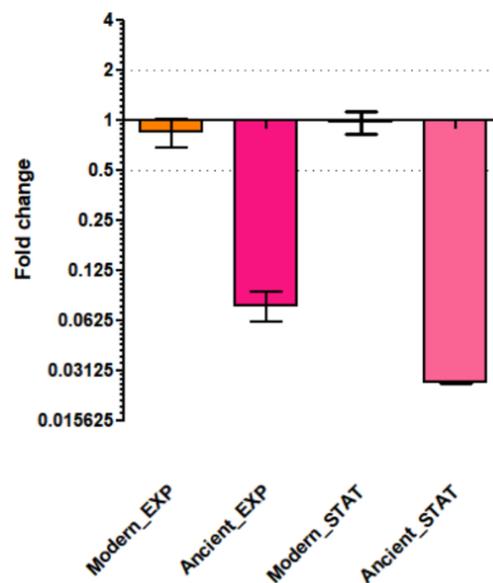
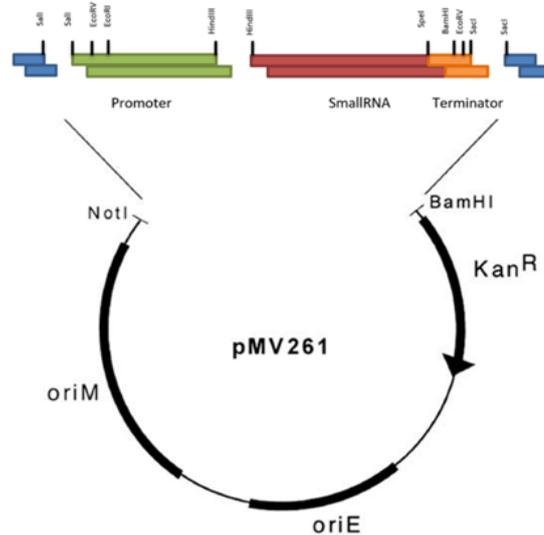


Fig. 5: qPCR of *ncRV0842c* smallRNA in modern and ancient MTB strains (normalized on H37Rv values). Result show that *ncRv0842c* is strongly downregulated in ancient lineages both in exponential and stationary planktonic growth phases

To better characterize the function of *ncRv0842c* we constructed an over-expressing plasmid using the vector pMV261 and cloning the smallRNA at



the restriction sites NotI and BamHI (Fig. 6).

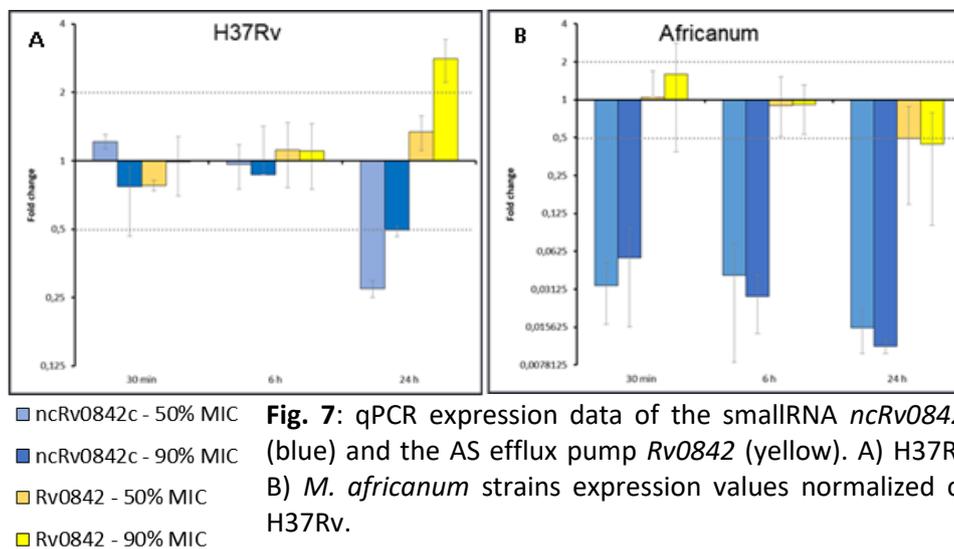
Fig.6: pMV261 overexpressing the smallRNA *ncRv0842c*. OriE: E. coli Origin of Replication; Kan^R: kanamycin resistance cassette, OriM: MTB Origin of Replication.

Our previous work based on RNAseq, highlighted the up-regulation of *Rv0842* and the simultaneous down-regulation of *ncRv0842c* during RIF stress (data not shown). Literature data showed that the *Rv0842* efflux pump was induced during several antibiotic-based stresses, including RIF¹². In particular, clinical isolates monoresistant to RIF showed over-expression of the efflux pump¹³. Accordingly, the subsequent experiments aimed to characterize the smallRNA function in H37Rv and *M. africanum* in response to the sub-inhibitory concentration of RIF (RIF-challenge).

Before proceeding with the RIF-challenge, the MIC for RIF was assessed on the strains considered and resulted 0.0625 µg/mL for H37Rv and the 2 *M. africanum* isolates; therefore, for the subsequent experiments, RIF concentrations equal to 0.0156 µg/L (25%), 0.0312 µg/mL (50%) and 0.0562 µg/mL (90%) were used. For this study, early (30 minutes), middle (6 hours) and late (24h) time points were selected.

The use of RIF at 25% MIC was not showing significant changes (data not shown). *ncRv0842c* was found down-regulated (-4 fold changes) in H37Rv treated with 50% of the MIC (p-value 0.0059). Despite the amplitude of the regulation decreased (-2 fold changes), a similar trend was observed at 90% of the MIC. The efflux pump *Rv0842* resulted significantly up-regulated (2 fold change), in comparison with the untreated control with 90% of the MIC (*P-value* 0.0089), whereas lower RIF concentrations (50% MIC) induced slight changes in the expression of the pump (Fig. 7 A).

Regarding the *M. africanum* strains, the expression of *Rv0842* resulted slightly induced under treatment with 90% of the MIC, however, the difference with the control was not found statistically significant (*P-value* 0.05). On the contrary, the smallRNA expression levels were mostly unaffected and strongly downregulated (-128 fold changes) compared to H37Rv (*P-value* <0.001) (Fig. 7 B), thus aligned with the baseline expression profiling showed above (Fig. 5).



MGIT TTP

To ensure that the over-expression of *ncRv0842c* does not affect the *in vitro* growth of the mutant strains, thus introducing a bias in the phenotypical evaluation discussed in the next paragraphs, the “time to

positivity” (TTP) of H37Rv strain carrying the overexpressing plasmid pMV*ncRv0842c* and of the H37Rv strain containing the empty plasmid was assessed by the BACTEC MGIT 960. Experiments were performed in basal conditions (no RIF), during the RIF-challenge (90% RIF MIC) and in extreme conditions (2x 90% RIF MIC).

In basal conditions, the growth rate resulted unaffected by the overexpression of the smallRNA. On the contrary, in both RIF-induced stress conditions, the constitutive overexpression of *ncRv0842c* causes growth delays during RIF challenge in MGIT TTP assay (Δ TTP>100 hr). (Fig. 8). Similar findings were confirmed by testing a RIF-S, modern lineage clinical isolate (data not shown).

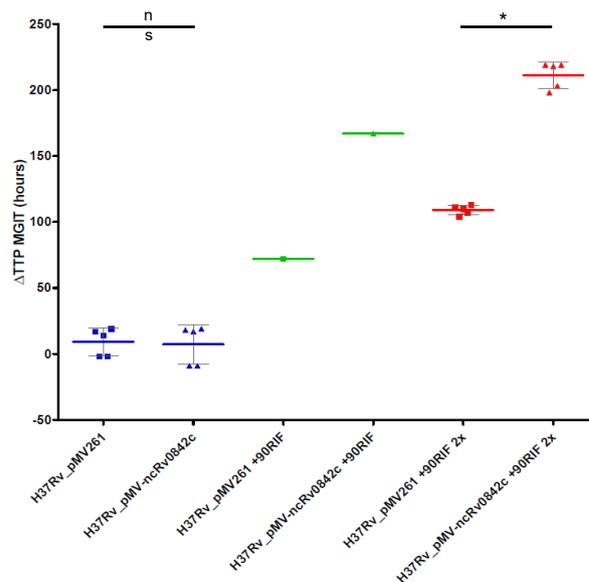


Fig. 8: TPP in MGIT of H37Rv under RIF stress. Blue) H37Rv with the empty control vector pMV261 and the *ncRv0842c* overexpressing plasmid w/o RIF-stress. Green) H37Rv with the empty control vector pMV261 and the *ncRv0842c* overexpressing plasmid with 90% of RIF-MIC. Red) H37Rv with the empty control vector pMV261 and the *ncRv0842c* overexpressing plasmid with 2x 90%RIF-MIC.

Minimum inhibitory concentration (MIC)

To evaluate the functional role of the smallRNA *ncRv0842c*, the minimum inhibitory concentrations (MICs), defined as the lower drug concentration

required to inhibit visible growth as detected by resazurin reduction, were assessed for six “S” (RIF-sensible) and five “iM” (increased RIF MIC) strains overexpressing *ncRv0842c* and on their respective controls (mock, pMV261 empty plasmid). As shown in the graph below, RIF MIC is reproducibly and consistently reduced by ≥ 1 MIC dilution when the smallRNA is overexpressed in all the modern strains selected, thus halving the concentration of RIF required to inhibit the bacterial growth (Fig. 9). This phenotype cannot be reproduced on strains carrying the L45L mutation on *Rv0842* gene (ancient lineages) in which the basal expression of the smallRNA is almost completely switched off.

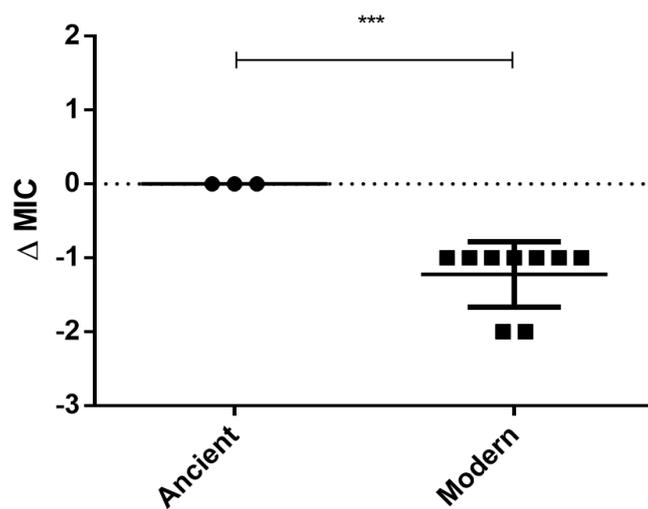


Fig. 9: MABA delta MIC variation of modern and ancient MTB strain carrying the smallRNA overexpressing plasmid. Modern strains’ RIF sensitivity is increased by about 1 MIC dilution. Each point represents the average delta MIC value of a strain. Experiments were performed at least 2 times, with 3 technical replicates each.

To confirm that the observed Δ MIC can be attributed to the downregulation of an efflux pump, we tested the effect of the well-characterized efflux pump inhibitor CCCP on mock control strains. The phenotypic effect of CCCP was assessed by calculating the RIF-MIC of the mock control strains undergoing sub-inhibitory (90% MIC) CCCP stress.

As a control, we tested the contribution of CCCP, randomly in three of the selected strains. The graphic below demonstrated that the sub-inhibitory administration (90% MIC) of CCCP has a phenotypic effect comparable to the overexpression of *ncRv0842c* (Fig. 10), thus supporting the hypothesis that the smallRNA is acting as a negative regulator of the efflux pump *Rv0842*.

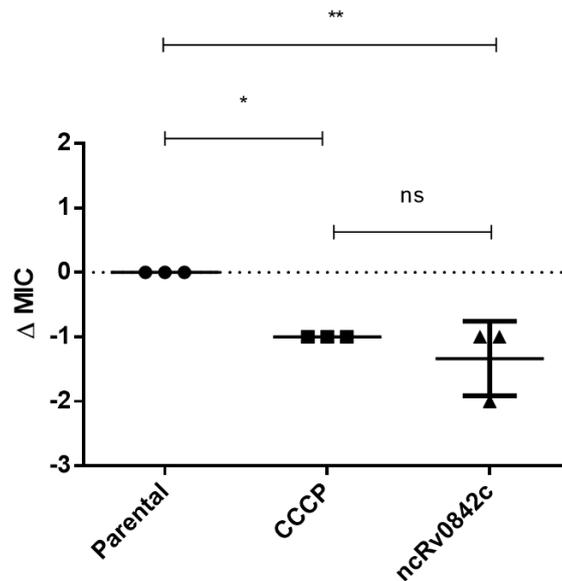


Fig. 10: Delta MIC variation. MABA assay demonstrated that the sub-inhibitory administration (90% MIC) of CCCP has a phenotypical effect comparable to the overexpression of *ncRv0842c*. Each point represents the average delta MIC value of a strain. Experiments were performed at least 2 times, with 3 technical replicates each.

Discussion

The DNA mutations, discovered and categorized thanks to extensively genomic studies conducted over the last three decades, failed to fully explain all the resistance found among the different strains; thus, suggesting that other, epigenetic, mechanisms are involved in the drug resistance or drug-tolerance development.

MTB smallRNAs have only recently become a topic of study; despite it being predicted that H37Rv genome encodes for at least 92 noncoding RNAs and a few additional hundreds have been predicted, just a few of them have been studied and characterized; therefore, their mechanisms of action and their functions within the cell are still mostly unidentified¹⁴. The mode of action of these regulatory molecules is very different from protein regulators, for which the waiting time is determined primarily by the turnover rate of the protein¹⁵.

Unraveling the biological functions and the mechanism of action of smallRNA may lead to the development of new therapeutic strategies to counteract the resistance development of MTB.

In the present study, we investigated whether MTB lineage-specific SNPs could map in smallRNA coding regions. The comparison between ancient and modern lineages identified a list of 65 lineage-specific SNPs involving smallRNA regions.

Interestingly, a synonymous mutation on the gene encoding for *Rv0842* (ctA→ctG) at codon 45 was found specific for the ancient lineages (L1, L5, L6) and mapped in the promoter region of *ncRv0842c*. We characterized the role of the smallRNA *ncRv0842c* cis-encoded to the *Rv0842* gene (a putative major facilitator superfamily (MFS) efflux pump) in different MTB lineages exposed to RIF stress.

The smallRNA was found to be strongly downregulated in ancient strains when compared to H37Rv and other modern strains, both in the exponential and in the stationary phase. This finding confirmed that the

synonymous mutation, L45L, abrogates the -10 promoter region of *ncRv0842c* consequently affecting the expression of this *cis*-encoded regulator without affecting the amino acidic sequence of the efflux pump. To study the mechanism of action of the smallRNA and to evaluate its capability to regulate the antisense gene, the expression levels of both the efflux pump and its smallRNA were analyzed under treatment with different RIF sub-inhibitory concentrations. In H37Rv (representative of modern lineages), RIF challenge induced the down-regulation of the smallRNA and the over-expression of the efflux pump. These findings suggest that the smallRNA acts as a negative post-transcriptional regulator by mediating or contributing to the degradation of *Rv0842* mRNA.

Concerning the ancient lineage strains, both the smallRNA and the efflux pump expression resulted unaffected by RIF-stress. On one side, these results confirm that the synonymous mutation impairs the expression of the smallRNA in ancient lineages; on the other side, it seems the efflux pump is unresponsive to RIF challenge, thus suggesting that additional stress response networks are missing. Future studies will help to shed light in the explanation of the efflux pump involvement in ancient MTB strains.

To characterize further the role of the smallRNA, we generated overexpressing mutants where the *ncRv0842c* is expressed by means of a constitutive promoter. Mutants were generated in clinical isolates belonging to both ancient and modern lineages. To evaluate the metabolic burden of the plasmid we considered the TTP in MGIT. Accordingly, we could not find any difference in TTP between mock strains harboring the empty vector (pMV261) and mutant strains overexpressing the smallRNA (pMV*ncRv0842c*) in a standard culture medium. However, a growth delay (approx. 3 days) was observed when RIF was added to the tube containing the mutant strains pMV*ncRv0842c*, whereas RIF was not affecting TTP in mock strains. These findings further support the hypothesis that the smallRNA and its *cis*-encoded target CDS are involved in RIF transportation. H37Rv pMV261*ncRv0842c* over-expressing mutant showed a 1-dilution RIF MIC reduction in several rounds of independent experiments. To validate

our findings, we selected 9 phylogenetically modern clinical strains showing different RIF MIC to be transformed with pMV261*ncRv0842c* over-expressing plasmid and evaluated if the overexpression of the smallRNA affected the strain-specific MIC for RIF. MABA assay showed that RIF MIC is reduced at least by 1-dilution when the smallRNA is overexpressed in all the strains selected. This effect on MIC cannot be reproduced in ancient strains pMV261*ncRv0842c* over-expressing mutant, at least, not with a low-copy plasmid. Also, the MIC reduction was not observed in RIF resistant clinical strains harboring mutations in the RRDR (the region of interaction between RIF and its target RNA polymerase) and engineered by pMV261*ncRv0842c* (data not shown).

To further elucidate the mechanism of action of *ncRv0842c*, we used the CCCP efflux pump inhibitor. CCCP has been reported to act on the *Rv0842* efflux pump, reducing its activity¹³. Mock strains (pMV261) treated with sub-inhibitory concentrations of CCCP recapitulate the phenotypic 1-dilution reduction on the MIC RIF, thus confirming that *ncRv0842c* is one of the regulators of *Rv0842*.

Despite our experimental does not permit a fine understanding of the dose-response profile, our results suggest that the down-regulation of *ncRv0842c* happens at lower RIF concentrations (50% MIC) compared to the up-regulation of the *Rv0842* pump (90%), thus pointing out for a possible mechanism of drug tolerance, rather than resistance. Further studies better focusing on the dose-response analysis are needed to further elucidate this aspect.

Besides characterizing for the first time the role of *ncRv0842c* in response to RIF, our study provides a few additional cues. First, the definition of synonymous mutation could be obsolete when cis-encoded post-transcriptional regulators are involved. Second, lineage-specific mutations can have relevant effects without directly affecting protein functions, thus suggesting that complex post-transcriptional regulatory networks are involved in determining lineage-specific features. These

findings provide the baseline for further studies in this underestimated area in TB.

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Chapter 4

Conclusions

The genetic variability among MTB clinical isolates may have dramatic consequences on the outcome of infection. Many *in vitro* and *in vivo* studies have demonstrated strain-dependent variation in key aspects of virulence such as stress survival, transmission, pathology, drug resistance and lethality ¹. On the other hand, the immunological state of the host is also important and is becoming increasingly relevant over the years. Indeed, the polarization states of macrophages have a huge impact on their functions and early phases of infection are crucial for the immune response activation and for the eradication of the infection ².

In order to keep into account both the host and the pathogen sides, we adopted an experimental plan specifically suited for considering as relevant variables the phenotypic diversity of macrophages and the genetic variability of MTB. The fate of TB infection highly depends on the activation status of the host immunity, and the early phases of the infection are crucial for immunity priming and for the development of a granuloma ³. The use of THP-1 cells allowed us to use a reproducible standardized model of host-pathogen interaction, which has been reported to act similar to human monocyte-derived macrophages ⁴. However, despite this model based on the THP-1 cell line being recognized in the field, the phenotypic status of the macrophage is often neglected when focusing on mycobacterial virulence. Similarly, only in recent years, researchers begun considering strains different from Herdman, H37Rv, H37Ra or CDC1551. Thus, our approach combining both human and MTB variability, despite its simplicity, represents the most comprehensive concept for unravelling MTB virulence. Indeed, virulence features of a given pathogen cannot be unlinked from the susceptibility (and resistance) features of the host.

We decided to focus our attention on the early phases of the MTB-macrophage interaction because of high importance in mounting an effective immune response, thus influencing the outcome of infection. In

addition, the laboratory is particularly interested in smallRNAs. As mentioned, these key regulatory molecules are among the most rapid, cost-effective and efficient modulators of gene expression in organisms. They act at the post-transcriptional level and are reported to be the earlier players in broad stress response with implications also for drug resistance and virulence for many bacteria. In this view, we found it more relevant to focus on the earlier phases of the host-pathogen interaction as this study will represent the baseline for further characterizing the role of smallRNAs in the very first stages of infection.

The use of M1 and M2 polarized THP-1-derived macrophages recapitulate the two extreme phenotypes found in the alveolar macrophages. This, combined with the different MTB lineages considered, provided a broader view of the diverse fates of the MTB-macrophage interaction.

In this project, we investigated the outcome of this interaction keeping into account different aspects and experimental strategies considering host response, bacterial escape, survival mechanisms. First, the host-pathogen interaction has been analyzed at a single-cell level by live-imaging confocal microscopy, giving us a large amount of information about how both host and MTB diversity could influence the infection, but also providing valuable details about the heterogeneity found even within the same infected macrophages. The analysis on the acidification of the phagolysosomes provided relevant hints on how much the genetic diversity of MTB affect this process, and highlighted those previous findings based on the laboratory strains are not always recapitulating the behaviour of clinical isolates. Furthermore, our approach based on earlier time points showed a clear discrepancy between the time of response in M1 and M2 macrophages. We could also show data about the percentage of macrophages effectively bringing the mycobacteria to an acidified compartment, and to shed light on the microenvironment of infection with an outlook on bystander cells. Similar conclusions can be drawn for the evaluation of apoptosis. The lack of induction of autophagy was probably the most common feature observed in all the settings. Several studies

highlighted a central role for autophagy in mycobacterial clearance; however, despite blocking autophagy seems a common strategy across MTB lineages, literature evidence suggests that different mechanisms to achieve this scope evolved ⁵.

In general, this project contributed to expanding our knowledge on the implications of MTB genetic diversity. Even though we cannot foresee a direct impact in the clinical management of TB, our study poses relevant bases for further dissection of the cellular and molecular mechanisms behind these observations. In the long term, this could further contribute in building evidence for translational applications related to the use of genotypic approaches for developing better diagnostic and treatment strategies tailored keeping into account the genetic background of MTB.

The reasons for selecting earlier time points in our study design have been outlined above. This brings together some limitations in the study as medium-long term effects of the host-pathogen interaction are lost. In particular, this could have hindered relevant findings related to autophagy and apoptosis/necrosis, both expected to happen later. Furthermore, whereas our single-cell approach tracked relevant mechanisms in the macrophages, we could only follow the processes in their entirety without dissecting specific molecular pathways. Both the use of additional time points and more specific molecular markers for sub-processes could represent further improvements to our approach.

In our second work, we decided to study the impact of genetic variability of the MTB strains directly on the smallRNAs. SmallRNAs are recently emerged as key factors in the regulation of critical bacterial features (as observed for microRNAs in humans), but MTB smallRNAs are poorly characterized ⁶. The advent of next-generation sequencing enabled access to WGS and this technology renewed the interest in identifying new markers of drug resistance in MTB. Indeed, molecular DST seems the most obvious and achievable approach to support the WHO End TB Strategy in providing universal DST access. In 2021, the WHO published the first

catalogue of mutations associated with drug resistance in MTB ⁷. Interestingly, despite the genotype-phenotype associations derived in the catalogue were considering data from nearly 40,000 clinical isolates, this could not help in identifying new genetic markers to fill the gaps in sensitivity for the detection of resistance. Indeed, the sensitivity of the genetic markers reported for rifampicin is still between 90-95%, whereas for isoniazid the catalogue is reporting a sensitivity close to 90%. The primary aim of this exercise was to provide confidence around the interpretation of mutations, rather than discover new markers; however, the fact that the analysis of so many isolates did not allow to go further close to 100% in sensitivity clearly suggests that new approaches and targets need to be considered.

For the first time, we identified and characterized a smallRNA (*ncRv0842c*) involved in the detoxification of antibiotic stress. Interestingly, we found a lineage-specific synonymous mutation abrogating the expression of the smallRNA, highlighting the existence of main differences in antibiotic tolerance between ancient and modern lineage, but also providing evidence on the need to redefine the concept of silent mutation that keeps into account the network of antisense regulators.

Among the limitations of this study, there is the lack of a specific inhibitory compound for the efflux pump (*Rv0842*) regulated by the smallRNA that prompted us to use a broad efflux pump inhibitor such as the CCCP. Also, it would be useful to achieve smallRNA silencing in modern strains and analyze how this affects rifampicin MIC. Given the fact that the efflux pump is an MSF, the inclusion of additional drugs could provide further data.

As a future perspective, we aim to increase the number of smallRNAs characterized for their role in both virulence and drug resistance. The availability of data for lineage-specific smallRNA variants will be an additional aid to the selection and analysis of the candidates.

In conclusion, this project contributed new relevant insights on the role of MTB genetic diversity in delineating pathogenic features, and on how it could affect yet unexplored smallRNA-based regulatory pathways.

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7. WHO Catalogue of mutations in *Mycobacterium tuberculosis* complex and their association with drug resistance 2021

ANNEX

Characterization of *Mycobacterium tuberculosis* smallRNA *ncRv0757c*

Introduction

Small non-coding RNAs (smallRNAs) have been identified as crucial elements that regulate gene expression in bacterial stress responses and virulence ¹. In this perspective, the investigation of smallRNAs in different lineages of MTB will provide further information about the regulation of the stress response in this pathogen.

We are investigating the role of *Mycobacterium tuberculosis* (MTB) smallRNAs during macrophage infection by means of in vitro and ex vivo infection models with the goal of determining whether these regulatory molecules are involved in the survival and in the virulence of the tubercle bacilli and if differences among lineages exist. Interestingly, several genes involved in the pathogenesis of tuberculosis and playing a relevant role in MTB survival during macrophages infection have been shown to be under the putative regulation of cis-encoded smallRNAs ². Whereas current research is starting to delineate the role of genetic diversity in determining the spectrum of virulence features across the 7 main MTB phylogenetic lineages described, no data are available on how this heterogeneity affects smallRNA expression and function. Moreover, since virulence is a relative capacity of a microbe to cause damage in a susceptible organism, the complex heterogeneity of the human immune system cannot be ignored ³.

In particular, the phenotypic diversity and plasticity of human macrophages (first line of defense against MTB) is an underestimated variable in experimental designs aiming at dissecting the pathogenesis of MTB strains.

PhoP (response regulator of a two-component system in which *phoR* is the response regulator) is a key regulator of pathogenesis in MTB: it is a transcriptional regulator that controls at least 2% of the genome of *M. tuberculosis* and among the genes under its regulation, there are relevant virulence factors and key players in acidic pH stress response, with the latter considered one of the crucial features enabling intramacrophagic survival to MTB⁴.

Despite the central role of PhoP in virulence, surprisingly its mechanism of action differs between modern and ancient strains⁵. *phoP* mutants have been considered for the development of new vaccine candidates because the deletion of this gene in SO2 mutant causes an attenuation of the virulence⁶. Even though, the precise mechanism by which PhoP is involved in pH response remains poorly clear. The study of the mechanism of regulation of *phoP*, including smallRNAs, is crucial to fully understand the complexity of the regulon and to develop an attenuated strain in order to test new vaccine candidates for TB. In this study, we decided to focus our attention on the smallRNA *ncRv0757c* which is antisense (AS) to the 5' region (promoter) of the gene *phoP*. In our laboratory, we previously confirmed the existence of this RNA by northern blot (data not shown).

Methods

To study the role of the smallRNA taking consideration of both human and bacteria variability, we decided to use a model of in vitro infection using THP-1 derived macrophages polarized toward M1-like (LPS) or M2-like (IL-4) macrophages. THP-1 (ATCC® TIB-202™) cells were differentiated in complete RPMI (with L-glutamine 2mM, Hepes 10mM, Non-essential amino acids 100nM, Sodium Pyruvate 1mM, Sigma Aldrich, Missouri, USA) supplemented with 10% FBS (Fetal Bovin Serum, Euroclone), antibiotics (penicillin G 100 U/ml and streptomycin sulfate 100 U/ml, Sigma Aldrich Missouri, USA), and 100 nM of phorbol 12-myristate 13-acetate (PMA,

Sigma-Aldrich Missouri, USA) for 3 days in Nunc™ 12 wells plates (500000 cells/well) (Thermo Fisher, Massachusset, USA) ⁷. Type I collagen (Sigma Aldrich, Missouri, USA) was used for cell adhesion. Three days after removal of the PMA, macrophage-like cells were polarized in cRPMI medium without antibiotics for 16 h with either lipopolysaccharide (50ng/mL, Sigma-Aldrich Missouri, USA) to induce an M1-like phenotype or human IL-4 (20 ng/mL, *E. coli*-derived human IL-4 protein, R&Dsystems) to induce an M2-like phenotype prior to the infection ^{7,8}. We tested the intramacrophagic expression of both *phoPR* regulon and the *ncRv0757c* smallRNA of different MTB lineages, including laboratory strains (H37Rv, H37Ra), and 13 clinical strains belonging to modern (more virulent) and ancient (less virulent) strains (Table 1). Intramacrophagic expression was evaluated by means of qPCR based on TaqMan chemistry. Ribosomal RNA 16S was used as endogenous control.

Table 1: Strains selected for this study

Strain	Lineage			Characteristics
	Name	N	Phylogenetic Origin	
<i>M. africanum</i>	West African	6	Ancient	Less virulent, clinical isolate
<i>M. tuberculosis</i>	EAI	1	Ancient	Less virulent, clinical isolate
<i>M. tuberculosis</i>	Beijing	2	Modern	More virulent, clinical isolate
<i>M. tuberculosis</i>	CDC155 1	4	Modern	More virulent, clinical isolate
<i>M. tuberculosis</i>	H37Ra	4	Modern	Less virulent, laboratory strain
<i>M. tuberculosis</i>	Haarlem	4	Modern	Virulent, clinical isolate
<i>M. tuberculosis</i>	H37Rv	4	Modern	Virulent, reference strain
<i>M. bovis</i>	Bovis	n.a.	Intermediate	Intermediate phenotype
<i>M. bovis</i> BCG	Bovis	n.a.	Intermediate	Vaccine strain

Results and discussion

We measured the smallRNA baseline expression in exponential and in stationary phases in modern and ancient strains and we found no statistical difference between the strain tested in planktonic growth (Fig. 1).

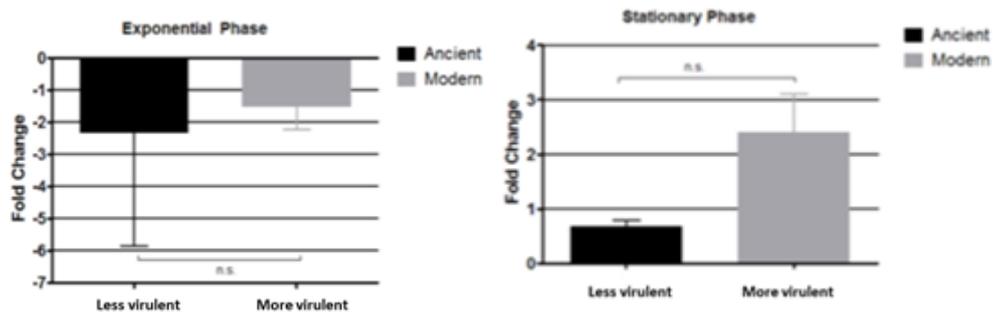


Fig. 1: *ncRv0757c* smallRNA expression in less virulent and more virulent MTB strains (normalized of H37Rv) in exponential and stationary growth phases.

We infected polarized macrophages at a MOI 10:1 and we measured the intramacrophagic expression of the smallRNA in the same MTB strains. In M1 macrophages the smallRNA *ncRv0757c* is highly upregulated in more virulent strains at 4h p.i. (compared to H37Rv) and its upregulation is maintained up to 48h, while the smallRNA is highly downregulated in less virulent strains (compared to H37Rv). We also observed a decreasing trend of the smallRNA upregulation during time, suggesting that the smallRNA is accomplish its function at early timepoints after infection but it is slowly disabled when infection time progresses (Fig. 2). In M2 macrophages the expression of the small-RNA does not change during time in both virulent and less virulent categories (Fig. 2).

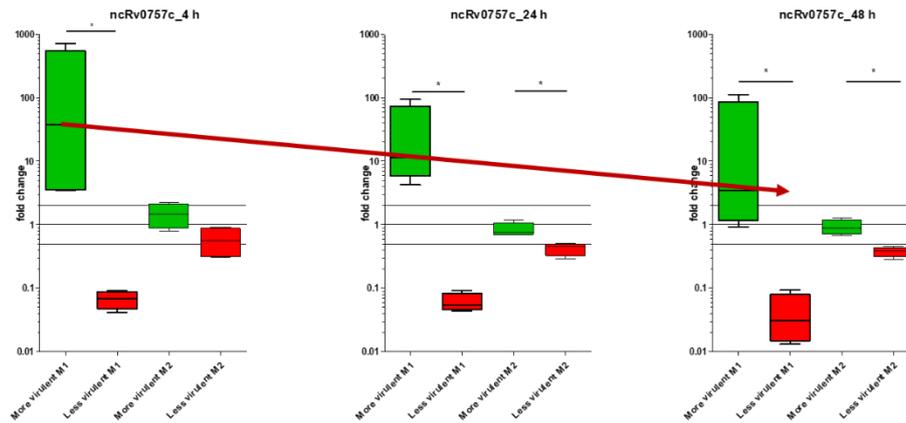


Fig. 2: MTB *ncRv0757c* expression in M1 and M2 THP-1 derived macrophages, at 4, 24 and 28h post-infection. Green: More virulent strains; Red: less virulent strains.

Concerning the expression of the response regulator *phoP*, we saw that in M1 macrophages its expression is slightly upregulated at 4h p.i. only in virulent strains while it remains invariable in less virulent strains. No changes in the expression of *phoP* were observed in M2 macrophages (Fig. 3).

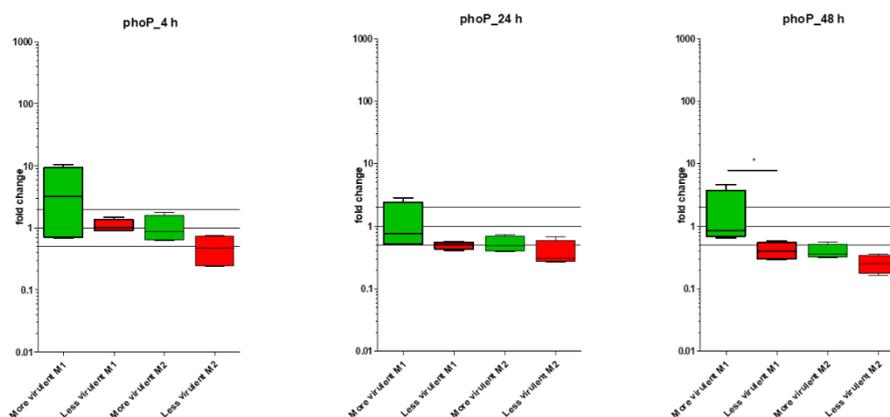


Fig. 3: MTB *phoP* expression in M1 and M2 THP-1 derived macrophages, at 4, 24 and 28h post-infection. Green: More virulent strains; Red: less virulent strains.

In order to better understand the mechanism of action of this smallRNA we decided to engineer over-expressing plasmids, taking into consideration

the work published by K. Arnvig et al. in which the target smallRNA is under the control of *M. smegmatis* *rrnB* promoter⁹. An artificial terminator to the smallRNA sequence is also included⁹. First attempts in obtaining detectable over-expression of *ncRv0757c* in MTB were unsuccessful. We decided to switch to the use of the low copy plasmid pMV261 in order to overexpress *ncRv0757c* under the control of a strong constitutive MTB promoter (P_{hsp60}). Our preliminary data show a colony phenotype similar to the SO2 *phoP* mutant vaccine candidate, with smaller colonies and with lower growth rate (Fig. 4). However, the phenotype obtained was found to be unstable.

We are now evaluating the use alternative over-expressing strategies. In particular, we working in collaboration with the University of Padua at the following constructs:

- Integrative plasmid pYUB413 with P_{rrnB} promoter
- Replicative inducible plasmid pFRA71 with P_{ptr} (pristinamycin inducible) promoter.

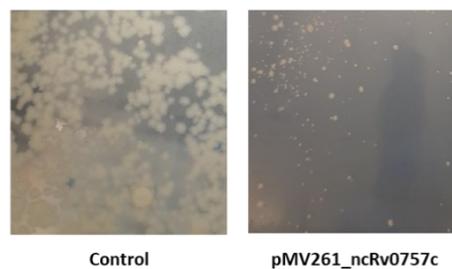


Fig. 4: preliminary phenotypical results of H37Rv overexpressing the smallRNA *ncRv0757c* with the replicative vector pMV261

In addition, recent findings showed that even few nucleotides (<5) modifying the 5' of the smallRNA can lead to severe and faster degradation of the regulator¹⁰. Given that two different 5' ends at a few nucleotides of

distance have been described for *ncRv0757c*, we are also evaluating how this can affect the overexpression strategy. This is particularly relevant for the inducible approach as it requires the inclusion of a nucleotidic stretch at the 5' end of the target smallRNA for pristinamycin binding ¹⁰.

Our hypothesis is that this smallRNA could act inhibiting *phoP* at a transcriptional or post-transcriptional level. The smallRNA exerts its effect at an early timepoint in more virulent strains but so far we don't have enough data to confirm that our proposed mechanism is correct. This is particularly complex because the smallRNA is part of a complicated (and yet not fully decoded in MTB) regulatory network tightly tuning *phoP* response to stresses (Fig 5).

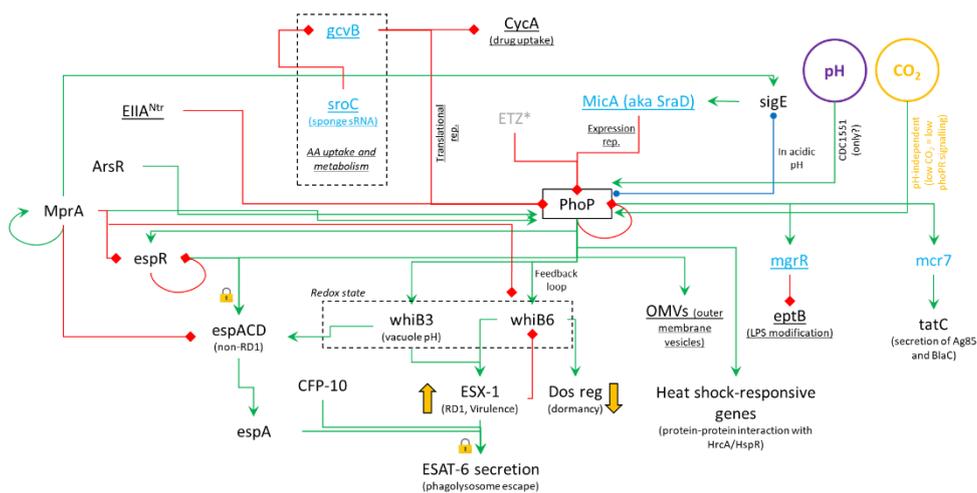


Fig. 5: MTB PhoP regulation network ¹¹⁻²⁸

Red lines with square ends: inhibition/repression

Blue lines with circle ends: interaction

Green lines with arrow ends: activation/induction

Underlined text: observed in bacteria other than mycobacteria

Blue text: smallRNAs

Grey text: chemical compounds

*The ethoxzolemid (ETZ) inhibits the *phoPR* operon (not *phoP* directly); observed in CDC1551 (not in other strains)

Lock: co-requirement of elements

The early induction of *Rv0757c* reduced *phoP* mediated virulence in “more virulent” strains at early timepoint after phagocytosis, contributing to hiding the bacteria to the immune system, while in less virulent strains this

pathway seems less relevant at the time point considered. This hypothesis seems aligned with our findings on phagolysosomal acidification, where the modern/more virulent strains were found to delay M1 macrophagic response (further information on Chapter 2).

As a next step, the use of *ncRv0757c* overexpressing mutant strains will allow to further elucidate the mechanism of action of the smallRNA and its effects on the *phoPR* operon, including broader influence on cytokines secretion, phagolysosomal acidification, apoptosis, and autophagy. The mutants will be analyzed for their ability to survive inside the macrophages and to kill macrophages.

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Supplementary Table

Category	Gen Name	Rv number	Description	Attenuation evidences	
				Model	Result
Lipids and Fatty Acid Metabolism	<i>kasB</i>	Rv2246	3-oxoacyl-[acyl-carrier protein] synthase 2 <i>kasB</i>	C57BL/6 mice (I _{da})	Reduced CFUs in organs and lung pathology Increased animal survival
Mycolic acid synthesis	<i>mmaA4</i>	Rv0642c	Methoxy mycolic acid synthase 4	C57BL/6 mice (I _{da} /iv)	Reduced CFUs in organs
				C57BL/6 mice (iv) [†]	Failed to persist in the spleens
	<i>pcaA</i>	Rv0470c	Mycolic acid synthase (cyclopropane synthase)	C57BL/6 mice (iv)	Failed to persist in organs Increased animal survival
				C57BL/6 mice (I _{da})	Reduced CFUs in lung
	<i>mymA</i> operon	Rv3083 to Rv3089	Probable Monooxygenase (Hydroxylase)	Activated J774 macrophages and guinea pigs (sc)	Reduced CFUs
	-	Rv2869c	Membrane bound metalloprotease	C57BL/6 mice (I _{da})	Reduced CFUs in lung
	<i>tre5</i>	Rv0126	Trehalose synthase	C57BL/6 mice (iv)	Reduced CFUs in lung Increased animal survival
Synthesis of complex lipids				C57BL/6J mice (in) and MAM MH-S	Reduced CFUs
PDIM	<i>pks15</i> <i>pks1</i>	Rv2946c Rv2947c	Probable polyketide synthases	B6D2 F1 mice (I _{da})	Increased animal survival
				Rabbits (intracisternally)	Reduced CFUs in cerebrospinal fluid and organs
	<i>pks10</i>	Rv1660	Possible chalcone synthase	C57BL/6J mice (in) and MAM MH-S	Reduced CFUs
	<i>pks12</i>	Rv2048c	Probable polyketide synthase	C57BL/6J mice (in) and MAM MH-S	Reduced CFUs
	<i>fadD26</i>	Rv2930	Fatty-acid-Coa synthase	C57BL/6 mice (iv)	Reduced CFUs in lung
				BALB/c mice (iv)	Reduced CFUs in lung
				BALB/c mice (in)	Reduced CFUs in organs
	<i>fadD28</i>	Rv2941	Fatty-acid-Coa synthase	C57BL/6 mice (iv)	Reduced CFUs in lung
	<i>mmpL7</i>	Rv294	Conserved transmembrane transport protein	C57BL/6 mice (iv)	Reduced CFUs in lung
				BALB/c mice (iv)	Reduced CFUs in lung
	<i>drvC</i>	Rv2938	Probable daurubicin-dim-integral membrane ABC-transporter	C57BL/6 mice (I _{da})	Reduced CFUs in organs. Increased animal survival
	<i>pks5</i>	Rv1527c	Probable polyketide synthase	BALB/c mice (I _{da})	Reduced CFUs in organs
	<i>pks7</i>	Rv1661	Probable polyketide synthase	BALB/c mice (I _{da})	Reduced CFUs in organs

Category	Gen Name	Rv number	Description	Attenuation evidences	
				Model	Result
SL	<i>mmpL8</i>	Rv3823c	Probable conserved integral membrane transport protein	C57BL/6 mice (iv)	Reduced CFUs in organs
Others genes related in lipid synthesis	<i>fadD33</i>	Rv1345	Possible polyketide synthase	C57BL/6 and B6D2/F1 mice (Iida)	Increased animal survival
	<i>icl1</i>	Rv0467	Isocitrate lyases	BALB/c mice (iv)	Reduced CFUs in liver
	<i>icl1</i> and <i>icl2</i>	Rv0467 and Rv1915-Rv1916	Isocitrate lyases	C57BL/6 and BALB/c mice (iv). Activated MBMDM	Failed to persist. Increased animal survival Reduced lung pathology
				C57BL/6; IFN- γ ^{-/-} and TNF-R1 ^{-/-} mice (iv). Non-activated and activated BMDM and HBMDM	Reduced CFUs, increased animal survival reduced lung pathology
	<i>plcA</i> <i>plcB</i> <i>plcC</i> <i>plcD</i>	Rv2351c Rv2350c Rv2349c Rv1755c	Probable phospholipase C	BALB/c mice (Iida)	Reduced CFUs in organs
Catabolism of cholesterol	<i>choD</i>	Rv3409c	Putative cholesterol oxidase	C57BL/6 (iv) and mouse peritoneal macrophages	Reduced CFUs
Cell Envelope Proteins	<i>hsaC</i>	<i>Rv3568c</i>	3,4-DHSA dioxygenase	SCID mice (iv)	Increase survival
	<i>igr</i> operon: <i>cyp125</i>	Rv3545c	Putative cytochrome P450	Guinea pig (a)	Modestly reduced CFUs and granulome formation in lungs.
				C57BL/6 mice (a)	Reduced CFUs
	<i>fadE28/29</i>	Rv3544c-Rv3543c	Acyl coenzyme A dehydrogenases		
	-	Rv3542c-Rv3541c	CHPs		
	<i>ltp2</i>	Rv3540c	Probable lipid carrier protein		
<i>erp</i>	Rv3810	Exported repetitive protein	MBMDM and BALB/c mice (iv) [†]	Reduced CFUs in macrophages and organs	
Cell wall proteins	<i>fbpA</i>	Rv3804	Fibronectin binding protein, mycolyltransferase	BALB/c mice (ip)* THP-1 and J774 macrophages	Reduced CFUs in organs. Reduced CFUs.
	<i>mce1</i>	Rv0166 to Rv0174	Mammalian cell entry proteins. Possible lipids ABC-transporters.	BALB/c mice (it) (<i>mce1</i> , 2 and 3 mutants)	Reduced CFUs in organs, reduced tissue pathology and increased animal survival
	<i>mce2</i>	Rv0586-0594		C57BL/6 mice (Iida)	Reduced CFUs and gross lesion in lung. Increased animal survival
	<i>mce3</i>	Rv1964 to Rv1971	Mammalian cell entry proteins. Possible lipids ABC-transporters.	C57BL/6 mice (Iida) (<i>mce3</i> , 4 mutants)	Reduced CFUs in organs and less tissue pathology in lung. Increased survival
	<i>mce4</i>	Rv3501c to Rv3494c	Cholesterol transporter		

Category	Gen Name	Rv number	Description	Attenuation evidences	
				Model	Result
	<i>ompATb</i>	Rv0899	Pore-forming protein	THP1 and MBMDM BALB/c mice (Iida)	Reduced CFUs in macrophages and organs
	<i>hbhA</i>	Rv0475	Heparin binding hemagglutinin protein (Adhesine)	A549 pneumocytes. BALB/c mice (Iida) ^f	Reduced adhesion and CFUs in pneumocytes and reduced CFUs in spleen
	<i>pstA1</i>	Rv0930	Inorganic phosphate-ABC transporter	Resting and activated MBMDM	Reduced CFUs
	<i>phoT</i>	Rv0820		Resting and activated MBMDM and C57BL/6J mice (iv)	Reduced CFUs in macrophages and lung
	<i>caeA</i>	Rv2224c	Carboxylesterase for esters of 3 to 7 carbon atoms	BALB/c mice (Iida)	Reduced CFUs and gross pathology in organs
				BALB/c mice (iv)	Increased survival and weight
				C57BL/6 mice (Iida)	Moderated reduction in CFUs
	<i>kefB</i>	Rv3236c	K ⁺ /H ⁺ antiporter, affecting ROS production	J774 macrophages ^f	Reduced phagosome ROS production
	<i>oppABCD</i>	Rv3666c to Rv3663c	Oligopeptide ABC-transporter	BALB/c mice (Iida)	Reduced CFUs in organs in the chronic infection. Increased survival
	<i>ctaC</i>	Rv2200c	Citrome C oxidase unit II	MBMDM	Reduced CFUs
Lipoproteins	<i>lppX</i>	Rv2945c	Carrier of DIM and antigen	BALB/c mice (Iida)	Reduced CFUs in lung
	<i>lpqH</i>	Rv3763	Antigen Apoptogenic	C57BL/6 mice (Iida)	Reduced CFUs in lung
	<i>lprG</i>	Rv1411c	Cell wall assembly TLR2 agonist	BALB/c mice (ip)*	Reduced bacterial load in spleens
				BALB/c mice (it)*	Reduced CFUs in lung
	<i>lprG-p55</i>	Rv1411c-Rv1410c	Antibiotic efflux pump (P55)	J774macrophages*	Reduced CFUs
	<i>pstS-1</i>	Rv0934	Inorganic phosphate transport. Antigen and apoptogenic	BALB/c and C57BL/6 mice (iv)	Reduced CFUs in organs
				Mouse peritoneal macrophages	Reduced multiplication
	<i>lpqY</i>	Rv1235	ABC-transporter (Recycling system of threalose)	SCID mice (BALB/c mice background) (Iida)	Increased survival
				C57BL/6 mice (Iida)	Reduced UFCs in organs.
	<i>modA</i>	Rv1857	Molybdenum ABC transporter	CBA/J mice (Iida)	Reduced CFUs in ungsand slight increased
				BALB/c mice (iv)	Reduced CFUs in lung
Secretion system	<i>esxA</i>	Rv3875	Esx-1 component or substrate (C or S)	Guinea pigs (sc)*	Reduced CFUs in spleen
	RD1	Rv3868 to Rv3875 and Rv3877	Esx-1 C or S	THP-1 macrophage	Reduced CFUs
				C57BL/6 mice (a)	Reduced CFUs in organs Total survival

Category	Gen Name	Rv number	Description	Attenuation evidences	
				Model	Result
	<i>esxB</i>	Rv3874	Esx-1 C or S	THP-1 macrophage	Reduced CFUs
	<i>espH</i>	Rv3867	Esx-1 C or S	BMDM	Reduced CFUs
				C57BL/6 mice (Iida)	Reduced CFUs in organs
	<i>espG1</i>	Rv3865	Esx-1 C or S	BMDM	Reduced CFUs
				C57BL/6 mice (Iida)	Reduced CFUs in organs Total survival
	<i>espA</i>	Rv3614	Esx-1 C or S	C57BL/6 mice (iv)	Reduced CFUs in organs Increased animal survival
	<i>espC</i>	Rv3615	Esx-1 C or S	BMDM	Reduced CFUs
	<i>eccCd</i>	Rv3877	Esx-1 C or S	BMDM	Reduced CFUs
	<i>espR</i>	Rv3849	Esx-1 C or S	C57BL/6 mice (iv)	Reduced CFUs in lung
	<i>mycP1</i>	Rv3883	Esx-5 C or S	MBMDM	Reduced CFUs
				BALB/c mice (Iida)	Reduced CFUs in lung
	<i>eccD5</i>	Rv1795	Esx-5 C or S	MBMDM	Reduced CFUs
	<i>ppe25 to ppe19</i>	Rv1787 to Rv1791	Signal peptidase for lipoproteins	MBMDM	Reduced CFUs
	<i>IspA</i>	Rv1539	Preprotein translocase ATPase	J774 macrophages	Reduced CFUs
				CBA/J mice	Reduced CFUs in lung
	<i>secA2</i>	Rv1821	Accessory SecA protein	C57BL/6 mice (iv)	Reduced CFUs in organs Increased animal survival
				C57BL/6 -SCID mice (iv)	Increased animal survival
	<i>secA2</i>	Rv1821	Accessory SecA protein	C57BL/6 mice (Iida)	Increased animal survival Reduced CFUs in organs
				BMDM from C57BL/6 mice, NOS2 ^{-/-} and pho ^{-/-} mice (Iida)	Reduced CFUs
Proteins Inhibiting Antimicrobial Effectors of the Macrophage	<i>acr1 (hspX)</i>	Rv2031c	Dormancy-associated protein	MBMDM /THP-1 macrophages	Reduced CFUs
	<i>acr2</i>	Rv0251c	Alpha-crystallin (Acr) family of molecular chaperones	C57BL/6 mice (iv)	Reduced weight loss
Oxidative and nitrosative stresses	-	Rv2136c	Likely involved in the synthesis of peptidoglycan	C57BL/6 mice (Iida)	Reduced CFUs in organs and reduced gross pathology in lung

Category	Gen Name	Rv number	Description	Attenuation evidences	
				Model	Result
	<i>ponA2</i>	Rv3682	Probable transglycosylase and transpeptidase	C57BL/6 mice (I _{da})	Moderated reduction in CFUs
	<i>ahpC</i>	Rv2428	Alkyl hydroperoxide reductase c	Guinea pigs (sc) (Antisense)* J774 macrophages	Reduced tissue pathology and CFUs Reduced CFUs
	<i>sodC</i>	Rv0432	Superoxide dismutase (SOD) protein	Activated murine peritoneal C57BL/6 mice and iNOS ^{-/-} macrophages	Reduced CFUs
	<i>mel2</i>	Rv1936 to Rv1941	Bioluminescence-related proteins	Activated J774, MBMDM from infected mice and human PBMC-derived macrophages	Reduced CFUs
				C57BL/6 and Phox ^{-/-} and iNOS ^{-/-} mice (I _{da})	Moderated reduction of CFUs in organs
				BALB/c mice (iv)	Reduced CFUs and increased survival
				Guinea pigs (im)	Reduced CFUs in spleen and reduced number of lesion in tissues
				C57BL/6 mice (iv)	Reduced CFUs
				NOS2 ^{-/-} mice (iv)	Reduced CFUs
	<i>katG</i>	Rv1908c	Catalase-peroxidase enzyme	BMDM from C57BL/6 and NOS2 ^{-/-} mice	Reduced CFUs
				Activated BMDM NOS2 ^{-/-}	Reduced CFUs
				Guinea pigs*	Moderated reduction in CFUs
				BALB/c mice and MHC class II-knockout mice (ip)	Reduced bacterial load in organs
	<i>tpX</i>	Rv1932	Thiol peroxidase	Resting and activated BMDM from BALB/c and C57BL/6 mice	Reduced CFUs
				BALB/c mice (iv)	Reduced CFUs and increased animal survival
	<i>ndk</i>	Rv2445c	Nucleoside diphosphate kinase	RAW 264.7 macrophages (Antisense) [†]	Reduced CFUs
	<i>ptpA</i>	Rv2234	Low-molecular weight tyrosine phosphatase	THP-1 macrophage	Reduced CFUs
				BALB/c mice (I _{da})	Reduced CFUs
	<i>pe_pgrs30</i>	Rv1651c	Member of the PE family	BALB/c mice (I _{da}) J774.1 and THP-1 macrophages	Reduced tissue damage Reduced CFUs

Category	Gen Name	Rv number	Description	Attenuation evidences	
				Model	Result
Inhibition of apoptosis	<i>nuoG</i>	Rv3151	Subunit of the type I NADH dehydrogenase, NADH-1	BALB/c mice (iv)	Increased animal survival and reduced CFUs in organs
				SCID mice (iv)	Increased animal survival
	<i>pknE</i>	Rv1743	Serine/threonine kinase E	THP-1 macrophages	Reduced bacterial survival
		-		Rv3654c-Rv3655c	CHP
Protein Kinases	<i>pknD</i>	Rv0931c	Protein kinase D	Brain microvascular endothelial cells (HBMEC)	Impaired invasion
				BALB/c mice (iv)	Reduced CFUs in brain
				BALB/c mice (iv)	Increased animal survival
	<i>pknG</i>	Rv0410c	Protein kinase G	BALB/c mice (iv)	Reduced CFUs in organs
SCID mice (iv)				Increased animal survival	
Proteases	<i>mycP1</i>	Rv3883c	Subtilisin-like serine protease.	MBMDM /BALB/c (ld a)	Reduced CFUs in organs
Serine proteases	<i>htrA2 (pepD)</i>	Rv0983	HtrA-like serine protease and chaperone.	BALB/c, SCID and C57BL/6 mice(iv)	Increased animal survival and less tissue pathology
		-		Rv3671c	Serine protease
ATP-dependent proteases	<i>clgR</i>	Rv2745c	Transcriptional regulator	MBMDM	Reduced CFUs in organs
Metallo-proteases	<i>zmp1</i>	Rv0198c	Zn ²⁺ Metallo-protease	J774and RAW264.7 macrophages	Reduced CFUs in organs
				BALB/c mice (lda)	
				THP1 macrophage	No differences in CFU's
				SCID mice (iv)	Reduced survival
<i>rip1</i>	Rv2869c	S2P class of metallo-proteases	C57BL/6 mice (lda)	Increased CFU's in organs	
			C57BL/6 mice (a)	Reduced CFUs in organ and reduced tissue pathology	
Proteasome-associated proteins	<i>pafA</i>	Rv2097c	Mycobacterial proteasomal ATPase	C57BL/6 and iNOS ^{-/-} mice	Reduced CFUs in organs and pathology
	<i>mpa</i>	Rv2115c	Mycobacterial proteasomal ATPase	Resting BMDM from both	
Metals-Transporter Proteins	<i>mbtB</i>	Rv2383c	Iron ABC Transporter	MBMDM	Reduced CFUs in organs and less tissue pathology. Increased animal survival
				BALB/c mice (lda)	
				Macrophage-like cell line THP-1.	Reduced CFUs and retarded for ability to grow in this cell line.

Category	Gen Name	Rv number	Description	Attenuation evidences	
				Model	Result
Metal importers	<i>irtAB</i>	Rv1348-Rv1349	Iron-dependent regulatory protein. Repressor of <i>mtb</i> and <i>mtb-2</i> loci.	THP-1 macrophages. C57B/6 mice (Iida)	Reduced CFUs in macrophages and lung
	<i>ideR</i>	Rv2711		-	Essential
	<i>mgtC</i>	Rv1811	Mg ²⁺ transport P-type ATPase. Mg ²⁺ uptake	HBMDM. BALB/c mice (iv)	Reduced CFUs in macrophages and organs
Metal exporters	<i>ctpC</i>	Rv3270	Zn ²⁺ efflux transporter P-type ATPase	Human macrophages	Reduced CFUs
	<i>ctpV</i>	Rv0969	Cu ²⁺ efflux transporter P-type ATPase	BALB/c mice (Iida)	Increased survival and lower tissue damage in lung
				Guinea pigs (Iida)	Reduced CFUs and lower tissue damage in lung
Gene Expression Regulators	<i>phoPR</i>	Rv0757 to Rv0758	TCS	THP1 macrophages	Reduced CFUs
				C57BL/6 MICE (Iida)	Reduced CFUs in lung
Two component system (TCS)	<i>phoP</i>	Rv0757	Transcriptional regulator	MBMDM	Reduced CFUs
				BALB/c mice (iv)	Reduced CFUs
				SCID mice (Iida/iv)	Improved survival
	<i>aprABC</i>	Rv2396abc	Expressed in acidic medium dependent on PhoP	Resting and activated C57BL/6 BMDM	Defects in intracellular replication
	<i>senX3-regX3</i>	Rv0490-Rv0491	TCS	THP-1 macrophages	Reduced CFUs
				DBA mice	Moderate reduced CFUs in lung
	<i>senX3</i>	Rv0490	Sensor	BALB/c mice (iv)	Reduced CFUs in lung
	<i>regX3</i>	Rv0491	Transcriptional Regulator	BALB/c mice (iv)	Reduced CFUs in lung
	<i>dosRS</i> <i>dosT</i>	Rv3133c-Rv3132c Rv2027c	TCS	C57BL/6 MICE (Iida)	Reduced lung pathology. Increased CFUs
	<i>dosR (devR)</i>	Rv3133c	Transcriptional Regulator	C57BL/6 mice (Iida)	Moderate reduction CFUs in lung
				Guinea pigs (Iida)	Reduction CFUs in lung
				Rabbit (Iida)	Moderate reduction CFUs in lung
				Guinea pigs (sc)	Reduced lung pathology
	<i>mprAB</i>	Rv0981-Rv0982	TCS	BALB/c mice (iv)	Reduced CFUs in lung latent stage

Category	Gen Name	Rv number	Description	Attenuation evidences	
				Model	Result
Sigma factors	<i>sigA</i>	Rv2703	Sigma factor A	C57BL/6 mice (Iida) (Antisense)	Reduced CFUs in lung
				MonoMac6 cells (Antisense)	Reduced CFUs
				DBA/2 mice (Iida)	Modest reduction of CFUs in lung, increased animal survival and reduced lung pathology
	<i>sigC</i>	Rv2069	Sigma factor C	SCID mice (Iida)	Modest reduction of CFUs in lung and increased animal survival
				Guinea pigs (Iida)	Reduced lung pathology
				DBA/2 mice (Iida/iv)	Increased survival and reduced lung pathology
				Guinea pigs (ida)	Reduced lung and spleen pathology
	<i>sigD</i>	Rv3414c	Sigma factor D	C3H mice (iv)	Increased animal survival
				BALB/c mice (iv)	Increased animal survival
				SCID mice	Increased animal survival
				BALB/c mice (iv)	Reduced CFUs in organs
	<i>sigE</i>	Rv1221	Sigma factor E	C3H/HeJ mice (a)	Increased animal survival
				Unactivated THP-1 and J774 or activated J774 macrophages	Reduced CFUs
	<i>sigF</i>	Rv3286c	Sigma factor F	BALB/c mice (iv)	Increased animal survival
				BALB/c mice (iv)	Reduced CFUs and pathology in tissues
				Guinea pigs (ida)	Moderated reduction in organ pathology
	<i>sigG</i>	Rv0182c	Sigma factor G	J774macrophages	Moderated reduction in CFUs
<i>sigH</i>	Rv3223c	Sigma factor H	C3H and C57BL/6 mice	Reduced pathology in tissues	
			C3H:He mice	Increased animal survival	
			BMDM from rhesus monkey	Reduced CFUs at late time post infection	
<i>sigL</i>	Rv0735	Sigma factor L	BALB/c mice (iv)	Increased animal survival	

Category	Gen Name	Rv number	Description	Attenuation evidences		
				Model	Result	
Other transcriptional regulators	-	Rv0485	Member of the nagc/xylr repressor family	BALB/c mice (iv)	Reduced lung pathology	
				BALB/c mice (iv)	Modestly increased survival	
				SCID (iv)	Modestly increased survival	
	-	Rv1931c	AraC transcriptional regulator	BALB/c mice (iv) and BMDM from BALB/c mice	Reduced CFUs	
	<i>hspR</i>	Rv0353	Transcriptional repressor	C57BL/6 mice (iv)	Reduced CFUs in organs	
	<i>whiB3</i>	Rv3416	Whib-like regulator family	C57BL/6 mice (iv)*	Increased animal survival	
	<i>mosR</i>	Rv0348	Mycobacterial operons of survival regulator	BALB/c mice (Ida)	Reduced CFUs	
	<i>virS</i>	Rv3082c	AraC family of transcriptional regulator	BALB/c mice (Ida)	Increased animal survival	
	<i>phoY2</i>	Rv0821c	Probable phosphate-transport system transcriptional regulator	Activated J774 macrophage. Guinea pig (sc)	Reduced CFUs	
Proteins of Unknown Function	<i>pe_pgrs33</i>	Rv1818c	PE_PGRS family protein	BALB/c mice (iv)	Reduced CFUs	
The PE/PPE families	<i>pe_pgrs51</i>	Rv3367	PE_PGRS family	J774 and BMDM macrophages [†]	Reduced CFUs	
	<i>ppe46</i>	Rv3018	PPE family	BALB/c mice (iv)	Moderate reduced CFUs in lung	
	-	Rv1099c	CHP	C57BL/6J mice (iv)	Reduced CFUs in organs	
	-	Rv0573c	CHP	C57BL/6J mice (iv)	Reduced CFUs in organs	
Others proteins with unknown function	-	Rv0204c	Integral membrane protein	BALB/c mice (iv)	Moderate reduced CFUs in lung	
	-	Rv2452c	Hypothetical proteins	BALB/c mice (iv)	Moderate reduced CFUs in lung	
	-	Rv1290c	CHP	CB-17/1cr SCID mice (iv)	Markedly increased survival	
	-	Rv1891	CHP	CB-17/1cr SCID mice (iv)	Moderate increased survival	
	-	Rv3404c	CHP	CB-17/1cr SCID mice (iv)	Moderate increased survival	
	-	Rv1503c to Rv1507c	CHP	BALB/c mice (Ida)	Markely reduced CFUs in organs	
	-	Rv0199	Conserved membrane protein	MBMDM	Reduced CFUs	
		<i>mmp14</i>	Rv0450c	Conserved membrane transport protein	C57BL/6 and (C57BL/6 x DBA2) F1 mice (a)	Reduced CFUs in organs and increased life survival
	-	Rv2136c	Conserved transmembrane protein	C57BL/6 mice (Ida)	Markedly reduced CFUs in organs. Decreased gross pathology in lung	

Category	Gen Name	Rv number	Description	Attenuation evidences	
				Model	Result
Other Virulence Factors	<i>RD2</i>	Rv1979c to Rv1982	Region of Difference	C57BL/6 mice (I _{da})	Reduced CFUs in lung and tissue pathology
				RAW macrophages	Reduced CFUs at late times
	<i>acg</i>	Rv2032	Uncertain	BALB/c mice (iv)	Reduced CFUs
				SCID mice (iv)	Increased animal survival
				BMDM from BALB/c mice	Reduced CFUs
	<i>pckA</i>	Rv0211	Phosphoenolpyruvate carboxykinase	BMDM from BALB/c mice [†]	Reduced CFUs
	<i>ptpB</i>	Rv0153c	Tyrosine phosphatase	BALB/c mice (iv) [†]	Reduced CFUs in spleen
				Guinea pigs	Reduced CFUs in spleen
	<i>hsp22.5</i>	Rv0990c	Novel heat shock protein	Activated J774macrophages	Reduced CFUs
				BALB/c mice (a)	Reduced CFUs in organs and increased animal survival

Supplementary Table: Principal MTB virulence factors ¹²

RELAZIONE SULL'ATTIVITÀ SVOLTA DA MATTEO CHIACCHIARETTA

Il dottorando Matteo Chiacchiaretta, iscritto al Dottorato in Biotecnologie Mediche dell'Università di Siena, ciclo XXXIV, ha svolto durante il corso di dottorato le seguenti attività:

ATTIVITÀ DI RICERCA

L'attività di ricerca del Dr. Matteo Chiacchiaretta si è fondamentalmente orientata allo studio di caratterizzazione della variabilità genetica di *M. tuberculosis* e del suo coinvolgimento nelle infezioni e nei meccanismi di tolleranza ai farmaci antitubercolari

Gli argomenti trattati sono stati:

- *Host-pathogen interaction* tra ceppi dell' MTB complex e macrofagi pro ed anti-infiammatori
- Caratterizzazione dello smallRNA *ncRv0842c* di MTB coinvolto nella risposta alla rifampicina, e il suo ruolo in diversi ceppi dell'MTB complex
- Dati preliminari sullo smallRNA *nc0757c* coinvolto nella regolazione del gene *phoP* in diversi lineage dell' MTB complex

TESI DI DOTTORATO

Titolo: M. tuberculosis lineages: genetic diversity and its involvement on macrophage infection and on drug tolerance

Tutore: Daniela Maria Cirillo

Co-tutore: Paolo Miotto

Descrizione delle tematiche e dei principali risultati

Lo studio ha permesso di indagare la variabilità genetica dei ceppi clinici dell'MTB complex e di correlarla ai meccanismi di patogenesi e di *drug-resistance acquisition*. I risultati hanno mostrato come sia per quanto riguarda la virulenza, sia per quanto riguarda la regolazione di geni importanti per la tolleranza ai farmaci, esista una variabilità non trascurabile tra i ceppi di MTB, che va ad influenzare drasticamente l'esito dell'infezione. Lo studio ha permesso di indagare con diverse strategie in contemporanea (*single-cell level*, produzione di citochine, sopravvivenza) le differenze che esistono tra i ceppi selezionati, tenendo anche in considerazione lo stato immunologico dell'ospite. Per la prima volta, è stato studiato uno smallRNA coinvolto nella regolazione di un gene importante per i meccanismi di tolleranza ai farmaci. Lo studio ha dimostrato l'importanza di mutazioni sinonime nell'espressione degli smallRNA e la differenza che esiste tra i ceppi dell'MTB complex.

PARTECIPAZIONE A CONGRESSI, SCUOLE E CORSI

- Matteo Chiacchiaretta, Rita Sorrentino, Federica Cugnata, Alessandra Aiello, Assunta Navarra, Valentina Vanini, Daniela M. Cirillo, Delia Goletti, Paolo Miotto. Interplay between macrophage heterogeneity and Mycobacterium tuberculosis complex phylogenetic lineages during in vitro infection 23-26 April 2022 (accepted abstract) – “32st European Congress of Clinical Microbiology and Infectious Diseases (ECCMID)”
- Russo G, Di Marco F, Chiacchiaretta M, Spitaleri A, Ambrosi A, Goletti D, Codecasa LR, Miotto P*, Cirillo DM. Serum human microRNAs in the differential diagnosis between active tuberculosis and tuberculosis infection. (19-22 October 2021 – “52nd Union World Conference on Lung Health – Lung Health for All, Solutions for a New Era. Joint meeting with TBScience 2021”, virtual event)
- Chiacchiaretta M, Sorrentino R*, Spitaleri A, Cirillo DM, Miotto P. ncRv0842c, a smallRNA regulating the efflux pump Rv0842 involved in rifampicin tolerance in Mycobacterium tuberculosis. (9-12 July 2021 – “31st European Congress of Clinical Microbiology and Infectious Diseases (ECCMID)”, virtual event)
- Cabibbe AM, Vanoni C, Chiacchiaretta M, Battaglia S, Spitaleri A, Cugnata F, Miotto P*, Cirillo DM. Epidemiological dynamics and characterization of a successful pre-XDR Mycobacterium tuberculosis clone actively transmitted in Italy. (9-12 July 2021 – “31st European Congress of Clinical Microbiology and Infectious Diseases (ECCMID)”, virtual event)
- Chiacchiaretta M, Sorrentino R, Audisio A, Goletti D, Vanini V, Aiello A, Cirillo DM, Miotto P. Interplay between macrophage polarization and M. tuberculosis lineages. (28-29 June 2021 – “41st Annual Congress of the European Society of Mycobacteriology”, virtual event)
- Battaglia S, Chiacchiaretta M, Gabro VS, Robinson E, Cirillo DM, Köser CU, Schön T, Miotto P. Lineage 1 has elevated pyrazinamide minimum inhibitory concentrations compared with other lineages of Mycobacterium tuberculosis complex. (28-29 June 2021 – “41st Annual Congress of the European Society of Mycobacteriology”, virtual event)

- Cabibbe AM*, Vanoni C, Chiacchiaretta M, Battaglia S, Spitaleri A, Cugnata F, Miotto P, Cirillo DM. In vitro characterization of a successful multidrug-resistant Mycobacterium tuberculosis clone actively transmitted in Italy. (28-29 June 2021 – “41st Annual Congress of the European Society of Mycobacteriology”, virtual event)
- Chiacchiaretta M, Bresciani N, Agresti A, Zambrano S, Mazza D, Cugnata F, Tassan Din C, Cirillo DM, Miotto P*. Classically (M1) and alternatively (M2) activated macrophages infected with generalists and specialists Mycobacterium tuberculosis complex isolates display differential phagosomal acidification patterns. (2-4 December 2020 – “Keystone eSymposium “Tuberculosis: science aimed at ending the epidemic”, virtual event)
- Chiacchiaretta M, Spitaleri A, Cirillo DM, Miotto P. ncRv0842c, a smallRNA regulating the efflux pump Rv0842 involved in rifampicin tolerance in Mycobacterium tuberculosis. (20-24 October 2020 – “51st Union World Conference on Lung Health – TBScience 2020”, virtual event)
- Chiacchiaretta M, Bresciani N, Russo G, Cirillo DM*, Miotto P. Characterization of the intramacrophagic expression profile of the smallRNA ncRv0757c in different Mycobacterium tuberculosis lineages. (29-30 October 2019 – “2nd TBScience meeting – The 50th Union World Conference on Lung Health”, Hyderabad, IND)
- Chiacchiaretta M*, Bresciani N, Russo G, Tassan Din C, Cirillo DM, Miotto P. Characterization of the intramacrophagic expression profile of the smallRNA ncRv0757c in different Mycobacterium tuberculosis lineages (30 June - 3 July 2019 – “40th Annual Congress of the European Society of Mycobacteriology”, Valencia, E)
- Chiacchiaretta M, Bresciani N, Agresti A, Zambrano S, Mazza D, Cugnata F, Tassan Din C, Cirillo DM, Miotto P*. The role of Mycobacterium tuberculosis complex genetic variation and of macrophage phenotype during early stages of infection (30 June - 3 July 2019 – “40th Annual Congress of the European Society of Mycobacteriology”, Valencia, E)
- Chiacchiaretta M*, Bresciani N, Agresti A, Zambrano S, Mazza D, Cugnata F, Tassan Din C, Cirillo DM, Miotto P. Mycobacterium tuberculosis complex genetic variation and macrophage polarization

status influence phagolysosomal acidification during infection (5-7 June 2019 – “11th Italian Conference on AIDS and Antiviral Research”, Milano, I)

· Chiacchiaretta M*, Bresciani N, Russo G, Tassan Din C, Cirillo DM, Miotto P. Characterization of the intramacrophagic expression profile of the smallRNA ncRv0757c in different Mycobacterium tuberculosis lineages. (16-18 March 2019 – “San Raffaele Scientific Retreat”, Baveno, I)

· Tassan Din C, Chiacchiaretta M, Bresciani N, Agresti A, Zambrano S, Mazza D, Cugnata F, Cirillo DM, Miotto P*. The role of Mycobacterium tuberculosis complex genetic variation during macrophage infection. (14-16 March 2019 – “San Raffaele Scientific Retreat”, Baveno, I)

· Chiacchiaretta M, Bresciani N, Agresti A, Zambrano S, Mazza D, Cugnata F, Tassan Din C, Cirillo DM, Miotto P*. Lysosomal acidification induced by different Mycobacterium tuberculosis complex lineages during human macrophage infection. (17-21 January 2019 – “Keystone Symposium on tuberculosis: Tuberculosis: Mechanisms, Pathogenesis and Treatment”, Banff – AB, CDN)

Chiacchiaretta M, Bresciani N, Agresti A, Zambrano S, Tassan Din C, Cirillo DM, Miotto P. Single-cell live confocal microscopy for monitoring lysosomal acidification induced by different M. tuberculosis complex lineages during macrophage infection. (1-4 July 2018 – “39th Annual Congress of the European Society of Mycobacteriology”, Dresden, D)

PUBBLICAZIONI

Durante il periodo del Dottorato il Dr Matteo Chiacchiaretta ha prodotto 2 pubblicazioni accettate e 3 manuscript in submission.

Articoli pubblicati:

· Gona F, Bongiorno D, Aprile A, et al. Emergence of two novel sequence types (3366 and 3367) NDM-1- and OXA-48-co-producing *K. pneumoniae* in Italy. *Eur J Clin Microbiol Infect Dis*. 2019;38(9):1687-1691. doi:10.1007/s10096-019-03597-w

· Fowler PW, Gibertoni Cruz AL, Hoosdally SJ, et al. Corrigendum: Automated detection of bacterial growth on 96-well plates for high-throughput drug susceptibility testing of Mycobacterium tuberculosis. Microbiology (Reading). 2019;165(5):585. doi:10.1099/mic.0.000795

In particolare sulle tematiche connesse alla ricerca svolta nell'ambito del dottorato ha pubblicato i seguenti articoli:

Manuscripts under submission:

· Chiacchiaretta M, Sorrentino R, Bresciani N, Agresti A, Zambrano S, Mazza D, Tassan Din C, Cugnata F, Aiello A, Navarra A, Goletti D, Cirillo DM, Miotto P. Classically (M1) and alternatively (M2) activated macrophages infected with generalists and specialists Mycobacterium tuberculosis complex isolates display differential phagosomal acidification patterns

· Chiacchiaretta M, Sorrentino R, Di Marco F, Spitaleri A, Cirillo DM, Miotto P *ncRv0842c*, a smallRNA regulating the efflux pump Rv0842 involved in rifampicin tolerance in Mycobacterium tuberculosis

· Lineage 1 has elevated pyrazinamide minimum inhibitory concentrations compared with other lineages of Mycobacterium tuberculosis complex

Data 28/01/2022

Firma del dottorando



Visto

(Il Tutore)



Phagolysosomal acidification

LysoSensor

Wells/sample	3
Acquisition positions/well	2
Channels	Hoechst, LysoSensor, dsRed
Z-stacks	0.89 μm x7

6 h p.i.

24 h p.i.

	NOT infected		Infected		TOT	
	N of cells	Replicates	N of cells	Replicates	Not inf	Inf
M1	CTRL	197	8	na	na	
	H37Rv	130	3	116	3	
	H37Ra	119	3	105	3	
	BCG	78	2	84	2	
	CDC1551	129	3	122	3	1141
	Beijing	182	5	199	5	909
	EAI	146	4	137	4	
	Afri	160	4	146	4	
M2	CTRL	190	8	na	na	
	H37Rv	111	3	113	3	
	BCG	73	2	84	2	
	H37Ra	131	3	121	3	
	CDC1551	134	3	123	3	1116
	Beijing	186	5	218	5	932
	EAI	142	4	134	4	
	Afri	149	4	139	4	
TOT	2257		1841		4098	

	NOT infected		Infected		TOT	
	N of cells	Replicates	N of cells	Replicates	Not inf	Inf
M1	CTRL	152	8	na	na	
	H37Rv	77	3	110	3	
	H37Ra	73	3	111	3	
	BCG	51	2	76	2	
	CDC1551	75	3	91	3	813
	Beijing	129	5	169	5	806
	EAI	136	4	127	4	
	Afri	120	4	122	4	
M2	CTRL	138	8	na	na	
	H37Rv	87	3	105	3	
	BCG	70	2	80	2	
	H37Ra	94	3	109	3	
	CDC1551	84	3	82	3	859
	Beijing	126	5	162	5	808
	EAI	141	4	141	4	
	Afri	119	4	129	4	
TOT	1672		1614		3286	

Total

	NOT infected		Infected		TOT	
	N of cells	Replicates	N of cells	Replicates	Not inf	Inf
M1	CTRL	349	8	na	na	
	H37Rv	207	3	226	3	
	H37Ra	192	3	216	3	
	BCG	129	2	160	2	
	CDC1551	204	3	213	3	1954
	Beijing	311	5	368	5	1715
	EAI	282	4	264	4	
	Afri	280	4	268	4	
M2	CTRL	328	8	na	na	
	H37Rv	198	3	218	3	
	BCG	143	2	164	2	
	H37Ra	225	3	230	3	
	CDC1551	218	3	205	3	1975
	Beijing	312	5	380	5	1740
	EAI	283	4	275	4	
	Afri	268	4	268	4	
TOT	3929		3455		7384	

Acquisition:

obj. 63x, oil NA 1.40
z-stacks: 0.89 μm x 7 stacks
Res. 16-bit format: 1024x1024
Laser 405 (for Hoechst) = 6%; gain: 898.9, offset: 0%
Laser 458 (for LysoSensor) = 10% (Argon power: 30%); gain: 724.8, offset: 0%
Laser 453 (for dsRed) = 15%; gain: 844.9, offset: 0%
Confocal: AP 24 FD 6 O INT 0 FIM 100%

Analysis:

4 h p.i. and 24 h p.i.; (N of phagolysosomes), phagolysosomes intensity and % of active cells, MOI, colocalization myc/acidic compartments

Hoechst (Ex/Em 350/461)

threshold (Huang, Red; no flags on Dark background and Stack histogram) = approx. 215
size = 40-inf μm^2 circularity = 0-1 (Show: outlines; flag only Add to Manager)

LysoSensor (Ex/Em 443/505)

threshold (Huang, Red; no flags on Dark background and Stack histogram) = 155
size = 0.5-1.8 μm^2 circularity = 0-1 (Show: overlay mask; NO flag on Add to Manager)

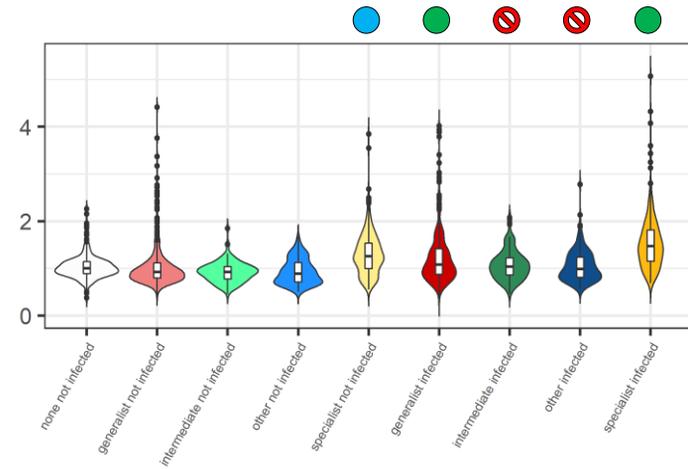
dsRed (Ex/Em 584-557)

threshold (Huang, Red; no flags on Dark background and Stack histogram) = 215
size = 2-100 μm^2 circularity = 0-0.9 (Show: overlay mask; NO flag on Add to Manager)

Grouping: Geo-host adaptation

M1 – 6 h p.i.

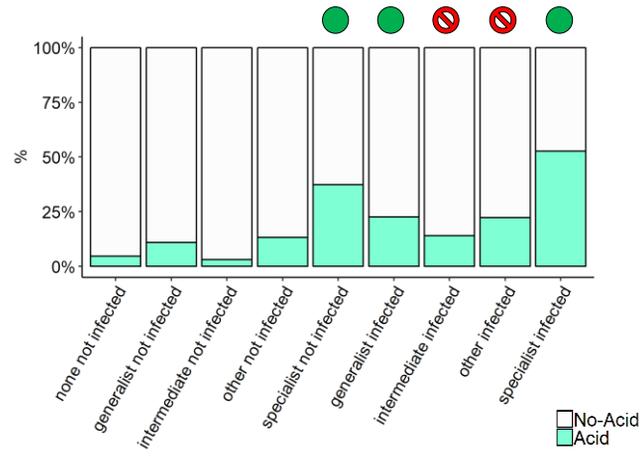
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	0.0034	0.0759	0.9648
groupgeneralist not infected	0.0463	0.0853	0.4025
groupintermediate not infected	-0.0365	0.0769	0.6354
grouphother not infected	-0.1016	0.0654	0.1208
groupspecialist not infected	0.1075	0.057	0.0597
groupgeneralist infected	0.1539	0.0553	0.0055
groupintermediate infected	0.1132	0.0771	0.142
grouphother infected	-0.0272	0.0656	0.6783
groupspecialist infected	0.2576	0.0572	0

Contrast	estimate	SE	p.value
none not infected-generalist not infected	-0.0463	0.0553	0.483
none not infected-intermediate not infected	0.0365	0.0769	0.663
none not infected-other not infected	0.1016	0.0654	0.1812
none not infected-specialist not infected	-0.1075	0.057	0.1303
none not infected-generalist infected	-0.1539	0.0553	0.0164
none not infected-intermediate infected	-0.1132	0.0771	0.2005
none not infected-other infected	0.0272	0.0656	0.6783
none not infected-specialist infected	-0.2576	0.0572	0
generalist not infected-intermediate not infected	0.0827	0.0749	0.3597
generalist not infected-other not infected	0.1478	0.0673	0.0675
generalist not infected-specialist not infected	-0.0612	0.0586	0.3748
intermediate not infected-other not infected	0.0651	0.0872	0.52
intermediate not infected-specialist not infected	-0.1439	0.0798	0.1317
other not infected-specialist not infected	-0.2091	0.0674	0.0067
generalist infected-intermediate infected	0.0407	0.0751	0.6414
generalist infected-other infected	0.1811	0.0675	0.0196
generalist infected-specialist infected	-0.1037	0.0588	0.1337
intermediate infected-other infected	0.1404	0.0874	0.1734
intermediate infected-specialist infected	-0.1445	0.0801	0.1317
other infected-specialist infected	-0.2849	0.0676	1e-04
generalist not infected-generalist infected	-0.1077	0.0141	0
intermediate not infected-intermediate infected	-0.1497	0.022	0
other not infected-other infected	-0.0744	0.0177	1e-04
specialist not infected-specialist infected	-0.1502	0.0143	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.5823	0.8396	0.0278	0
groupgeneralist not infected	0.6037	0.6792	1.8289	0.3741
groupintermediate not infected	0.0514	0.9665	1.0527	0.9576
grouphother not infected	0.3002	0.735	1.3502	0.6829
groupspecialist not infected	1.5245	0.6226	4.5927	0.0143
groupgeneralist infected	2.1602	0.6538	8.6725	0.001
groupintermediate infected	1.826	0.862	6.2087	0.0341
grouphother infected	0.9688	0.7215	2.6349	0.1793
groupspecialist infected	2.675	0.6242	14.513	0

Contrast	estimate	SE	p.value
none not infected-generalist not infected	-0.6037	0.6792	0.4988
none not infected-intermediate not infected	-0.0514	0.9665	0.9576
none not infected-other not infected	-0.3002	0.735	0.745
none not infected-specialist not infected	-1.5245	0.6226	0.0492
none not infected-generalist infected	-2.1602	0.6538	0.0057
none not infected-intermediate infected	-1.826	0.862	0.1024
none not infected-other infected	-0.9688	0.7215	0.3074
none not infected-specialist infected	-2.675	0.6242	1e-04
generalist not infected-intermediate not infected	0.5523	0.9195	0.6923
generalist not infected-other not infected	0.3035	0.7255	0.745
generalist not infected-specialist not infected	-0.2489	0.5871	0.2157
intermediate not infected-other not infected	-0.2489	1.0007	0.8385
intermediate not infected-specialist not infected	-1.4731	0.9031	0.2057
other not infected-specialist not infected	-1.2242	0.6569	0.1497
generalist infected-intermediate infected	0.3342	0.7838	0.745
generalist infected-other infected	1.1913	0.6837	0.1777
generalist infected-specialist infected	-0.5149	0.5613	0.4988
intermediate infected-other infected	0.8571	0.887	0.4988
intermediate infected-specialist infected	-0.8491	0.7907	0.4527
other infected-specialist infected	-1.7962	0.6456	0.0329
generalist not infected-generalist infected	-1.5563	0.339	1e-04
intermediate not infected-intermediate infected	-1.7746	0.5888	0.0124
other not infected-other infected	-0.6686	0.3247	0.1052
specialist not infected-specialist infected	-1.1506	0.2256	0

OBSERVATIONS

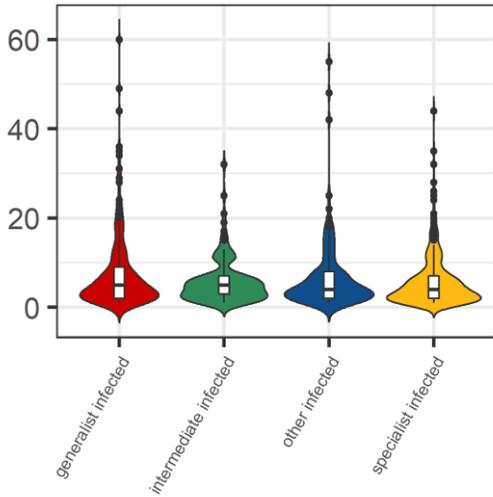
In M1 macrophages, only categories intermediate and other displayed blocking of phagolysosomal acidification at 6 h p.i., and specialist strains showed the highest number of acidified macrophages (50% vs <25% of all the other categories). A bystander effect in non-infected M1 macrophages was observed for specialist strains at 6 h p.i, despite it was not found statistically significant.

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

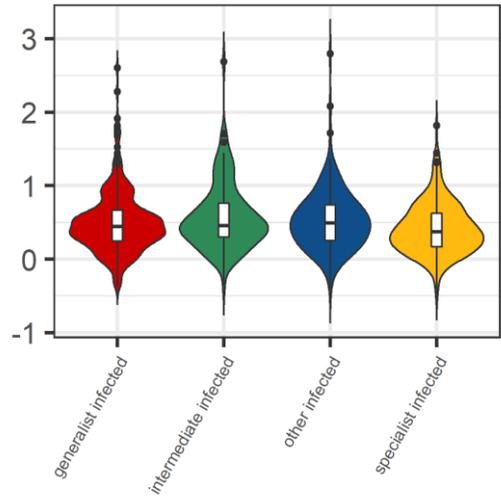
Grouping: Geo-host adaptation

M1 – 6 h p.i.

c) Effective MOI



d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	1.4895	0.1074	0
groupintermediate infected	-0.1421	0.1621	0.3841
groupother infected	-0.0297	0.1467	0.8405
groupspecialist infected	-0.0751	0.1277	0.5587

Contrast	estimate	SE	p.value
generalist infected - intermediate infected	0.1421	0.1621	0.8405
generalist infected - other infected	0.0297	0.1467	0.8405
generalist infected - specialist infected	0.0751	0.1277	0.8405
intermediate infected - other infected	-0.1124	0.188	0.8405
intermediate infected - specialist infected	-0.067	0.1726	0.8405
other infected - specialist infected	0.0454	0.1459	0.8405

Par	Value	Std.Error	p.value
(Intercept)	0.0473	0.08	0.5543
groupintermediate infected	0.1246	0.1341	0.3566
groupother infected	0.1538	0.121	0.2084
groupspecialist infected	-0.1465	0.1058	0.1712

Contrast	estimate	SE	p.value
generalist infected - intermediate infected	-0.1246	0.1341	0.428
generalist infected - other infected	-0.1538	0.121	0.3126
generalist infected - specialist infected	0.1465	0.1058	0.3126
intermediate infected - other infected	-0.0292	0.1519	0.8482
intermediate infected - specialist infected	0.2711	0.1402	0.1733
other infected - specialist infected	0.3003	0.1198	0.0893

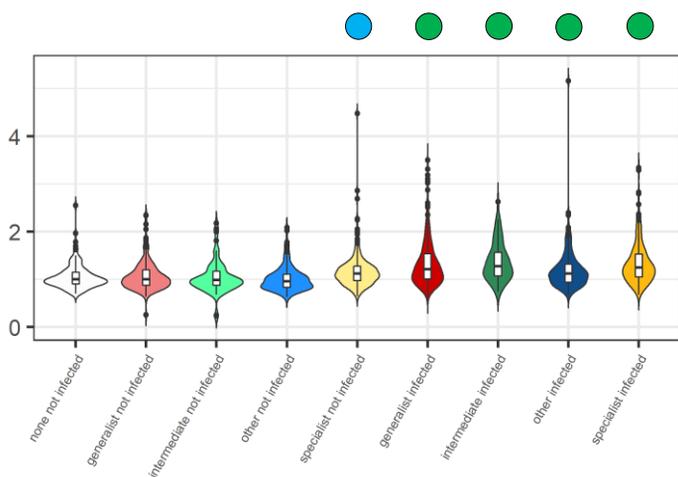
OBSERVATIONS

In M1 macrophages, no statistically relevant differences in effective MOI were observed. Colocalization with acidified compartments was similar among all the groups.

Grouping: Geo-host adaptation

M1 – 24 h p.i.

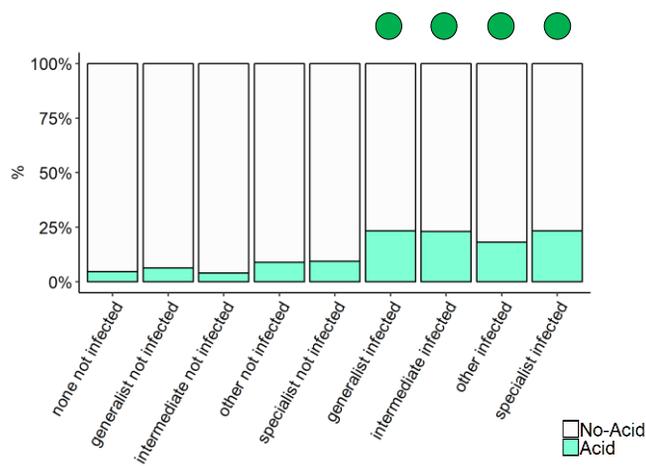
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	0.0091	0.0429	0.8315
groupgeneralist not infected	0.0412	0.0323	0.2028
groupintermediate not infected	0.0206	0.0444	0.643
grouphother not infected	-0.0215	0.0381	0.5724
groupspecialist not infected	0.0263	0.0323	0.416
groupgeneralist infected	0.1879	0.0318	0
groupintermediate infected	0.2727	0.0435	0
grouphother infected	0.1259	0.0367	6e-04
groupspecialist infected	0.1353	0.0324	0

Contrast	estimate	SE	p.value
none not infected-generalist not infected	-0.0412	0.0323	0.3042
none not infected-intermediate not infected	-0.0206	0.0444	0.7014
none not infected-other not infected	0.0215	0.0381	0.7014
none not infected-specialist not infected	-0.0263	0.0323	0.5547
none not infected-generalist infected	-0.1879	0.0318	0
none not infected-intermediate infected	-0.2727	0.0435	0
none not infected-other infected	-0.1259	0.0367	0.0019
none not infected-specialist infected	-0.1353	0.0324	1e-04
generalist not infected-intermediate not infected	0.0206	0.0426	0.7014
generalist not infected-other not infected	0.0627	0.0377	0.1653
generalist not infected-specialist not infected	0.0149	0.0321	0.7014
intermediate not infected-other not infected	0.0421	0.0492	0.5538
intermediate not infected-specialist not infected	-0.0057	0.0446	0.8977
other not infected-specialist not infected	-0.0478	0.0372	0.3042
generalist infected-intermediate infected	-0.0848	0.0412	0.0864
generalist infected-other infected	0.062	0.0358	0.1653
generalist infected-specialist infected	0.0526	0.0316	0.1653
intermediate infected-other infected	0.1468	0.0473	0.0047
intermediate infected-specialist infected	0.1374	0.0438	0.0047
other infected-specialist infected	-0.0093	0.036	0.8296
generalist not infected-generalist infected	-0.1467	0.0191	0
intermediate not infected-intermediate infected	-0.2521	0.0325	0
other not infected-other infected	-0.1475	0.0236	0
specialist not infected-specialist infected	-0.109	0.0181	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.1976	0.4593	0.0409	0
groupgeneralist not infected	0.3895	0.5143	1.4763	0.4488
groupintermediate not infected	0.1034	0.7496	1.109	0.8902
grouphother not infected	0.9072	0.5479	2.4774	0.0977
groupspecialist not infected	0.5002	0.4771	1.649	0.2945
groupgeneralist infected	1.9578	0.4494	7.0841	0
groupintermediate infected	2.0397	0.5309	7.6883	1e-04
grouphother infected	1.7688	0.4842	5.8637	3e-04
groupspecialist infected	1.6527	0.4506	5.2213	2e-04

Contrast	estimate	SE	p.value
none not infected-generalist not infected	-0.3895	0.5143	0.6732
none not infected-intermediate not infected	-0.1034	0.7496	0.8902
none not infected-other not infected	-0.9072	0.5479	0.2607
none not infected-specialist not infected	-0.5002	0.4771	0.5468
none not infected-generalist infected	-1.9578	0.4494	1e-04
none not infected-intermediate infected	-2.0397	0.5309	7e-04
none not infected-other infected	-1.7688	0.4842	0.001
none not infected-specialist infected	-1.6527	0.4506	0.001
generalist not infected-intermediate not infected	0.2861	0.6951	0.8168
generalist not infected-other not infected	-0.5177	0.4955	0.5468
generalist not infected-specialist not infected	-0.1106	0.4089	0.8583
intermediate not infected-other not infected	-0.8038	0.7382	0.5468
intermediate not infected-specialist not infected	-0.3967	0.6866	0.728
other not infected-specialist not infected	0.407	0.4483	0.5822
generalist infected-intermediate infected	-0.0818	0.3691	0.8604
generalist infected-other infected	0.1891	0.3384	0.728
generalist infected-specialist infected	0.3051	0.2853	0.5468
intermediate infected-other infected	0.2709	0.4453	0.728
intermediate infected-specialist infected	0.387	0.4073	0.5822
other infected-specialist infected	0.116	0.3328	0.8313
generalist not infected-generalist infected	-1.5683	0.3303	0
intermediate not infected-intermediate infected	-1.9363	0.646	0.0093
other not infected-other infected	-0.8616	0.3756	0.0655
specialist not infected-specialist infected	-1.1526	0.2684	1e-04

OBSERVATIONS

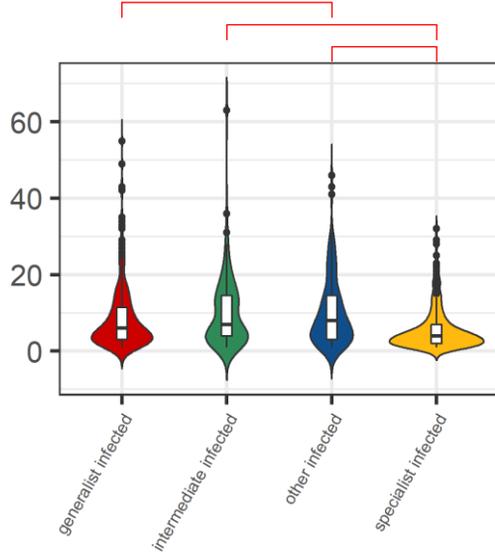
At 24 h p.i. none of the category is blocking the acidification of the phagolysosomes. However, despite an increase in the acidification of the phagolysosomes was observed, the number of macrophages displaying such acidification was found generally low (<25%).

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, stastically not significant

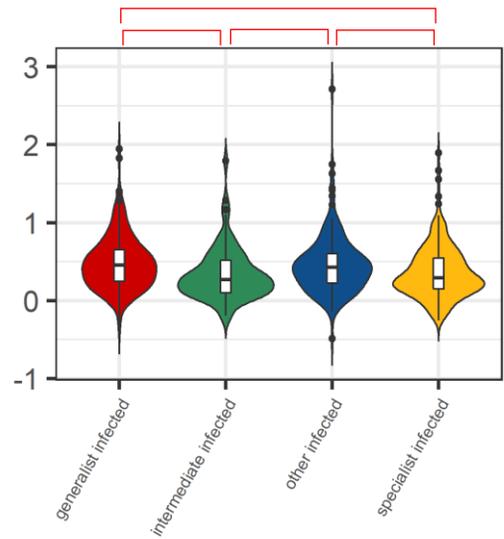
Grouping: Geo-host adaptation

M1 – 24 h.p.i.

c) Effective MOI



d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	1.6426	0.1203	0
groupintermediate infected	0.1433	0.1195	0.2348
groupother infected	0.4187	0.105	2e-04
groupspecialist infected	-0.1752	0.0937	0.0664

Contrast	estimate	SE	p.value
generalist infected - intermediate infected	-0.1433	0.1195	0.2348
generalist infected - other infected	-0.4187	0.105	5e-04
generalist infected - specialist infected	-0.1752	0.0937	0.0796
intermediate infected - other infected	-0.2753	0.1417	0.0796
intermediate infected - specialist infected	0.3185	0.1325	0.0385
other infected - specialist infected	0.5938	0.1065	0

Par	Value	Std.Error	p.value
(Intercept)	0.1697	0.0802	0.0347
groupintermediate infected	-0.535	0.1412	8e-04
groupother infected	-0.0769	0.1162	0.5107
groupspecialist infected	-0.3273	0.103	0.0023

Contrast	estimate	SE	p.value
generalist infected - intermediate infected	0.535	0.1412	0.0021
generalist infected - other infected	0.0769	0.1162	0.5107
generalist infected - specialist infected	0.3273	0.103	0.007
intermediate infected - other infected	-0.4581	0.1592	0.0111
intermediate infected - specialist infected	-0.2077	0.1496	0.2043
other infected - specialist infected	0.2504	0.1166	0.0537

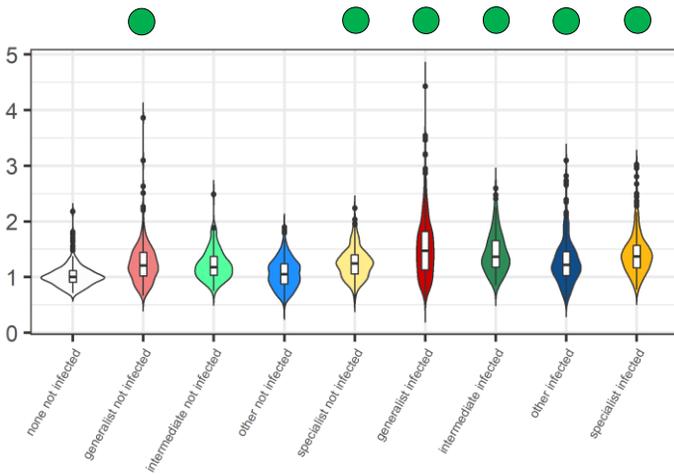
OBSERVATIONS

Specialist strains showed lower MOI values compared to the other groups. Colocalization with acidified compartments was found lower for specialist and intermediate strains.

Grouping: Geo-host adaptation

M2 – 6 h p.i.

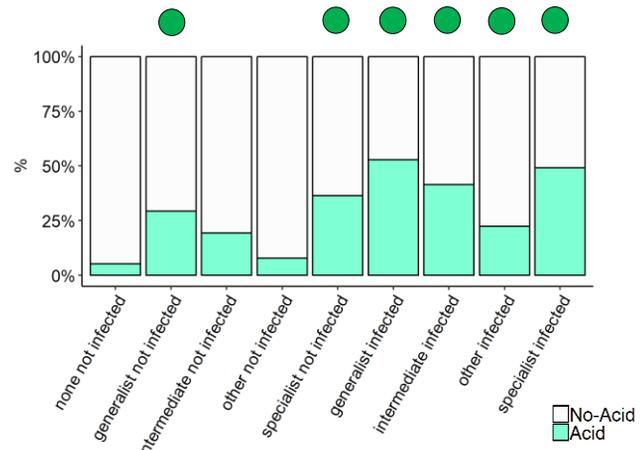
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	-0.0103	0.0505	0.8387
groupgeneralist not infected	0.1843	0.0441	0
groupintermediate not infected	0.1018	0.0612	0.0966
grouphother not infected	0.0694	0.0518	0.1809
groupspecialist not infected	0.1644	0.0458	3e-04
groupgeneralist infected	0.3254	0.044	0
groupintermediate infected	0.2311	0.0615	2e-04
grouphother infected	0.2086	0.052	1e-04
groupspecialist infected	0.2734	0.0459	0

Contrast	estimate	SE	p.value
none not infected-generalist not infected	-0.1843	0.0441	1e-04
none not infected-intermediate not infected	-0.1018	0.0612	0.1656
none not infected-other not infected	-0.0694	0.0518	0.2553
none not infected-specialist not infected	-0.1644	0.0458	3e-04
none not infected-generalist infected	-0.3254	0.044	0
none not infected-intermediate infected	-0.2311	0.0615	5e-04
none not infected-other infected	-0.2086	0.052	2e-04
none not infected-specialist infected	-0.2734	0.0459	0
generalist not infected-intermediate not infected	0.0825	0.0598	0.251
generalist not infected-other not infected	0.1149	0.0535	0.064
generalist not infected-specialist not infected	0.0199	0.0468	0.6993
intermediate not infected-other not infected	0.0324	0.0691	0.6973
intermediate not infected-specialist not infected	-0.0626	0.0634	0.3883
other not infected-specialist not infected	-0.095	0.0555	0.1403
generalist infected-intermediate infected	0.0943	0.06	0.1855
generalist infected-other infected	0.1168	0.0537	0.064
generalist infected-specialist infected	0.052	0.0468	0.337
intermediate infected-other infected	0.0225	0.0694	0.7457
intermediate infected-specialist infected	-0.0423	0.0637	0.5796
other infected-specialist infected	-0.0648	0.0537	0.3039
generalist not infected-generalist infected	-0.1411	0.0135	0
intermediate not infected-intermediate infected	-0.1294	0.0207	0
other not infected-other infected	-0.1393	0.0168	0
specialist not infected-specialist infected	-0.106	0.0139	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.2009	0.6363	0.0407	0
groupgeneralist not infected	1.6574	0.5541	5.2456	0.0028
groupintermediate not infected	0.8911	0.7029	2.4377	0.2049
grouphother not infected	1.2315	0.6648	3.4263	0.064
groupspecialist not infected	2.0146	0.5667	7.498	4e-04
groupgeneralist infected	3.1396	0.5477	23.0938	0
groupintermediate infected	2.4219	0.6849	11.2672	4e-04
grouphother infected	2.6032	0.6298	13.5067	0
groupspecialist infected	2.8956	0.5666	18.0946	0

Contrast	estimate	SE	p.value
none not infected-generalist not infected	-1.6574	0.5541	0.0067
none not infected-intermediate not infected	-0.8911	0.7029	0.3383
none not infected-other not infected	-1.2315	0.6648	0.1396
none not infected-specialist not infected	-2.0146	0.5667	0.0011
none not infected-generalist infected	-3.1396	0.5477	0
none not infected-intermediate infected	-2.4219	0.6849	0.0011
none not infected-other infected	-2.6032	0.6298	1e-04
none not infected-specialist infected	-2.8956	0.5666	0
generalist not infected-intermediate not infected	0.7663	0.6158	0.3383
generalist not infected-other not infected	0.4259	0.5959	0.5697
generalist not infected-specialist not infected	-0.3572	0.4812	0.5697
intermediate not infected-other not infected	-0.3404	0.748	0.6773
intermediate not infected-specialist not infected	-1.1236	0.6569	0.1744
other not infected-specialist not infected	-0.7832	0.5857	0.3345
generalist infected-intermediate infected	0.7177	0.5921	0.3383
generalist infected-other infected	0.5364	0.5454	0.4593
generalist infected-specialist infected	0.244	0.474	0.6619
intermediate infected-other infected	-0.1813	0.6974	0.7949
intermediate infected-specialist infected	-0.4737	0.6363	0.5697
other infected-specialist infected	-0.2924	0.5424	0.6619
generalist not infected-generalist infected	-1.4822	0.2261	0
intermediate not infected-intermediate infected	-1.5308	0.3463	1e-04
other not infected-other infected	-1.3717	0.326	1e-04
specialist not infected-specialist infected	-0.881	0.2259	3e-04

OBSERVATIONS

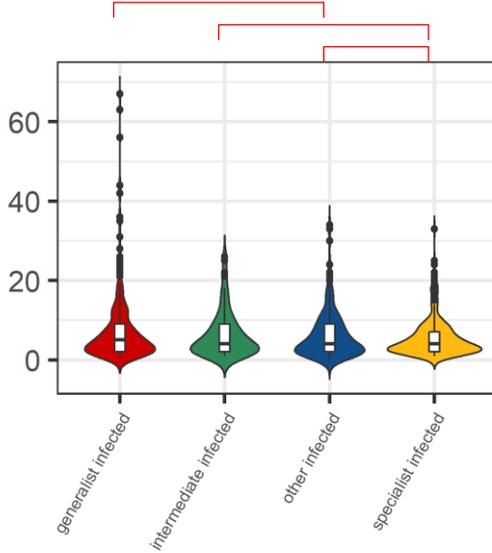
In M2 macrophages, none of the category is blocking the acidification of the phagolysosomes at 6 h p.i. and between 25% (other) and 50% (specialist and generalist) of macrophages showed acidification. A bystander effect in non-infected M2a macrophages was observed for specialist strains at 6 h p.i., with approx. 40% of bystander cells showing acidification.

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, stastically not significant

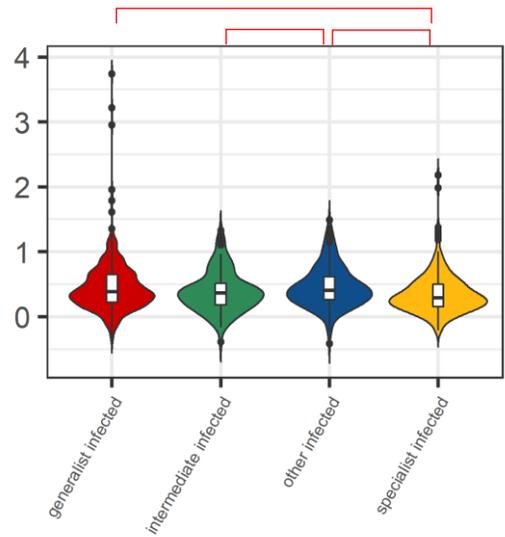
Grouping: Geo-host adaptation

M2 – 6 h p.i.

c) Effective MOI



d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	1.6426	0.1203	0
groupintermediate infected	0.1433	0.1195	0.2348
groupother infected	0.4187	0.105	2e-04
groupspecialist infected	-0.1752	0.0937	0.0664

Contrast	estimate	SE	p.value
generalist infected - intermediate infected	-0.1433	0.1195	0.2348
generalist infected - other infected	-0.4187	0.105	2e-04
generalist infected - specialist infected	0.1752	0.0937	0.0796
intermediate infected - other infected	-0.2753	0.1417	0.0796
intermediate infected - specialist infected	0.3185	0.1325	0.0385
other infected - specialist infected	0.5938	0.1065	0

Par	Value	Std.Error	p.value
(Intercept)	-0.0507	0.0532	0.3402
groupintermediate infected	-0.1478	0.0841	0.0839
groupother infected	0.1194	0.0758	0.1201
groupspecialist infected	-0.2237	0.0687	0.0018

Contrast	estimate	SE	p.value
generalist infected - intermediate infected	0.1478	0.0841	0.1259
generalist infected - other infected	-0.1194	0.0758	0.1441
generalist infected - specialist infected	0.2237	0.0687	0.0055
intermediate infected - other infected	-0.2672	0.0968	0.0153
intermediate infected - specialist infected	0.0759	0.0912	0.4084
other infected - specialist infected	0.3432	0.0769	2e-04

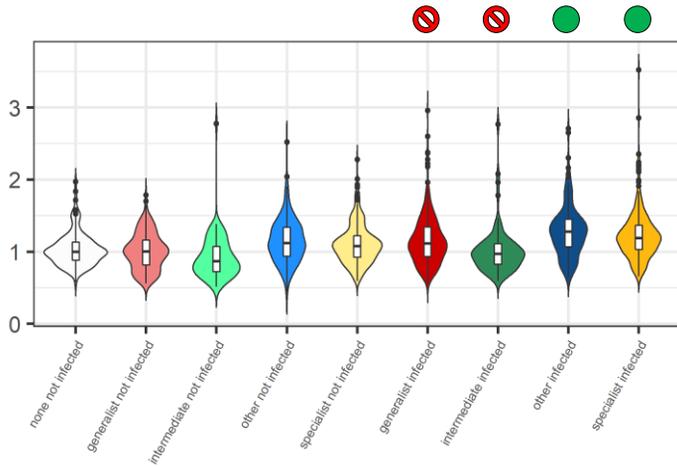
OBSERVATIONS

Specialist strains showed lower MOI values compared to the other groups. Colocalization with acidified compartments was found lower for specialist and intermediate strains.

Grouping: Geo-host adaptation

M2 – 24 h.p.i.

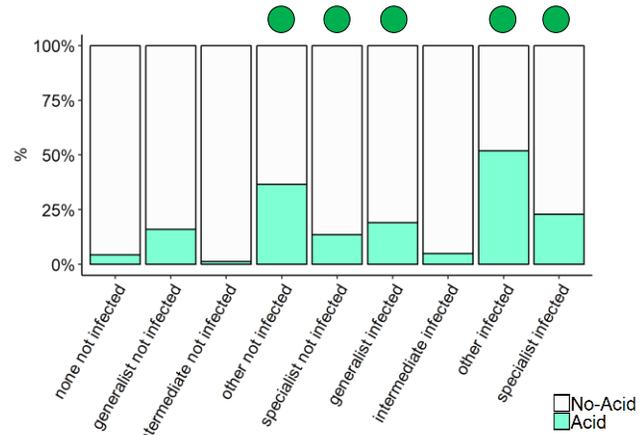
a) Phagolysosomal acidification



Par	Value	Std.Error	p-value
(Intercept)	0.0089	0.0545	0.87
groupgeneralist not infected	-0.0689	0.0316	0.0292
groupintermediate not infected	-0.0716	0.0421	0.0894
groupoth not infected	0.0026	0.0364	0.9437
groupspecialist not infected	0.0109	0.0315	0.7289
groupgeneralist infected	0.0347	0.031	0.2632
groupintermediate infected	-0.0083	0.0426	0.8447
groupoth infected	0.1243	0.0356	5e-04
groupspecialist infected	0.1061	0.0316	8e-04

Contrast	estimate	SE	p-value
none not infected-generalist not infected	0.0689	0.0316	0.0585
none not infected-intermediate not infected	0.0716	0.0421	0.1431
none not infected-other not infected	-0.0026	0.0364	0.9471
none not infected-specialist not infected	-0.0109	0.0315	0.8747
none not infected-generalist infected	-0.0347	0.031	0.3716
none not infected-intermediate infected	0.0083	0.0426	0.9215
none not infected-other infected	-0.1243	0.0356	0.003
none not infected-specialist infected	-0.1061	0.0316	0.0038
generalist not infected-intermediate not infected	0.0027	0.0403	0.9471
generalist not infected-other not infected	-0.0715	0.0362	0.0882
generalist not infected-specialist not infected	-0.0799	0.0314	0.0294
intermediate not infected-other not infected	-0.0742	0.0465	0.1664
intermediate not infected-specialist not infected	-0.0825	0.0423	0.0882
other not infected-specialist not infected	-0.0084	0.0356	0.9215
generalist infected-intermediate infected	0.0431	0.0404	0.3812
generalist infected-other infected	-0.0895	0.0351	0.0294
generalist infected-specialist infected	-0.0714	0.0309	0.0507
intermediate infected-other infected	-0.1326	0.0463	0.017
intermediate infected-specialist infected	-0.1145	0.0429	0.0264
other infected-specialist infected	0.0182	0.035	0.7625
generalist not infected-generalist infected	-0.1037	0.0171	0
intermediate not infected-intermediate infected	-0.0633	0.0281	0.0529
other not infected-other infected	-0.1217	0.0195	0
specialist not infected-specialist infected	-0.0952	0.0158	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.574	0.6275	0.028	0
groupgeneralist not infected	1.1131	0.5049	3.0436	0.0275
groupintermediate not infected	-0.0353	1.1207	0.9653	0.9749
groupoth not infected	1.5681	0.4806	4.7973	0.0011
groupspecialist not infected	1.1996	0.4875	3.3188	0.0139
groupgeneralist infected	1.6838	0.4892	5.3859	6e-04
groupintermediate infected	1.4906	0.7107	4.4397	0.036
groupoth infected	2.5577	0.4779	12.906	0
groupspecialist infected	1.9581	0.4754	7.0859	0

Contrast	estimate	SE	p-value
none not infected-generalist not infected	-1.1131	0.5049	0.066
none not infected-intermediate not infected	0.0353	1.1207	0.9749
none not infected-other not infected	-1.5681	0.4806	0.0053
none not infected-specialist not infected	-1.1996	0.4875	0.0416
none not infected-generalist infected	-1.6838	0.4892	0.0035
none not infected-intermediate infected	-1.4906	0.7107	0.0784
none not infected-other infected	-2.5577	0.4779	0
none not infected-specialist infected	-1.9581	0.4754	5e-04
generalist not infected-intermediate not infected	1.1483	1.0546	0.3489
generalist not infected-other not infected	-0.455	0.3167	0.2412
generalist not infected-specialist not infected	-0.0865	0.3425	0.8353
intermediate not infected-other not infected	-1.6033	1.0551	0.2205
intermediate not infected-specialist not infected	-1.2349	1.0548	0.3223
other not infected-specialist not infected	0.3685	0.2927	0.2939
generalist infected-intermediate infected	0.1932	0.5786	0.8056
generalist infected-other infected	-0.8739	0.2967	0.011
generalist infected-specialist infected	-0.2743	0.2897	0.4125
intermediate infected-other infected	-1.0671	0.5957	0.1352
intermediate infected-specialist infected	-0.4675	0.5895	0.4889
other infected-specialist infected	0.5996	0.2669	0.0659
generalist not infected-generalist infected	-0.5707	0.2952	0.1064
intermediate not infected-intermediate infected	-1.5259	1.1313	0.2661
other not infected-other infected	-0.9896	0.2527	7e-04
specialist not infected-specialist infected	-0.7585	0.2485	0.0091

OBSERVATIONS

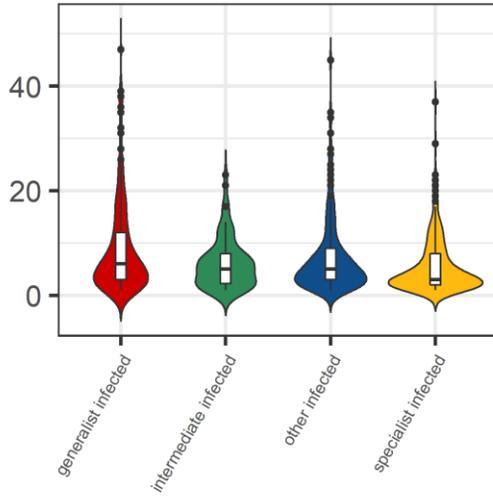
As the infection progresses, generalist and intermediate categories are blocking the acidification of the phagolysosomes. The number of acidified macrophages also decreased below 25% except for the category other (50%). A bystander effect in non-infected M2a macrophages was observed for specialist strains at 24 h p.i, despite it was not found statistically significant.

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

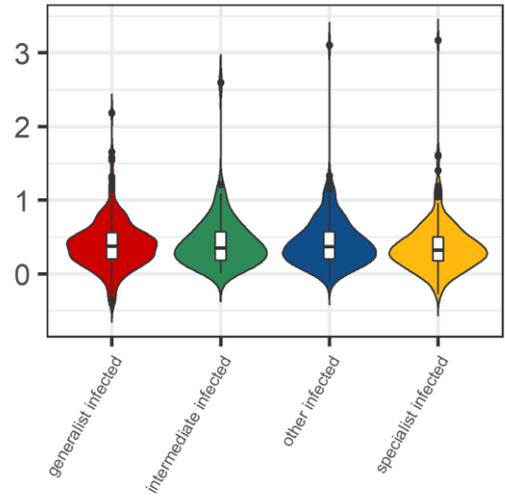
Grouping: Geo-host adaptation

M2 – 24 h p.i.

c) Effective MOI



d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	1.6645	0.1221	0
groupintermediate infected	-0.2831	0.156	0.0745
groupother infected	0.0267	0.1361	0.8448
groupspecialist infected	-0.2481	0.1199	0.0427

Contrast	estimate	SE	p.value
generalist infected - intermediate infected	0.2831	0.156	0.1367
generalist infected - other infected	-0.0267	0.1361	0.8448
generalist infected - specialist infected	0.2481	0.1199	0.1362
intermediate infected - other infected	-0.3098	0.1805	0.1367
intermediate infected - specialist infected	-0.035	0.1669	0.8448
other infected - specialist infected	0.2748	0.1345	0.1362

Par	Value	Std.Error	p.value
(Intercept)	-0.0522	0.0609	0.3923
groupintermediate infected	0.0401	0.1112	0.7194
groupother infected	0.0632	0.0879	0.4751
groupspecialist infected	-0.1392	0.0807	0.0897

Contrast	estimate	SE	p.value
generalist infected - intermediate infected	-0.0401	0.1112	0.8459
generalist infected - other infected	-0.0632	0.0879	0.7127
generalist infected - specialist infected	0.1392	0.0807	0.2326
intermediate infected - other infected	-0.023	0.118	0.8459
intermediate infected - specialist infected	0.1793	0.1125	0.2326
other infected - specialist infected	0.2023	0.0858	0.1299

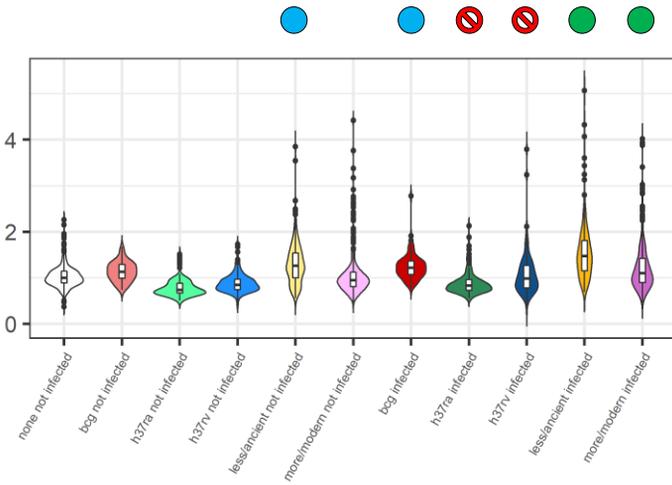
OBSERVATIONS

No statistically relevant differences in effective MOI at 24 h p.i., even though generalist strains displayed an higher distribution of MOI values. Colocalization with acidified compartments was similar among all the groups.

Grouping: Basic

M1 – 6 h p.i.

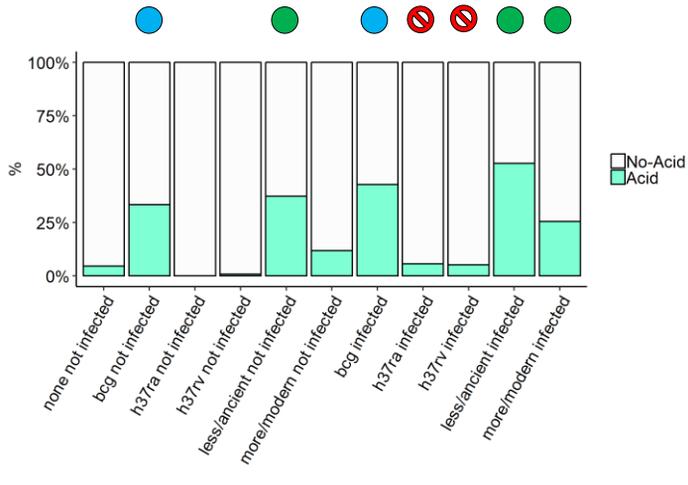
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	0.0057	0.0692	0.9345
groupbcg not infected	0.0151	0.1015	0.882
group h37ra not infected	-0.1856	0.0793	0.0193
group h37rv not infected	-0.0647	0.0798	0.4178
group less/ancient not infected	0.1341	0.058	0.0208
group more/modern not infected	0.0508	0.057	0.3733
groupbcg infected	0.0803	0.1013	0.4281
group h37ra infected	-0.1042	0.0796	0.1907
group h37rv infected	0.0489	0.0803	0.5429
group less/ancient infected	0.2842	0.0581	0
group more/modern infected	0.1729	0.057	0.0025

Contrast	estimate	SE	p.value
none not infected-bcg not infected	-0.0151	0.1015	0.882
none not infected-h37ra not infected	0.1856	0.0793	0.052
none not infected-h37rv not infected	0.0647	0.0798	0.4995
none not infected-less/ancient not infected	-0.1341	0.058	0.052
none not infected-bcg infected	-0.0508	0.057	0.4791
none not infected-h37ra infected	-0.0803	0.1013	0.4995
none not infected-h37rv infected	0.1042	0.0796	0.267
none not infected-h37rv infected	-0.0489	0.0803	0.5939
none not infected-less/ancient infected	-0.2842	0.0581	0
none not infected-more/modern infected	-0.1729	0.057	0.0095
bcg not infected-h37ra not infected	0.2006	0.1223	0.186
bcg not infected-h37rv not infected	0.0798	0.1234	0.5848
bcg not infected-less/ancient not infected	-0.1191	0.0966	0.2933
bcg not infected-more/modern not infected	-0.0357	0.1063	0.7816
h37ra not infected-h37rv not infected	-0.1209	0.0895	0.258
h37ra not infected-less/ancient not infected	-0.3197	0.087	0.0014
h37ra not infected-more/modern not infected	-0.2363	0.0834	0.0163
h37rv not infected-less/ancient not infected	-0.1988	0.0892	0.0605
h37rv not infected-more/modern not infected	-0.1155	0.0836	0.2549
less/ancient not infected-more/modern not infected	0.0833	0.0589	0.2504
bcg infected-h37ra infected	0.1845	0.1224	0.2306
bcg infected-h37rv infected	0.0314	0.1236	0.8228
bcg infected-less/ancient infected	-0.2039	0.0966	0.0762
bcg infected-more/modern infected	-0.0926	0.1062	0.4791
h37ra infected-h37rv infected	-0.1531	0.0903	0.1753
h37ra infected-less/ancient infected	-0.3885	0.0874	1e-04
h37ra infected-more/modern infected	-0.2771	0.0837	0.0041
h37rv infected-less/ancient infected	-0.2354	0.0897	0.0279
h37rv infected-more/modern infected	-0.124	0.084	0.2335
less/ancient infected-more/modern infected	0.1113	0.0591	0.1227
bcg not infected-bcg infected	-0.0653	0.0265	0.0407
h37ra not infected-h37ra infected	-0.0814	0.0237	0.003
h37rv not infected-h37rv infected	-0.1136	0.0231	0
less/ancient not infected-less/ancient infected	-0.1501	0.0143	0
more/modern not infected-more/modern infected	-0.1222	0.0138	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.5104	0.7705	0.0299	0
groupbcg not infected	0.8759	0.942	2.401	0.3525
group h37ra not infected	-14.8252	582.0863	0	0.9797
group h37rv not infected	-1.2669	1.3848	0.2817	0.3603
group less/ancient not infected	1.7648	0.6365	5.8405	0.0056
group more/modern not infected	0.8433	0.6914	2.324	0.2226
groupbcg infected	1.2845	0.9366	3.6128	0.1702
group h37ra infected	0.8777	0.9127	2.4054	0.3362
group h37rv infected	0.7321	1.0357	2.0794	0.4797
group less/ancient infected	2.9099	0.6374	18.3541	0
group more/modern infected	2.4233	0.6719	11.2825	3e-04

Contrast	estimate	SE	p.value
none not infected-bcg not infected	-0.8759	0.942	0.5483
none not infected-h37ra not infected	14.8252	582.0863	0.9814
none not infected-h37rv not infected	1.2669	1.3848	0.5483
none not infected-less/ancient not infected	-1.7648	0.6365	0.0389
none not infected-more/modern not infected	-0.8433	0.6914	0.4411
none not infected-bcg infected	-1.2845	0.9366	0.3911
none not infected-h37ra infected	-0.8777	0.9127	0.5483
none not infected-h37rv infected	-0.7321	1.0357	0.6715
none not infected-less/ancient infected	-2.9099	0.6374	1e-04
none not infected-more/modern infected	-2.4233	0.6719	0.0027
bcg not infected-h37ra not infected	15.701	582.0868	0.9814
bcg not infected-h37rv not infected	2.1427	1.5937	0.3911
bcg not infected-less/ancient not infected	-0.889	0.8381	0.5055
bcg not infected-more/modern not infected	-0.0326	0.9672	0.9814
h37ra not infected-h37rv not infected	-13.5583	582.0872	0.9814
h37ra not infected-less/ancient not infected	-16.59	582.0863	0.9814
h37ra not infected-more/modern not infected	-15.6685	582.0863	0.9814
h37rv not infected-less/ancient not infected	-3.0317	1.4184	0.1628
h37rv not infected-more/modern not infected	-2.1101	1.422	0.3445
less/ancient not infected-more/modern not infected	0.9215	0.5751	0.3053
bcg infected-h37ra infected	0.4068	1.183	0.9475
bcg infected-h37rv infected	0.5524	1.2976	0.9023
bcg infected-less/ancient infected	-1.6254	0.839	0.205
bcg infected-more/modern infected	-1.1388	0.949	0.4411
h37ra infected-h37rv infected	1.456	1.0474	0.9814
h37ra infected-less/ancient infected	-2.0321	0.9258	0.1628
h37ra infected-more/modern infected	-1.5455	0.9097	0.2843
h37rv infected-less/ancient infected	-2.1778	1.0791	0.1906
h37rv infected-more/modern infected	-1.6912	1.0683	0.3053
less/ancient infected-more/modern infected	0.4866	0.5523	0.5516
bcg not infected-bcg infected	-0.4086	0.3474	0.4411
h37ra not infected-h37ra infected	-15.7029	582.0859	0.9814
h37rv not infected-h37rv infected	-1.9989	1.1041	0.2458
less/ancient not infected-less/ancient infected	-1.145	0.2251	0
more/modern not infected-more/modern infected	-1.58	0.3054	0

OBSERVATIONS

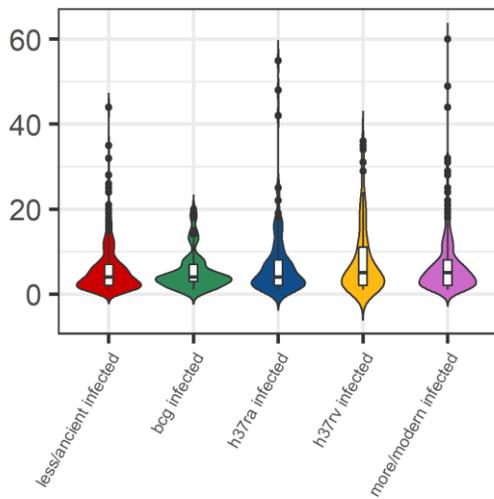
BCG, H37Rv, and H37Ra were found to block acidification of the phagolysosomes in M1 macrophages at 6 h p.i. Less/ancient strains also induced acidification in a higher percentage (50%) of cells compared to the other groups.

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Basic

M1 – 6 h p.i.

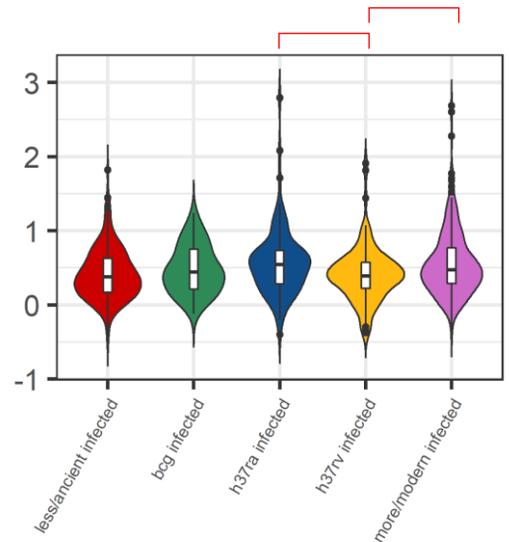
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.3998	0.1168	0
groupbcg infected	-0.0196	0.2119	0.9268
grouph37ra infected	0.1092	0.1902	0.5682
grouph37rv infected	0.1091	0.1949	0.5779
groupmore/modern infected	0.0388	0.1302	0.767

Contrast	estimate	SE	p.value
less/ancient infected - bcg infected	-0.0196	0.2119	0.9996
less/ancient infected - h37ra infected	-0.1092	0.1902	0.9996
less/ancient infected - h37rv infected	-0.1091	0.1949	0.9996
less/ancient infected - more/modern infected	-0.0388	0.1302	0.9996
bcg infected - h37ra infected	-0.1287	0.2669	0.9996
bcg infected - h37rv infected	-0.1286	0.2689	0.9996
bcg infected - more/modern infected	-0.0583	0.2326	0.9996
h37ra infected - h37rv infected	1e-04	0.2007	0.9996
h37ra infected - more/modern infected	0.0704	0.1824	0.9996
h37rv infected - more/modern infected	0.0703	0.1825	0.9996

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	-0.0522	0.0991	0.5983
groupbcg infected	0.1116	0.1679	0.5087
grouph37ra infected	0.2712	0.1543	0.0839
grouph37rv infected	-0.1589	0.1582	0.3193
groupmore/modern infected	0.2451	0.1047	0.0226

Contrast	estimate	SE	p.value
less/ancient infected - bcg infected	-0.1116	0.1679	0.5653
less/ancient infected - h37ra infected	-0.2712	0.1543	0.2097
less/ancient infected - h37rv infected	0.1589	0.1582	0.5321
less/ancient infected - more/modern infected	-0.2451	0.1047	0.0753
bcg infected - h37ra infected	-0.1596	0.2157	0.5653
bcg infected - h37rv infected	0.2705	0.2173	0.4363
bcg infected - more/modern infected	-0.1335	0.1868	0.5653
h37ra infected - h37rv infected	0.4301	0.16	0.0464
h37ra infected - more/modern infected	0.0261	0.1468	0.8594
h37rv infected - more/modern infected	-0.4039	0.1456	0.0464

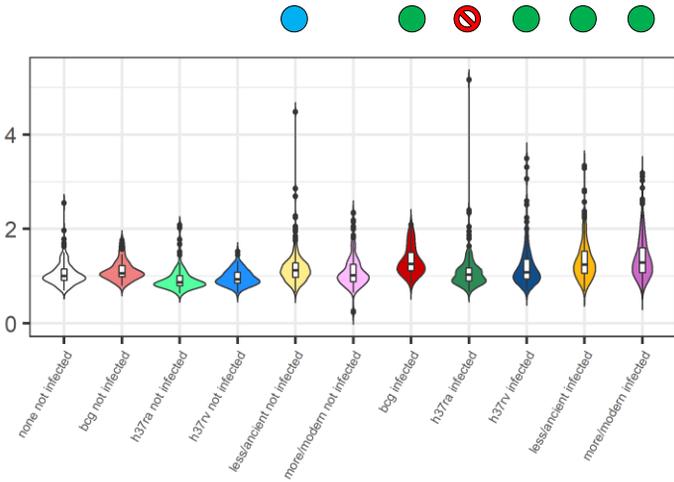
OBSERVATIONS

H37Rv showed lower colocalization with acidified compartments. No statistically relevant differences in effective MOI were observed among the categories in M1 macrophages, irrespectively from the time point considered.

Grouping: Basic

M1 – 24 h.p.i.

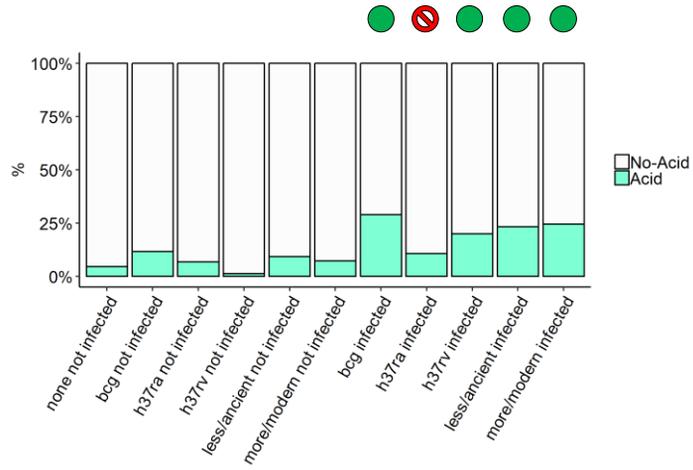
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	0.0174	0.0363	0.6328
groupbcg not infected	0.0522	0.0553	0.3454
grouph37ra not infected	-0.0886	0.0465	0.0567
grouph37rv not infected	-0.0205	0.0461	0.6568
groupless/ancient not infected	0.044	0.0325	0.1757
groupmore/modern not infected	0.0294	0.033	0.3727
groupbcg infected	0.2095	0.0529	1e-04
grouph37ra infected	0.0464	0.0447	0.2994
grouph37rv infected	0.1162	0.0449	0.0097
groupless/ancient infected	0.1544	0.0326	0
groupmore/modern infected	0.2248	0.0323	0

Contrast	estimate	SE	p.value
none not infected-bcg not infected	-0.0522	0.0553	0.4318
none not infected-h37ra not infected	0.0886	0.0465	0.1103
none not infected-h37rv not infected	0.0205	0.0461	0.7183
none not infected-less/ancient not infected	-0.044	0.0325	0.2863
none not infected-more/modern not infected	-0.0294	0.033	0.4499
none not infected-bcg infected	-0.2095	0.0529	3e-04
none not infected-h37ra infected	-0.0464	0.0447	0.3881
none not infected-h37rv infected	-0.1162	0.0449	0.0308
none not infected-less/ancient infected	-0.1544	0.0326	0
none not infected-more/modern infected	-0.2248	0.0323	0
bcg not infected-h37ra not infected	0.1408	0.0672	0.0747
bcg not infected-h37rv not infected	0.0727	0.0674	0.3876
bcg not infected-less/ancient not infected	0.0082	0.0522	0.8752
bcg not infected-more/modern not infected	0.0228	0.0575	0.7329
h37ra not infected-h37rv not infected	-0.0681	0.0508	0.2863
h37ra not infected-less/ancient not infected	-0.1326	0.0486	0.0225
h37ra not infected-more/modern not infected	-0.118	0.0471	0.0335
h37rv not infected-less/ancient not infected	-0.0645	0.0492	0.2889
h37rv not infected-more/modern not infected	-0.0499	0.0469	0.3876
less/ancient not infected-more/modern not infected	0.0146	0.0323	0.7183
bcg infected-h37ra infected	0.1631	0.064	0.032
bcg infected-h37rv infected	0.0933	0.0646	0.2607
bcg infected-less/ancient infected	0.0551	0.0498	0.3876
bcg infected-more/modern infected	-0.0152	0.0548	0.8038
h37ra infected-h37rv infected	-0.0699	0.0478	0.2607
h37ra infected-less/ancient infected	-0.108	0.047	0.0506
h37ra infected-more/modern infected	-0.1784	0.045	3e-04
h37rv infected-less/ancient infected	-0.0382	0.0481	0.4983
h37rv infected-more/modern infected	-0.1085	0.045	0.0399
less/ancient infected-more/modern infected	-0.0703	0.0318	0.059
bcg not infected-bcg infected	-0.1573	0.0359	1e-04
h37ra not infected-h37ra infected	-0.135	0.0309	1e-04
h37rv not infected-h37rv infected	-0.1367	0.03	0
less/ancient not infected-less/ancient infected	-0.1104	0.018	0
more/modern not infected-more/modern infected	-0.1954	0.0197	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.1636	0.4495	0.0423	0
groupbcg not infected	0.844	0.6733	2.3256	0.21
grouph37ra not infected	0.9602	0.6738	2.6121	0.1542
grouph37rv not infected	-1.0824	1.1144	0.3388	0.3314
groupless/ancient not infected	0.5498	0.4805	1.7329	0.2526
groupmore/modern not infected	0.528	0.5087	1.6956	0.2993
groupbcg infected	1.9837	0.5712	7.2697	5e-04
grouph37ra infected	1.4416	0.5806	4.2274	0.013
grouph37rv infected	1.8728	0.5358	6.5968	5e-04
groupless/ancient infected	1.6986	0.454	5.4662	2e-04
groupmore/modern infected	1.9572	0.4554	7.0796	0

Contrast	estimate	SE	p-value
none not infected-bcg not infected	-0.844	0.6733	0.4901
none not infected-h37ra not infected	-0.9602	0.6738	0.3855
none not infected-h37rv not infected	1.0824	1.1144	0.5927
none not infected-less/ancient not infected	-0.5498	0.4805	0.5525
none not infected-more/modern not infected	-0.528	0.5087	0.5819
none not infected-bcg infected	-1.9837	0.5712	0.003
none not infected-h37ra infected	-1.4416	0.5806	0.057
none not infected-h37rv infected	-1.8728	0.5358	0.003
none not infected-less/ancient infected	-1.6986	0.454	0.0016
none not infected-more/modern infected	-1.9572	0.4554	2e-04
bcg not infected-h37ra not infected	-0.1162	0.7856	0.9359
bcg not infected-h37rv not infected	1.9263	1.1877	0.3336
bcg not infected-less/ancient not infected	0.2942	0.5809	0.7503
bcg not infected-more/modern not infected	0.316	0.6403	0.7503
h37ra not infected-h37rv not infected	2.0425	1.1462	0.2617
h37ra not infected-less/ancient not infected	0.4104	0.6159	0.6846
h37ra not infected-more/modern not infected	0.4321	0.6217	0.6846
h37rv not infected-less/ancient not infected	-1.6322	1.083	0.3679
h37rv not infected-more/modern not infected	-1.6104	1.082	0.3679
less/ancient not infected-more/modern not infected	0.0218	0.4011	0.9567
bcg infected-h37ra infected	0.5421	0.6118	0.5927
bcg infected-h37rv infected	1.1109	0.5731	0.926
bcg infected-less/ancient infected	0.2851	0.4313	0.6846
bcg infected-more/modern infected	0.0265	0.4825	0.9567
h37ra infected-h37rv infected	-0.4313	0.4824	0.5927
h37ra infected-less/ancient infected	-0.257	0.4874	0.7503
h37ra infected-more/modern infected	-0.5156	0.466	0.5528
h37rv infected-less/ancient infected	0.1743	0.4381	0.8059
h37rv infected-more/modern infected	-0.0844	0.3987	0.926
less/ancient infected-more/modern infected	-0.2586	0.2933	0.5927
bcg not infected-bcg infected	-1.1397	0.5064	0.0949
h37ra not infected-h37ra infected	-0.4814	0.5595	0.5927
h37rv not infected-h37rv infected	-2.9552	1.0384	0.0221
less/ancient not infected-less/ancient infected	-1.1488	0.2682	2e-04
more/modern not infected-more/modern infected	-1.4292	0.3142	2e-04

OBSERVATIONS

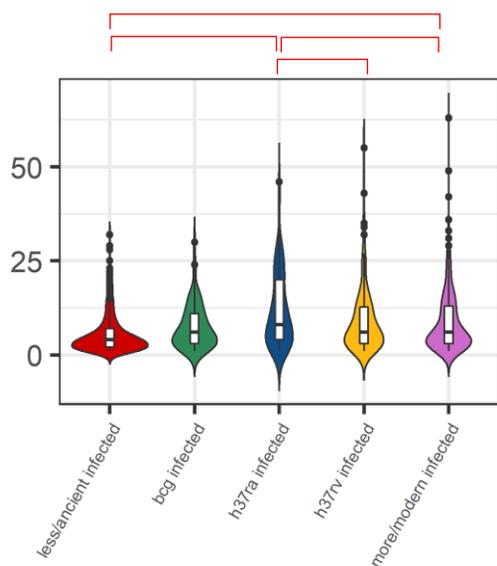
At 24 h.p.i., only H37Ra was retaining the blocking of the acidification of the phagolysosomes. However, despite an increase in the acidification of the phagolysosomes was observed, the number of macrophages displaying such acidification was found generally low (25%).

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Basic

M1 – 24 h.p.i.

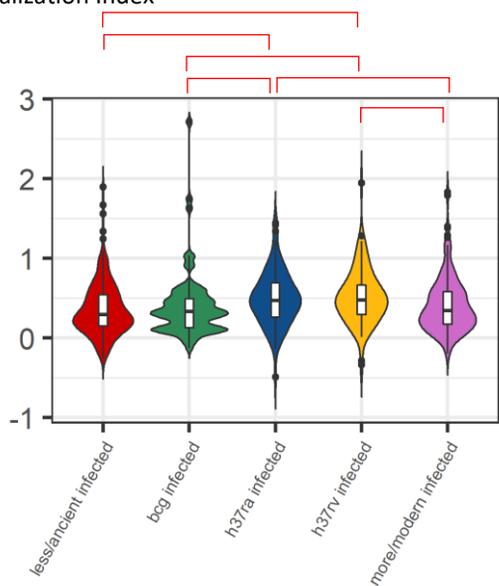
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.4775	0.134	0
groupbcg infected	0.4793	0.1543	0.0029
grouph37ra infected	0.6163	0.1567	2e-04
grouph37rv infected	0.0909	0.1604	0.5728
groupmore/modern infected	0.2335	0.0988	0.0214

Contrast	estimate	SE	p.value
less/ancient infected - bcg infected	-0.4793	0.1543	0.0096
less/ancient infected - h37ra infected	-0.6163	0.1567	0.0014
less/ancient infected - h37rv infected	-0.0909	0.1604	0.5728
less/ancient infected - more/modern infected	-0.2335	0.0988	0.0428
bcg infected - h37ra infected	-0.137	0.2153	0.5728
bcg infected - h37rv infected	0.3884	0.2177	0.1325
bcg infected - more/modern infected	0.2458	0.1797	0.2521
h37ra infected - h37rv infected	0.5254	0.1358	0.0014
h37ra infected - more/modern infected	0.3828	0.143	0.0239
h37rv infected - more/modern infected	-0.1426	0.142	0.3992

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	-0.2816	0.1095	0.0104
groupbcg infected	0.0636	0.16	0.6924
grouph37ra infected	0.7087	0.1571	0
grouph37rv infected	0.7889	0.1571	0
groupmore/modern infected	0.135	0.1031	0.1956

Contrast	estimate	SE	p.value
less/ancient infected - bcg infected	-0.0636	0.16	0.6976
less/ancient infected - h37ra infected	-0.7087	0.1571	1e-04
less/ancient infected - h37rv infected	-0.7889	0.1571	1e-04
less/ancient infected - more/modern infected	-0.135	0.1031	0.2794
bcg infected - h37ra infected	-0.6451	0.215	0.0066
bcg infected - h37rv infected	-0.7253	0.2145	0.0026
bcg infected - more/modern infected	-0.0714	0.1829	0.6976
h37ra infected - h37rv infected	-0.0802	0.1438	0.6976
h37ra infected - more/modern infected	0.5737	0.1479	7e-04
h37rv infected - more/modern infected	0.6538	0.1423	1e-04

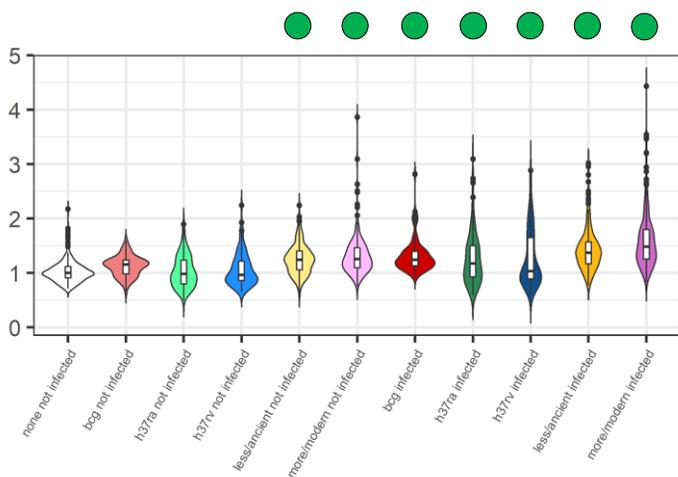
OBSERVATIONS

H37Rv and H37Ra showed higher colocalization with acidified compartments. No statistically relevant differences in effective MOI were observed among the categories in M1 macrophages, irrespectively from the time point considered.

Grouping: Basic

M2 – 6 h p.i.

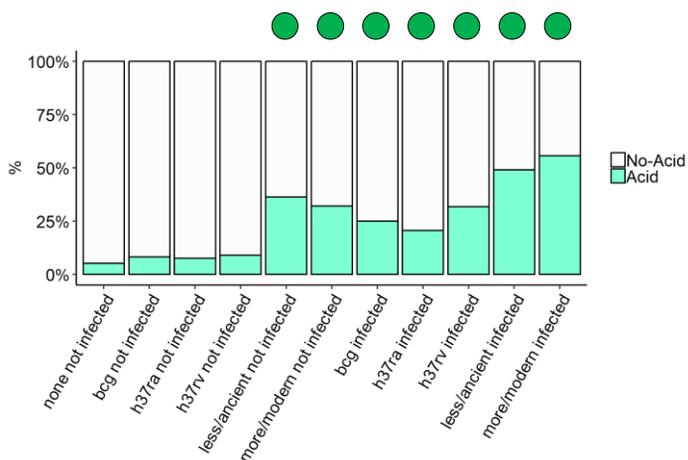
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	-0.0077	0.042	0.8545
groupbcg not infected	0.123	0.0795	0.1222
grouph37ra not infected	0.0073	0.0617	0.9055
grouph37rv not infected	0.0416	0.0623	0.5042
groupless/ancient not infected	0.1856	0.0462	1e-04
groupmore/modern not infected	0.2127	0.0456	0
groupbcg infected	0.2296	0.0794	0.0104
grouph37ra infected	0.1687	0.0621	0.0167
grouph37rv infected	0.1675	0.0624	0.0167
groupless/ancient infected	0.2946	0.0464	0
groupmore/modern infected	0.3585	0.0455	0

Contrast	estimate	SE	p.value
none not infected-bcg not infected	-0.123	0.0795	0.1944
none not infected-h37ra not infected	-0.0073	0.0617	0.9321
none not infected-h37rv not infected	-0.0416	0.0623	0.5847
none not infected-less/ancient not infected	-0.1856	0.0462	3e-04
none not infected-more/modern not infected	-0.2127	0.0456	0
none not infected-bcg infected	-0.2296	0.0794	0.0104
none not infected-h37ra infected	-0.1687	0.0621	0.0167
none not infected-h37rv infected	-0.1675	0.0624	0.0167
none not infected-less/ancient infected	-0.2946	0.0464	0
none not infected-more/modern infected	-0.3585	0.0455	0
bcg not infected-h37ra not infected	0.1157	0.0941	0.3196
bcg not infected-h37rv not infected	0.0814	0.0949	0.5102
bcg not infected-less/ancient not infected	-0.0626	0.0763	0.5148
bcg not infected-more/modern not infected	-0.0897	0.0827	0.3894
h37ra not infected-h37rv not infected	-0.0343	0.0718	0.6713
h37ra not infected-less/ancient not infected	-0.1783	0.0667	0.0167
h37ra not infected-more/modern not infected	-0.2053	0.0646	0.0053
h37rv not infected-less/ancient not infected	-0.144	0.0685	0.0692
h37rv not infected-more/modern not infected	-0.1711	0.0651	0.0178
less/ancient not infected-more/modern not infected	-0.0271	0.0467	0.6145
bcg infected-h37ra infected	0.0609	0.0942	0.5847
bcg infected-h37rv infected	0.0621	0.0948	0.5847
bcg infected-less/ancient infected	-0.065	0.0762	0.5102
bcg infected-more/modern infected	-0.1289	0.0825	0.1944
h37ra infected-h37rv infected	0.0012	0.0723	0.9866
h37ra infected-less/ancient infected	-0.1259	0.0672	0.1125
h37ra infected-more/modern infected	-0.1898	0.0649	0.0102
h37rv infected-less/ancient infected	-0.1271	0.0687	0.1125
h37rv infected-more/modern infected	-0.191	0.0651	0.0102
less/ancient infected-more/modern infected	-0.0639	0.0467	0.2611
bcg not infected-bcg infected	-0.1066	0.0266	3e-04
h37ra not infected-h37ra infected	-0.1614	0.0219	0
h37rv not infected-h37rv infected	-0.1258	0.0223	0
less/ancient not infected-less/ancient infected	-0.109	0.014	0
more/modern not infected-more/modern infected	-0.1458	0.0132	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.172	0.602	0.0419	0
groupbcg not infected	0.755	0.9531	2.1276	0.4283
grouph37ra not infected	1.2814	0.7846	3.6017	0.1024
grouph37rv not infected	-0.0263	0.821	0.974	0.9744
groupless/ancient not infected	2.078	0.5798	7.9888	3e-04
groupmore/modern not infected	1.8699	0.5617	6.4876	0e-04
groupbcg infected	2.1862	0.8833	8.9015	0.0133
grouph37ra infected	2.6218	0.7381	13.7602	4e-04
grouph37rv infected	2.3266	0.7461	10.2435	0.0018
groupless/ancient infected	2.9587	0.5796	19.2736	0
groupmore/modern infected	3.2052	0.5582	24.6607	0

Contrast	estimate	SE	p.value
none not infected-bcg not infected	-0.755	0.9531	0.5774
none not infected-h37ra not infected	-1.2814	0.7846	0.239
none not infected-h37rv not infected	-0.0263	0.821	0.9744
none not infected-less/ancient not infected	-2.078	0.5798	0.0019
none not infected-more/modern not infected	-1.8699	0.5617	0.0038
none not infected-bcg infected	-2.1862	0.8833	0.0359
none not infected-h37ra infected	-2.6218	0.7381	0.0019
none not infected-h37rv infected	-2.3266	0.7461	0.0064
none not infected-less/ancient infected	-2.9587	0.5796	0
none not infected-more/modern infected	-3.2052	0.5582	0
bcg not infected-h37ra not infected	-0.5264	1.0983	0.7311
bcg not infected-h37rv not infected	0.7813	1.1336	0.6361
bcg not infected-less/ancient not infected	-1.323	0.8506	0.2622
bcg not infected-more/modern not infected	-1.1149	0.929	0.4084
h37ra not infected-h37rv not infected	1.3077	0.8827	0.2851
h37ra not infected-less/ancient not infected	-0.7966	0.7783	0.5023
h37ra not infected-more/modern not infected	-0.5885	0.744	0.5774
h37rv not infected-less/ancient not infected	-2.1044	0.8302	0.0328
h37rv not infected-more/modern not infected	-1.8962	0.7762	0.0364
less/ancient not infected-more/modern not infected	0.2082	0.4859	0.7311
bcg infected-h37ra infected	-0.4356	1.0034	0.7311
bcg infected-h37rv infected	-0.1404	1.0198	0.9167
bcg infected-less/ancient infected	-0.7725	0.77	0.5023
bcg infected-more/modern infected	-1.019	0.8551	0.4084
h37ra infected-h37rv infected	0.2951	0.7547	0.7379
h37ra infected-less/ancient infected	-0.337	0.7293	0.7311
h37ra infected-more/modern infected	-0.5834	0.6888	0.5774
h37rv infected-less/ancient infected	-0.6321	0.7565	0.5774
h37rv infected-more/modern infected	-0.8786	0.699	0.406
less/ancient infected-more/modern infected	-0.2465	0.4817	0.7311
bcg not infected-bcg infected	1.4312	0.5083	0.0155
h37ra not infected-h37ra infected	-1.3404	0.4263	0.0064
h37rv not infected-h37rv infected	-2.353	0.5066	0
less/ancient not infected-less/ancient infected	-0.8807	0.2259	7e-04
more/modern not infected-more/modern infected	-1.3353	0.2061	0

OBSERVATIONS

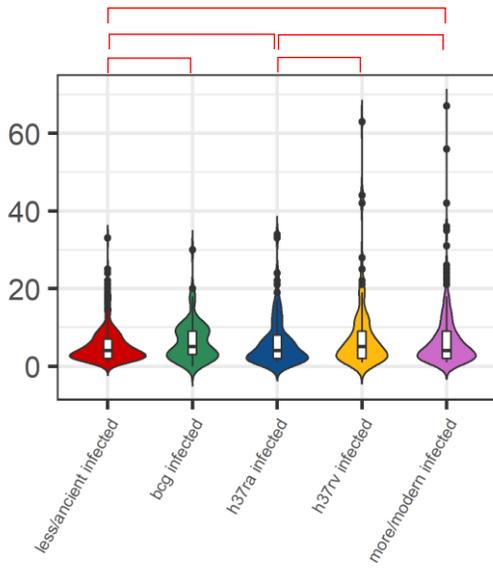
In M2 macrophages, none of the group displayed the blockage of the acidification of the phagolysosomes and between 25% (H37Rv, H37Ra) and 50% (more virulent/modern) of macrophages showed acidification.

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Basic

M2 – 6 h p.i.

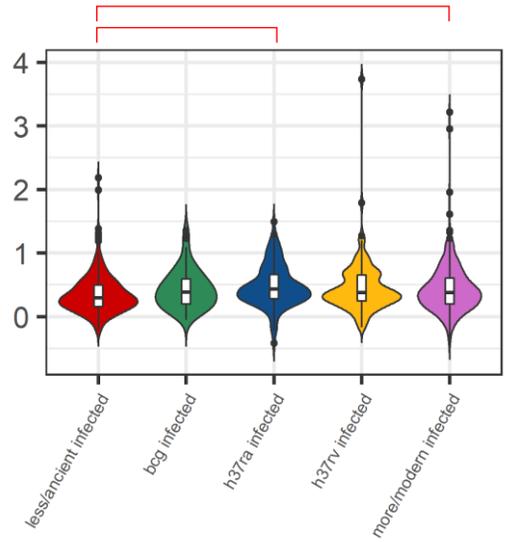
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.4775	0.134	0
groupbcg infected	0.4793	0.1543	0.0029
grouph37ra infected	0.6163	0.1567	2e-04
grouph37rv infected	0.0909	0.1604	0.5728
groupmore/modern infected	0.2335	0.0988	0.0214

Contrast	estimate	SE	p.value
less/ancient infected - bcg infected	-0.4793	0.1543	0.0096
less/ancient infected - h37ra infected	-0.6163	0.1567	0.0014
less/ancient infected - h37rv infected	-0.9909	0.1604	0.5728
less/ancient infected - more/modern infected	-0.2335	0.0988	0.0428
bcg infected - h37ra infected	-0.137	0.2153	0.5728
bcg infected - h37rv infected	0.3884	0.2177	0.1325
bcg infected - more/modern infected	0.2458	0.1797	0.2521
h37ra infected - h37rv infected	0.5254	0.1358	0.0014
h37ra infected - more/modern infected	0.3828	0.143	0.0239
h37rv infected - more/modern infected	-0.1426	0.142	0.3992

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	-0.2909	0.0544	0
groupbcg infected	0.2359	0.1063	0.0303
grouph37ra infected	0.4195	0.0929	0
grouph37rv infected	0.2242	0.0962	0.0232
groupmore/modern infected	0.2158	0.0678	0.0023

Contrast	estimate	SE	p.value
less/ancient infected - bcg infected	-0.2359	0.1063	0.0606
less/ancient infected - h37ra infected	-0.4195	0.0929	8e-04
less/ancient infected - h37rv infected	-0.2242	0.0962	0.0606
less/ancient infected - more/modern infected	-0.2158	0.0678	0.0116
bcg infected - h37ra infected	-0.1836	0.1265	0.2172
bcg infected - h37rv infected	0.0117	0.1281	0.9273
bcg infected - more/modern infected	0.0201	0.1106	0.9273
h37ra infected - h37rv infected	0.1953	0.106	0.1173
h37ra infected - more/modern infected	0.2037	0.0888	0.0606
h37rv infected - more/modern infected	0.0084	0.0909	0.9273

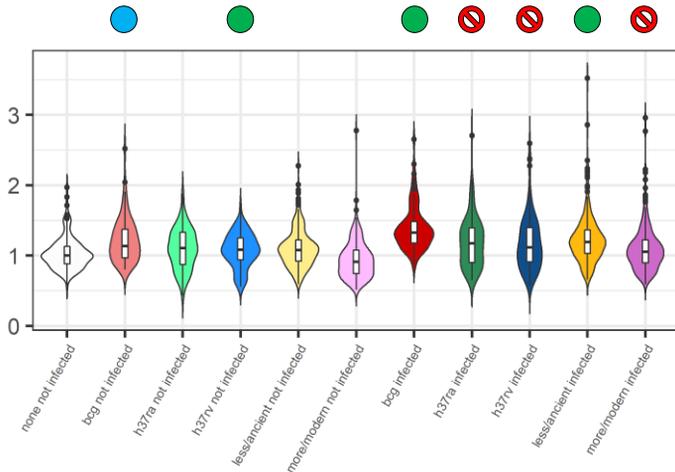
OBSERVATIONS

Less virulent ancient strains showed lower MOI at 6 h p.i. compared to the other groups. Colocalization with acidified compartments was similar among all the groups.

Grouping: Basic

M2 – 24 h p.i.

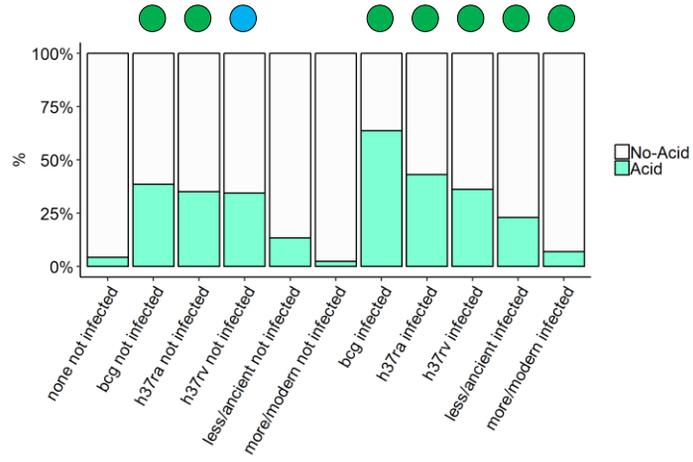
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	0.0131	0.0546	0.8106
groupbcg not infected	0.0701	0.0522	0.1795
grouph37ra not infected	-0.0471	0.0442	0.2871
grouph37rv not infected	-0.1166	0.0449	0.0095
groupless/ancient not infected	0.0251	0.0314	0.4241
groupmore/modern not infected	-0.0696	0.0316	0.0277
groupbcg infected	0.221	0.0516	0
grouph37ra infected	0.0611	0.0429	0.1542
grouph37rv infected	-0.0506	0.0438	0.2482
groupless/ancient infected	0.1256	0.0314	1e-04
groupmore/modern infected	0.0386	0.0312	0.2161

Contrast	estimate	SE	p.value
none not infected-bcg not infected	-0.0701	0.0522	0.2327
none not infected-h37ra not infected	0.0471	0.0442	0.3349
none not infected-h37rv not infected	0.1166	0.0449	0.0237
none not infected-less/ancient not infected	-0.0251	0.0314	0.4498
none not infected-more/modern not infected	0.0696	0.0316	0.0511
none not infected-bcg infected	0.221	0.0516	2e-04
none not infected-h37ra infected	-0.0611	0.0429	0.2159
none not infected-h37rv infected	0.0506	0.0438	0.2996
none not infected-less/ancient infected	-0.1256	0.0314	3e-04
none not infected-more/modern infected	-0.0386	0.0312	0.2701
bcg not infected-h37ra not infected	0.1172	0.0642	0.1086
bcg not infected-h37rv not infected	0.1867	0.0652	0.012
bcg not infected-less/ancient not infected	0.045	0.0486	0.3879
bcg not infected-more/modern not infected	0.1397	0.0544	0.024
h37ra not infected-h37rv not infected	0.0695	0.0479	0.2137
h37ra not infected-less/ancient not infected	-0.0722	0.0471	0.1914
h37ra not infected-more/modern not infected	0.0225	0.0451	0.6186
h37rv not infected-less/ancient not infected	-0.1417	0.0488	0.0119
h37rv not infected-more/modern not infected	-0.0471	0.046	0.3455
less/ancient not infected-more/modern not infected	0.0947	0.0309	0.0078
bcg infected-h37ra infected	0.1599	0.0629	0.0242
bcg infected-h37rv infected	0.2716	0.064	2e-04
bcg infected-less/ancient infected	0.0954	0.0479	0.0779
bcg infected-more/modern infected	0.1824	0.0535	0.0026
h37ra infected-h37rv infected	0.1117	0.0457	0.0286
h37ra infected-less/ancient infected	-0.0645	0.046	0.2167
h37ra infected-more/modern infected	0.0225	0.0437	0.6186
h37rv infected-less/ancient infected	-0.1762	0.0478	0.001
h37rv infected-more/modern infected	-0.0892	0.0444	0.0779
less/ancient infected-more/modern infected	0.087	0.0306	0.012
bcg not infected-bcg infected	-0.1509	0.0295	0
h37ra not infected-h37ra infected	-0.1082	0.0259	2e-04
h37rv not infected-h37rv infected	-0.066	0.0263	0.0249
less/ancient not infected-less/ancient infected	-0.1005	0.0158	0
more/modern not infected-more/modern infected	-0.1081	0.0176	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.5118	0.6157	0.0298	0
groupbcg not infected	1.7216	0.5477	3.5932	0.0017
grouph37ra not infected	1.4539	0.585	4.2798	0.0129
grouph37rv not infected	1.2307	0.5873	3.4235	0.0361
groupless/ancient not infected	1.2008	0.5041	3.3227	0.0172
groupmore/modern not infected	0.4251	0.6559	1.5297	0.5169
groupbcg infected	2.7484	0.5419	15.6178	0
grouph37ra infected	2.3708	0.5715	10.7059	0
grouph37rv infected	1.7556	0.5749	5.7871	0.0023
groupless/ancient infected	1.9427	0.4906	6.9778	1e-04
groupmore/modern infected	1.4646	0.5336	4.3258	0.0061

Contrast	estimate	SE	p.value
none not infected-bcg not infected	-1.7216	0.5477	0.013
none not infected-h37ra not infected	-1.4539	0.585	0.0412
none not infected-h37rv not infected	-1.2307	0.5873	0.0843
none not infected-less/ancient not infected	-1.2008	0.5041	0.0464
none not infected-more/modern not infected	-0.4251	0.6559	0.6238
none not infected-bcg infected	2.7484	0.5419	0
none not infected-h37ra infected	2.3708	0.5715	6e-04
none not infected-h37rv infected	1.7556	0.5749	0.013
none not infected-less/ancient infected	-1.9427	0.4906	9e-04
none not infected-more/modern infected	-1.4646	0.5336	0.0236
bcg not infected-h37ra not infected	0.2676	0.594	0.6918
bcg not infected-h37rv not infected	0.4909	0.592	0.5349
bcg not infected-less/ancient not infected	0.5208	0.3434	0.2157
bcg not infected-more/modern not infected	1.2965	0.5924	0.0716
h37ra not infected-h37rv not infected	0.2233	0.3902	0.6404
h37ra not infected-less/ancient not infected	0.2531	0.5402	0.6918
h37ra not infected-more/modern not infected	1.0288	0.6494	0.2021
h37rv not infected-less/ancient not infected	0.0299	0.5461	0.9564
h37rv not infected-more/modern not infected	0.8056	0.6525	0.3037
less/ancient not infected-more/modern not infected	0.7757	0.5309	0.2191
bcg infected-h37ra infected	0.3776	0.5684	0.6238
bcg infected-h37rv infected	0.9928	0.5778	0.1667
bcg infected-less/ancient infected	0.8057	0.3235	0.0412
bcg infected-more/modern infected	1.2838	0.4482	0.0183
h37ra infected-h37rv infected	0.6152	0.3908	0.2021
h37ra infected-less/ancient infected	0.4281	0.5	0.5276
h37ra infected-more/modern infected	0.9062	0.4999	0.1438
h37rv infected-less/ancient infected	-0.1871	0.5115	0.7355
h37rv infected-more/modern infected	0.291	0.5021	0.6404
less/ancient infected-more/modern infected	0.4781	0.3476	0.2463
bcg not infected-bcg infected	-1.0269	0.3387	0.013
h37ra not infected-h37ra infected	-0.9169	0.3755	0.0426
h37rv not infected-h37rv infected	-0.525	0.3581	0.2191
less/ancient not infected-less/ancient infected	-0.7419	0.2464	0.013
more/modern not infected-more/modern infected	-1.0395	0.5248	0.1042

OBSERVATIONS

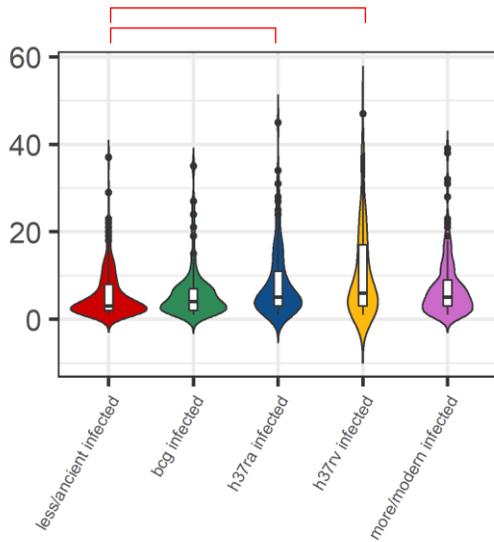
At 24 h p.i. only H37Rv, H37Ra and more virulent/modern strains displayed a blockade of the acidification of the phagolysosomes; however, whereas the percentage of acidified cells remained high for BCG (60%), H37Rv and H37Ra (40%), it dropped for less virulent and ancient strains (25%) and for more virulent and modern strains (<10%).

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Basic

M2 – 24 h.p.i.

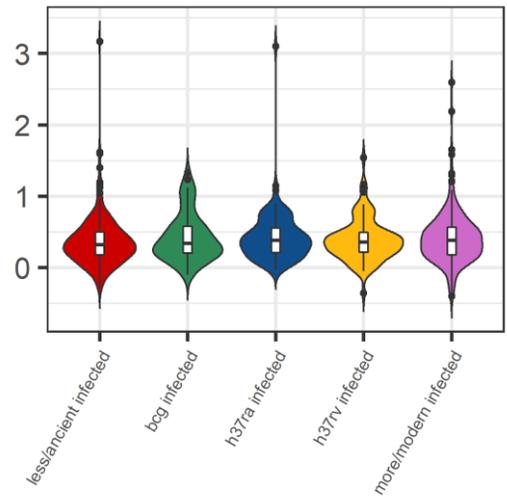
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.3419	0.1293	0
groupbcg infected	0.1459	0.1918	0.4498
grouph37ra infected	0.5265	0.1857	0.0062
grouph37rv infected	0.5935	0.1938	0.0033
groupmore/modern infected	0.1622	0.1228	0.1913

Contrast	estimate	SE	p.value
less/ancient infected - bcg infected	-0.1459	0.1918	0.5622
less/ancient infected - h37ra infected	-0.5265	0.1857	0.0312
less/ancient infected - h37rv infected	-0.5935	0.1938	0.0312
less/ancient infected - more/modern infected	-0.1622	0.1228	0.2733
bcg infected - h37ra infected	-0.3806	0.2569	0.2396
bcg infected - h37rv infected	-0.4476	0.2621	0.1856
bcg infected - more/modern infected	-0.0163	0.2196	0.9411
h37ra infected - h37rv infected	-0.067	0.1794	0.7891
h37ra infected - more/modern infected	0.3643	0.1749	0.1037
h37rv infected - more/modern infected	0.4313	0.178	0.0615

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	-0.1921	0.0656	0.0035
groupbcg infected	0.2212	0.1216	0.074
grouph37ra infected	0.1895	0.1101	0.0904
grouph37rv infected	0.069	0.1169	0.5571
groupmore/modern infected	0.1861	0.0832	0.0291

Contrast	estimate	SE	p.value
less/ancient infected - bcg infected	-0.2212	0.1216	0.3014
less/ancient infected - h37ra infected	-0.1895	0.1101	0.3014
less/ancient infected - h37rv infected	-0.069	0.1169	0.7958
less/ancient infected - more/modern infected	-0.1861	0.0832	0.2909
bcg infected - h37ra infected	0.0317	0.1507	0.9267
bcg infected - h37rv infected	0.1522	0.155	0.5503
bcg infected - more/modern infected	0.0351	0.1348	0.9267
h37ra infected - h37rv infected	0.1205	0.1193	0.5503
h37ra infected - more/modern infected	0.0033	0.1089	0.9756
h37rv infected - more/modern infected	-0.1171	0.1143	0.5503

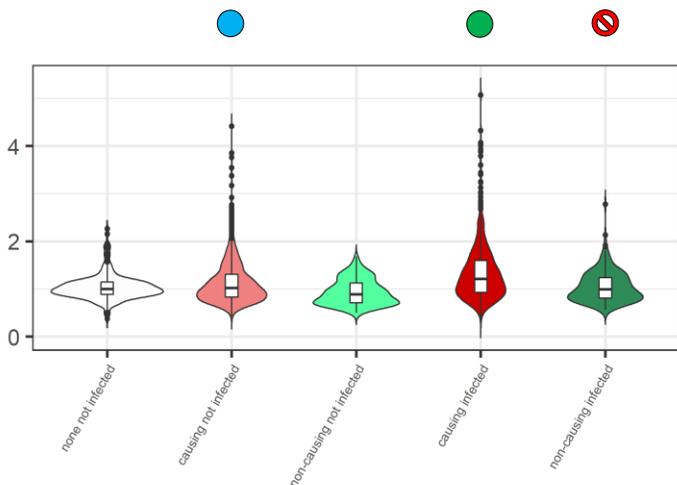
OBSERVATIONS

Less virulent ancient strains showed lower MOI compared to H37Ra and H37Rv only. Colocalization with acidified compartments was similar among all the groups.

Grouping: Disease

M1 – 6 h p.i.

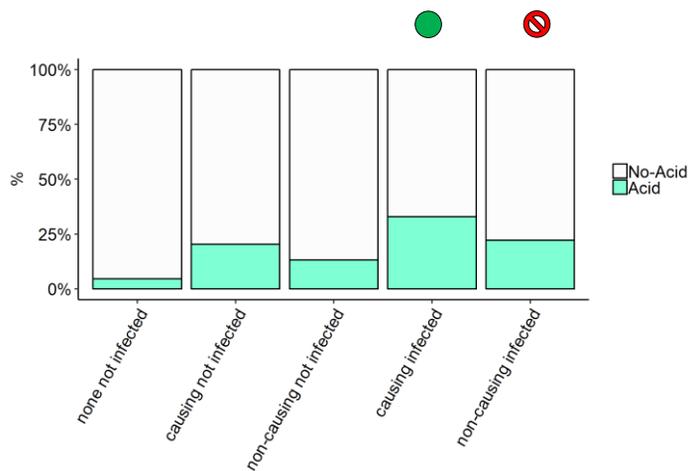
a) Phagolysosomal acidification



Par	Value	Std.Error	p-value
(Intercept)	0.0033	0.0806	0.9677
groupcausing not infected	0.0601	0.0468	0.1988
groupnon-causing not infected	-0.1065	0.0655	0.1044
groupcausing infected	0.1912	0.0468	0
groupnon-causing infected	-0.0322	0.0657	0.624

Contrast	estimate	SE	p-value
none not infected-causing not infected	-0.0601	0.0468	0.2272
none not infected-non-causing not infected	0.1065	0.0655	0.1392
none not infected-causing infected	-0.1912	0.0468	1e-04
none not infected-non-causing infected	0.0322	0.0657	0.624
causing not infected-non-causing not infected	0.1666	0.0602	0.0091
causing infected-non-causing not infected	0.2234	0.0604	4e-04
causing not infected-causing infected	-0.1311	0.0091	0
non-causing not infected-non-causing infected	-0.0743	0.0176	1e-04

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.6546	0.8901	0.0259	0
groupcausing not infected	1.0725	0.5749	2.9227	0.0621
groupnon-causing not infected	0.2613	0.7424	1.2986	0.7249
groupcausing infected	2.3925	0.572	10.9406	0
groupnon-causing infected	0.933	0.729	2.5422	0.2006

Contrast	estimate	SE	p-value
none not infected-causing not infected	-1.0725	0.5749	0.0993
none not infected-non-causing not infected	-0.2613	0.7424	0.7249
none not infected-causing infected	-2.3925	0.572	1e-04
none not infected-non-causing infected	-0.933	0.729	0.2293
causing not infected-non-causing not infected	0.8112	0.6205	0.2293
causing infected-non-causing not infected	1.4595	0.602	0.0409
causing not infected-causing infected	-1.32	0.1771	0
non-causing not infected-non-causing infected	-0.6717	0.3259	0.0785

OBSERVATIONS

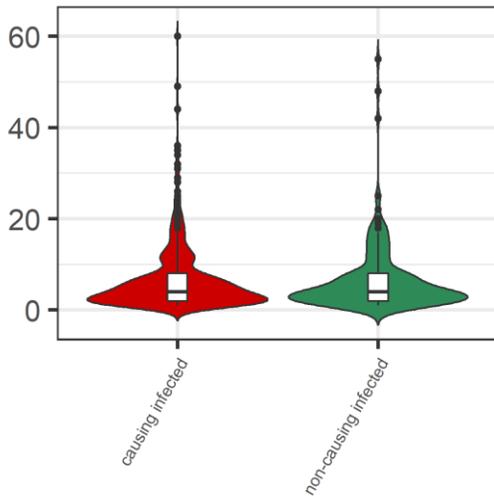
In M1 macrophages, non-causing disease strains are blocking the phagolysosomal acidification at 6 h p.i., but the percentage of acidified cells remains low (30%) also in the category disease causing.

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Disease

M1 – 6 h p.i.

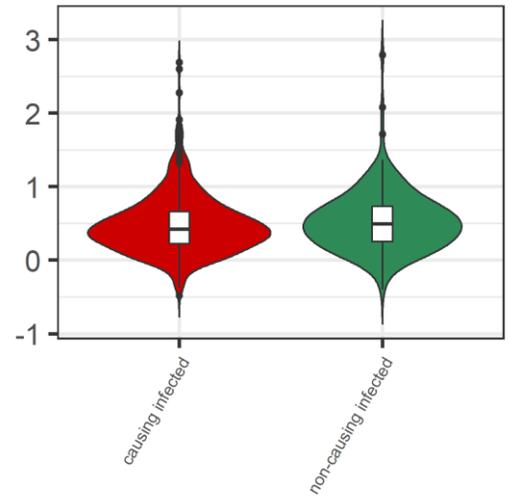
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.4382	0.0858	0
groupnon-causing infected	0.0226	0.1292	0.8619

Contrast	estimate	SE	p.value
causing infected - non-causing infected	-0.0226	0.1292	0.8619

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	0.0046	0.0699	0.9479
groupnon-causing infected	0.1905	0.1097	0.0874

Contrast	estimate	SE	p.value
causing infected - non-causing infected	-0.1905	0.1097	0.0874

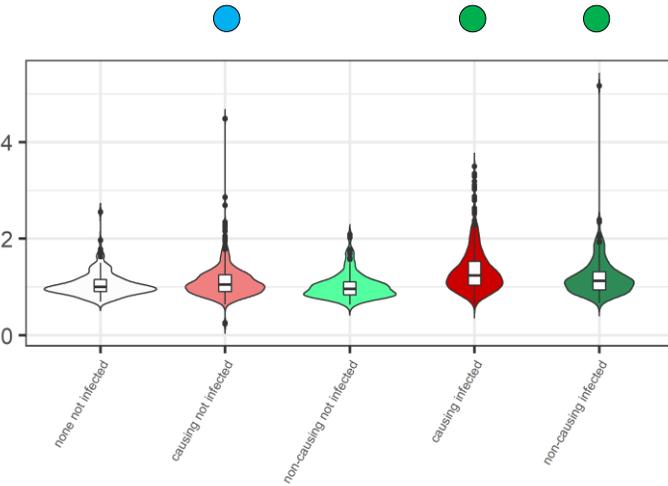
OBSERVATIONS

Non-causing disease strains showed higher MOI values. Colocalization with acidified compartments was similar among all the groups.

Grouping: Disease

M1 – 24 h p.i.

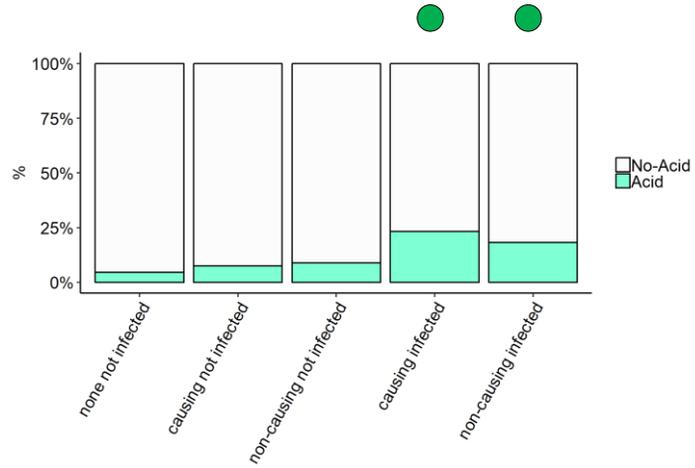
a) Phagolysosomal acidification



Par	Value	Std.Error	p-value
(Intercept)	0.0097	0.0408	0.8124
groupecausing not infected	0.0341	0.0281	0.225
groupnon-causing not infected	-0.0209	0.0389	0.5903
groupecausing infected	0.1785	0.0279	0
groupnon-causing infected	0.1265	0.0376	8e-04

Contrast	estimate	SE	p-value
none not infected-causing not infected	-0.0341	0.0281	0.2572
none not infected-non-causing not infected	0.0209	0.0389	0.5903
none not infected-causing infected	-0.1785	0.0279	0
none not infected-non-causing infected	-0.1265	0.0376	0.0015
causing not infected-non-causing not infected	0.055	0.0342	0.1492
causing infected-non-causing not infected	0.052	0.0327	0.1492
causing not infected-causing infected	-0.1444	0.0122	0
non-causing not infected-non-causing infected	-0.1475	0.0236	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.186	0.456	0.0413	0
groupecausing not infected	0.4347	0.4456	1.5446	0.3292
groupnon-causing not infected	0.9095	0.5479	2.4832	0.0969
groupecausing infected	1.8308	0.4246	6.2387	0
groupnon-causing infected	1.7686	0.4844	5.8628	3e-04

Contrast	estimate	SE	p-value
none not infected-causing not infected	-0.4347	0.4456	0.3763
none not infected-non-causing not infected	-0.9095	0.5479	0.155
none not infected-causing infected	-1.8308	0.4246	1e-04
none not infected-non-causing infected	-1.7686	0.4844	7e-04
causing not infected-non-causing not infected	-0.4748	0.4191	0.343
causing infected-non-causing not infected	0.0621	0.3017	0.8368
causing not infected-causing infected	1.396	0.1963	0
non-causing not infected-non-causing infected	-0.8591	0.3752	0.0441

OBSERVATIONS

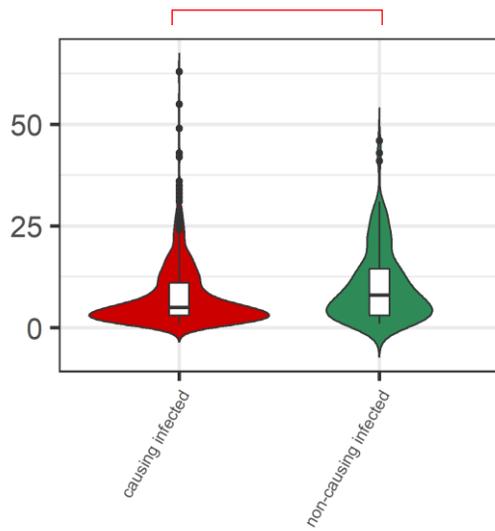
At 24 h p.i. none of the category is blocking the acidification of the phagolysosomes. However, despite an increase in the acidification of the phagolysosomes was observed, the number of macrophages displaying such acidification was found generally low (<25%).

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Disease

M1 – 24 h p.i.

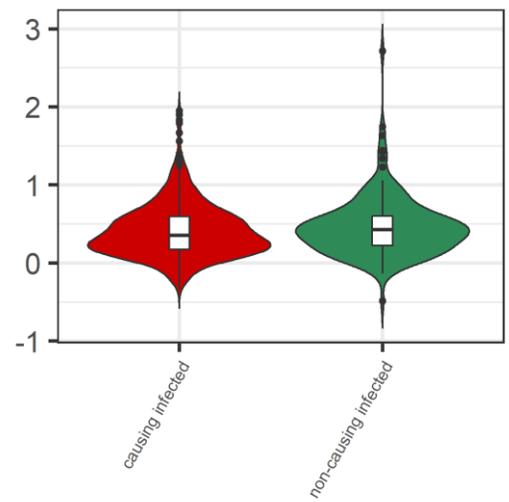
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.5863	0.1249	0
groupnon-causing infected	0.4806	0.0974	0

Contrast	estimate	SE	p.value
causing infected - non-causing infected	-0.4806	0.0974	0

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	-0.0421	0.0717	0.5571
groupnon-causing infected	0.1552	0.1161	0.1859

Contrast	estimate	SE	p.value
causing infected - non-causing infected	-0.1552	0.1161	0.1859

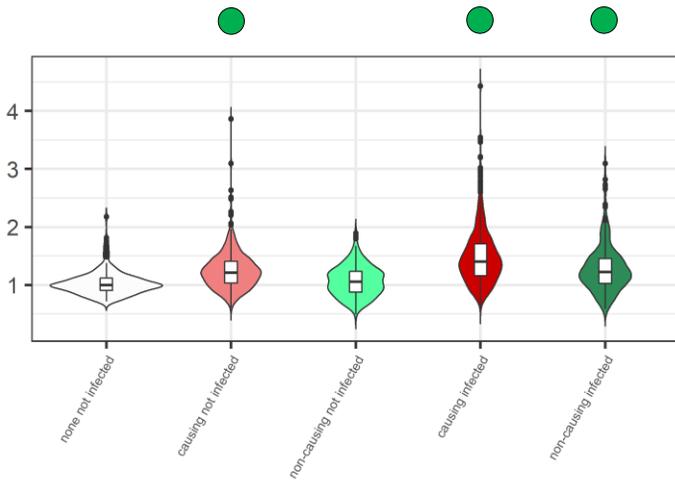
OBSERVATIONS

Non-causing disease strains showed higher MOI values. Colocalization with acidified compartments was similar among all the groups.

Grouping: Disease

M2 – 6 h p.i.

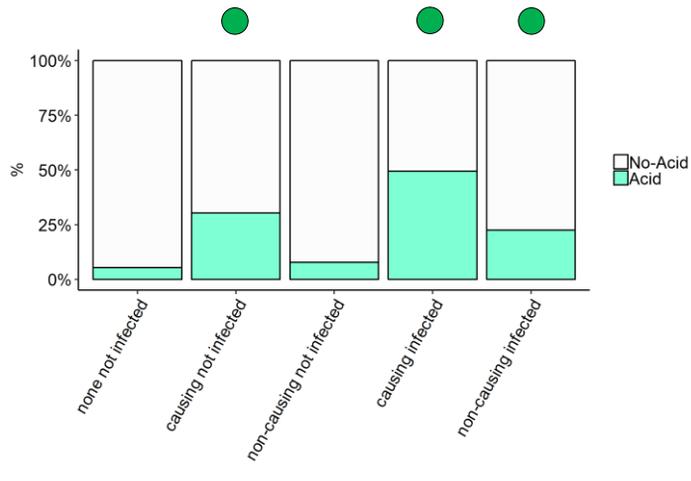
a) Phagolysosomal acidification



Par	Value	Std.Error	p-value
(Intercept)	-0.0092	0.0487	0.8495
groupcausing not infected	0.1646	0.0377	0
groupnon-causing not infected	0.0662	0.0521	0.2044
groupcausing infected	0.2905	0.0377	0
groupnon-causing infected	0.2055	0.0523	1e-04

Contrast	estimate	SE	p-value
none not infected-causing not infected	-0.1646	0.0377	0
none not infected-non-causing not infected	-0.0662	0.0521	0.2044
none not infected-causing infected	-0.2905	0.0377	0
none not infected-non-causing infected	-0.2055	0.0523	1e-04
causing not infected-non-causing not infected	0.0984	0.0479	0.0536
causing infected-non-causing infected	0.085	0.0481	0.0883
causing not infected-causing infected	-0.1259	0.0088	0
non-causing not infected-non-causing infected	-0.1393	0.0168	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.197	0.6331	0.0409	0
groupcausing not infected	1.7001	0.5059	5.4745	8e-04
groupnon-causing not infected	1.179	0.6744	3.2511	0.0804
groupcausing infected	2.9537	0.5039	19.1775	0
groupnon-causing infected	2.5538	0.6396	12.8565	1e-04

Contrast	estimate	SE	p-value
none not infected-causing not infected	-1.7001	0.5059	0.0012
none not infected-non-causing not infected	-1.179	0.6744	0.1072
none not infected-causing infected	-2.9537	0.5039	0
none not infected-non-causing infected	-2.5538	0.6396	1e-04
causing not infected-non-causing not infected	0.5211	0.5464	0.3889
causing infected-non-causing infected	0.3999	0.4975	0.4214
causing not infected-causing infected	-1.2536	0.1449	0
non-causing not infected-non-causing infected	-1.3749	0.3264	1e-04

OBSERVATIONS

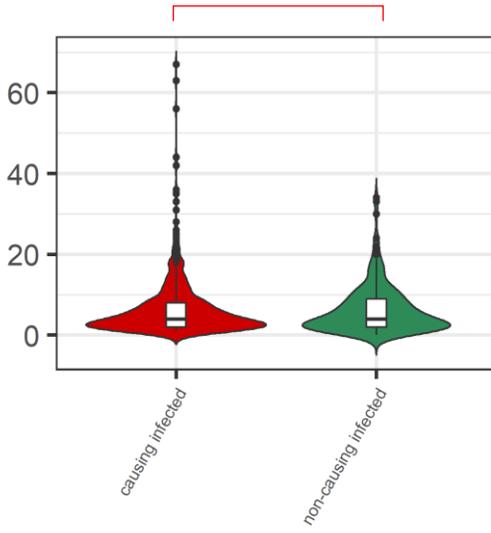
In M2 macrophages, none of the category is blocking the acidification of the phagolysosomes at 6 h p.i.. Whereas the category causing disease displayed acidification in 50% of cells, non-causing disease strains showed acidification in less than 25% of macrophages.

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, stastically not significant

Grouping: Disease

M2 – 6 h p.i.

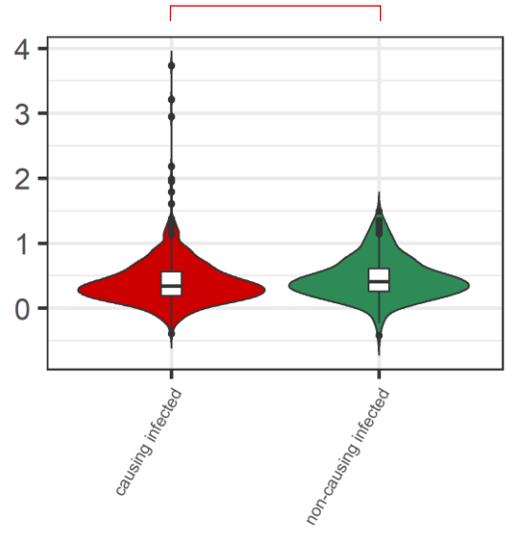
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.5863	0.1249	0
groupnon-causing infected	0.4806	0.0974	0

Contrast	estimate	SE	p.value
causing infected - non-causing infected	-0.4806	0.0974	0

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	-0.1642	0.0535	0.0022
groupnon-causing infected	0.2582	0.0714	6e-04

Contrast	estimate	SE	p.value
causing infected - non-causing infected	-0.2582	0.0714	6e-04

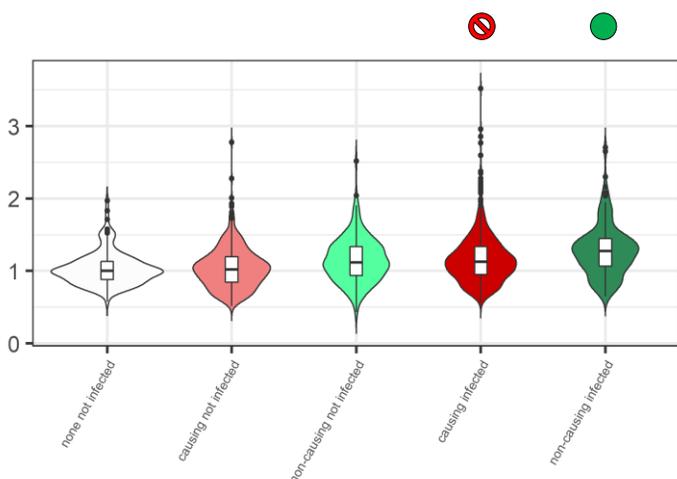
OBSERVATIONS

Non-causing disease strains showed also higher MOI values. Colocalization with acidified compartments was found lower for disease causing strains.

Grouping: Disease

M2 – 24 h.p.i.

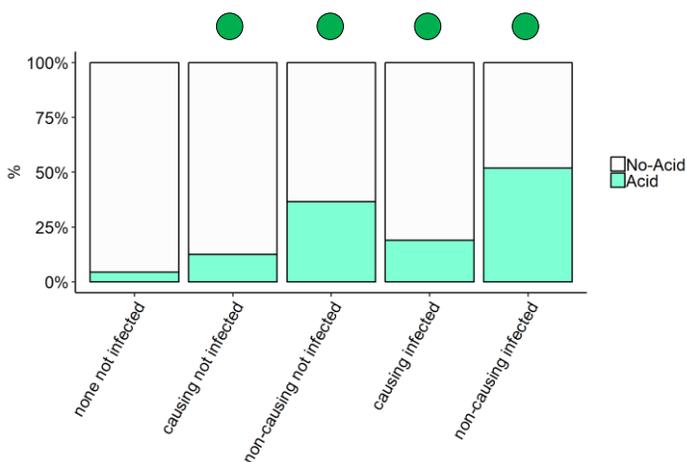
a) Phagolysosomal acidification



Par	Value	Std.Error	p-value
(Intercept)	0.0118	0.057	0.8355
groupcausing not infected	-0.0395	0.0277	0.1538
groupnon-causing not infected	-0.0037	0.0378	0.9227
groupcausing infected	0.0537	0.0276	0.052
groupnon-causing infected	0.121	0.0371	0.0011

Contrast	estimate	SE	p-value
none not infected-causing not infected	0.0395	0.0277	0.2051
none not infected-non-causing not infected	0.0037	0.0378	0.9227
none not infected-causing infected	-0.0537	0.0276	0.0833
none not infected-non-causing infected	-0.121	0.0371	0.003
causing not infected-non-causing not infected	-0.0358	0.0334	0.3235
causing infected-non-causing infected	-0.0674	0.0325	0.0772
causing not infected-causing infected	-0.0932	0.0107	0
non-causing not infected-non-causing infected	-0.1247	0.0194	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.6245	0.6384	0.0267	0
groupcausing not infected	1.1272	0.4645	3.087	0.0152
groupnon-causing not infected	1.5586	0.4802	4.7521	0.0012
groupcausing infected	1.8374	0.4592	6.2804	1e-04
groupnon-causing infected	2.5484	0.4778	12.7873	0

Contrast	estimate	SE	p-value
none not infected-causing not infected	-1.1272	0.4645	0.0174
none not infected-non-causing not infected	-1.5586	0.4802	0.0019
none not infected-causing infected	-1.8374	0.4592	2e-04
none not infected-non-causing infected	-2.5484	0.4778	0
causing not infected-non-causing not infected	-0.4314	0.2499	0.0843
causing infected-non-causing infected	-0.711	0.2394	0.004
causing not infected-causing infected	-0.7102	0.1873	3e-04
non-causing not infected-non-causing infected	-0.9899	0.2525	2e-04

OBSERVATIONS

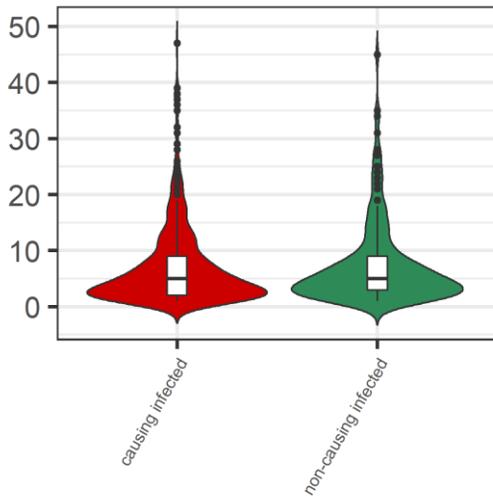
As the infection progresses, disease causing strains are blocking the acidification of the phagolysosomes. In this case, the percentage of macrophages displaying acidification was higher for non-causing disease strains (50%) compared to causing disease strains (<25%).

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Disease

M2 – 24 h p.i.

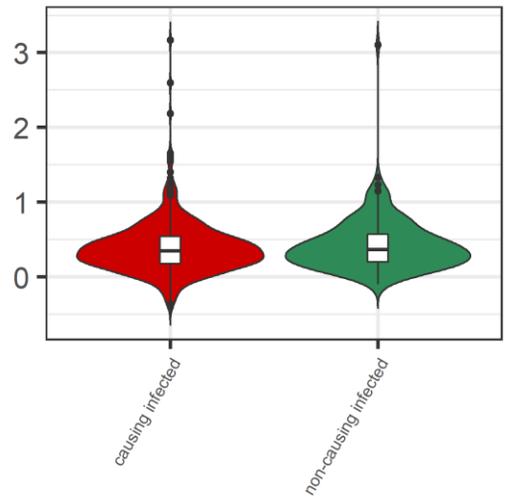
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.5343	0.1207	0
groupnon-causing infected	0.1865	0.1183	0.1198

Contrast	estimate	SE	p.value
causing infected - non-causing infected	-0.1865	0.1183	0.1198

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	-0.1051	0.047	0.0258
groupnon-causing infected	0.1232	0.0796	0.1267

Contrast	estimate	SE	p.value
causing infected - non-causing infected	-0.1232	0.0796	0.1267

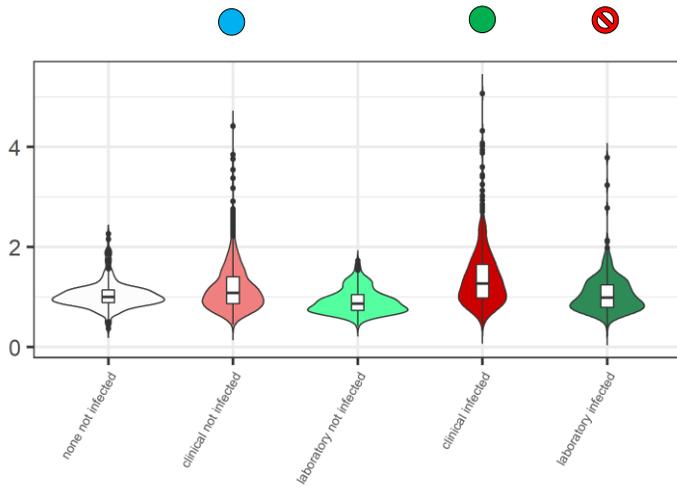
OBSERVATIONS

Colocalization with acidified compartments was similar among all the groups.

Grouping: Lab adaptation

M1 – 6 h p.i.

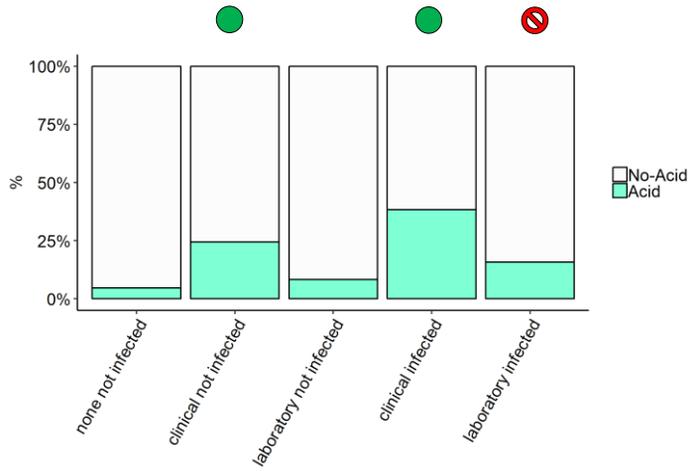
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	0.0048	0.0752	0.9496
groupclinical not infected	0.0872	0.0495	0.0782
grouplaboratory not infected	-0.0864	0.0584	0.139
groupclinical infected	0.2227	0.0495	0
grouplaboratory infected	3e-04	0.0585	0.9957

Contrast	estimate	SE	p.value
none not infected-clinical not infected	-0.0872	0.0495	0.1043
none not infected-laboratory not infected	0.0864	0.0584	0.1589
none not infected-clinical infected	-0.2227	0.0495	0
none not infected-laboratory infected	-3e-04	0.0585	0.9957
clinical not infected-laboratory not infected	0.1735	0.0575	0.0041
clinical infected-laboratory infected	0.2223	0.0576	2e-04
clinical not infected-clinical infected	-0.1355	0.0099	0
laboratory not infected-laboratory infected	-0.0867	0.014	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.6022	0.8358	0.0273	0
groupclinical not infected	1.379	0.5906	3.9711	0.0195
grouplaboratory not infected	0.043	0.6915	1.0439	0.9504
groupclinical infected	2.6821	0.5897	14.6163	0
grouplaboratory infected	0.8566	0.6756	2.3551	0.2049

Contrast	estimate	SE	p.value
none not infected-clinical not infected	-1.379	0.5906	0.0313
none not infected-laboratory not infected	-0.043	0.6915	0.9504
none not infected-clinical infected	-2.6821	0.5897	0
none not infected-laboratory infected	-0.8566	0.6756	0.2341
clinical not infected-laboratory not infected	1.336	0.6205	0.0417
clinical infected-laboratory infected	1.8256	0.6025	0.0065
clinical not infected-clinical infected	-1.3031	0.1806	0
laboratory not infected-laboratory infected	-0.8136	0.3045	0.0151

OBSERVATIONS

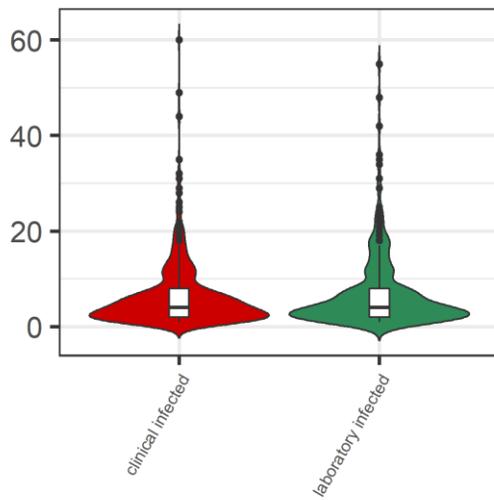
In M1 macrophages, laboratory strains are blocking the phagolysosomal acidification at 6 h p.i. but the percentage of acidified cells remains low (30%) also in the category of clinical strains.

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Lab adaptation

M1 – 6 h p.i.

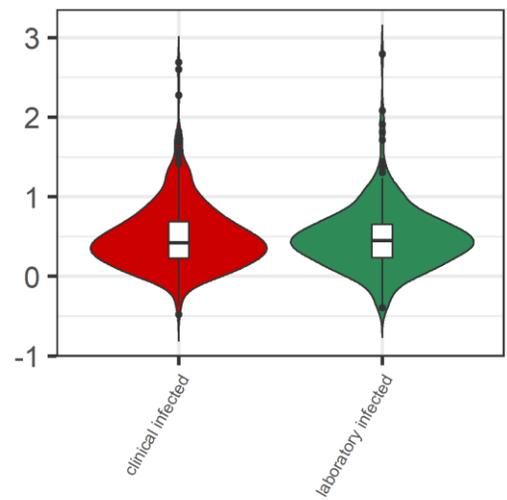
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.4239	0.0933	0
grouplaboratory infected	0.0521	0.1209	0.6679

Contrast	estimate	SE	p.value
clinical infected - laboratory infected	-0.0521	0.1209	0.6679

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	0.0535	0.08	0.5039
grouplaboratory infected	-0.034	0.103	0.7425

Contrast	estimate	SE	p.value
clinical infected - laboratory infected	0.034	0.103	0.7425

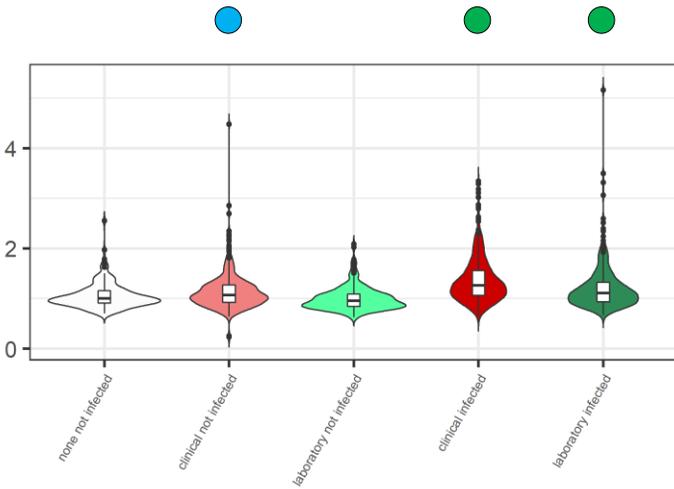
OBSERVATIONS

Colocalization with acidified compartments was similar among all the groups. Laboratory strains showed higher MOI values.

Grouping: Lab adaptation

M1 – 24 h p.i.

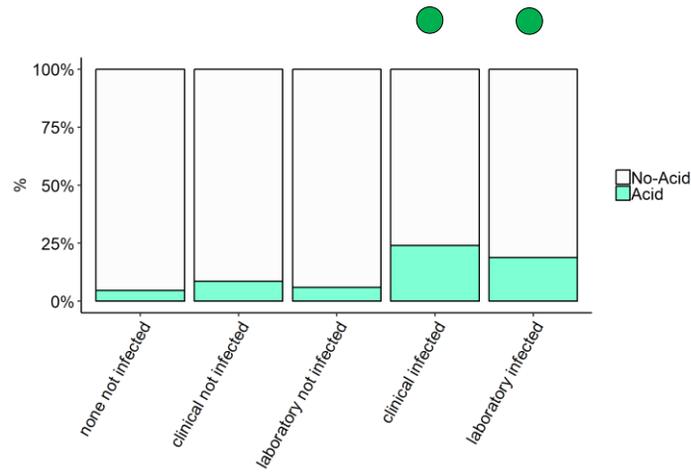
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	0.0141	0.0388	0.7169
groupclinical not infected	0.0404	0.0293	0.1686
grouplaboratory not infected	-0.0191	0.0352	0.5874
groupclinical infected	0.1864	0.0292	0
grouplaboratory infected	0.1256	0.0344	3e-04

Contrast	estimate	SE	p.value
none not infected-clinical not infected	-0.0404	0.0293	0.1927
none not infected-laboratory not infected	0.0191	0.0352	0.5874
none not infected-clinical infected	-0.1864	0.0292	0
none not infected-laboratory infected	-0.1256	0.0344	3e-04
clinical not infected-laboratory not infected	0.0595	0.0325	0.0902
clinical infected-laboratory not infected	0.0608	0.0316	0.0875
clinical not infected-clinical infected	-0.1459	0.0134	0
laboratory not infected-laboratory infected	-0.1447	0.0186	0

b) Percentage of cells showing acidified phagolysosomes



Contrast	estimate	SE	p.value
none not infected-clinical not infected	-0.5354	0.4509	0.3761
none not infected-laboratory not infected	-0.485	0.5275	0.4771
none not infected-clinical infected	-1.8155	0.4306	1e-04
none not infected-laboratory infected	-1.8084	0.4605	2e-04
clinical not infected-laboratory not infected	0.0504	0.4028	0.9796
clinical infected-laboratory not infected	0.0071	0.2774	0.9796
clinical not infected-clinical infected	-1.2801	0.2035	0
laboratory not infected-laboratory infected	-1.3234	0.3381	2e-04

OBSERVATIONS

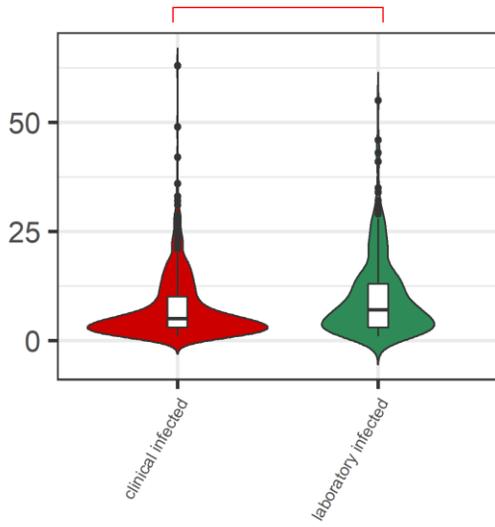
At 24 h p.i. none of the category is blocking the acidification of the phagolysosomes. However, despite an increase in the acidification of the phagolysosomes was observed, the number of macrophages displaying such acidification was found generally low (<25%).

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, stastically not significant

Grouping: Lab adaptation

M1 – 24 h p.i.

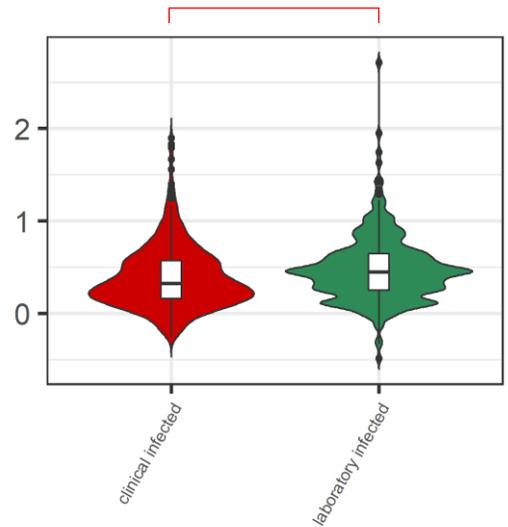
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.5888	0.1164	0
grouplaboratory infected	0.2835	0.1099	0.0122

Contrast	estimate	SE	p.value
clinical infected - laboratory infected	-0.2835	0.1099	0.0122

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	-0.1587	0.08	0.0478
grouplaboratory infected	0.4017	0.1017	2e-04

Contrast	estimate	SE	p.value
clinical infected - laboratory infected	-0.4017	0.1017	2e-04

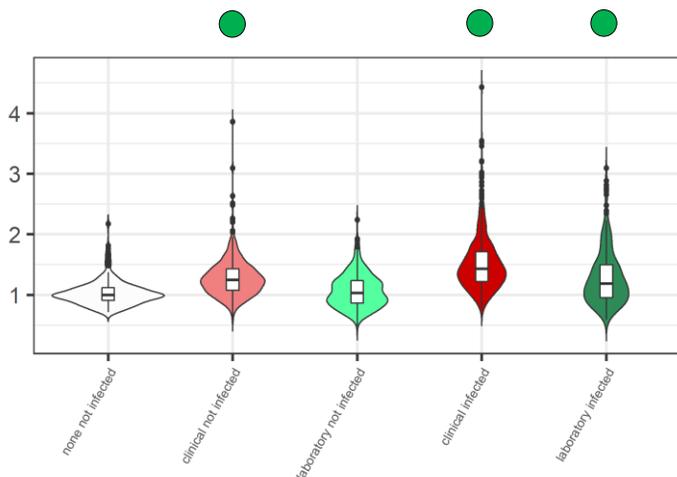
OBSERVATIONS

Laboratory strains showed higher MOI values. Colocalization with acidified compartments was higher for laboratory strains.

Grouping: Lab adaptation

M2 – 6 h p.i.

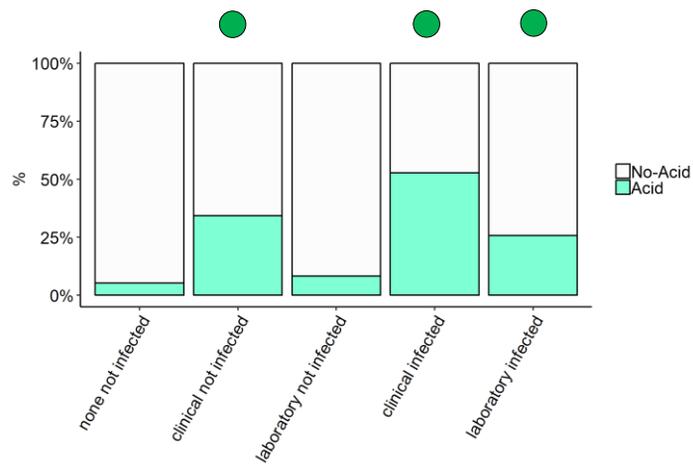
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	-0.0099	0.0439	0.8208
groupclinical not infected	0.1958	0.039	0
grouplaboratory not infected	0.0543	0.045	0.2276
groupclinical infected	0.3222	0.039	0
grouplaboratory infected	0.1888	0.0451	0

Contrast	estimate	SE	p.value
none not infected-clinical not infected	-0.1958	0.039	0
none not infected-laboratory not infected	-0.0543	0.045	0.2276
none not infected-clinical infected	-0.3222	0.039	0
none not infected-laboratory infected	-0.1888	0.0451	0
clinical not infected-laboratory not infected	0.1414	0.0437	0.0016
clinical infected-laboratory infected	0.1334	0.0438	0.0027
clinical not infected-clinical infected	-0.1265	0.0095	0
laboratory not infected-laboratory infected	-0.1345	0.0134	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.1627	0.5886	0.0423	0
groupclinical not infected	1.9879	0.5178	7.3005	1e-04
grouplaboratory not infected	0.6482	0.608	1.912	0.2864
groupclinical infected	3.1213	0.5166	22.6749	0
grouplaboratory infected	2.3513	0.5791	10.4992	0

Contrast	estimate	SE	p.value
none not infected-clinical not infected	-1.9879	0.5178	2e-04
none not infected-laboratory not infected	-0.6482	0.608	0.2864
none not infected-clinical infected	-3.1213	0.5166	0
none not infected-laboratory infected	-2.3513	0.5791	1e-04
clinical not infected-laboratory not infected	1.3398	0.5031	0.0103
clinical infected-laboratory infected	0.77	0.4652	0.1119
clinical not infected-clinical infected	-1.1333	0.1521	0
laboratory not infected-laboratory infected	-1.7031	0.2761	0

OBSERVATIONS

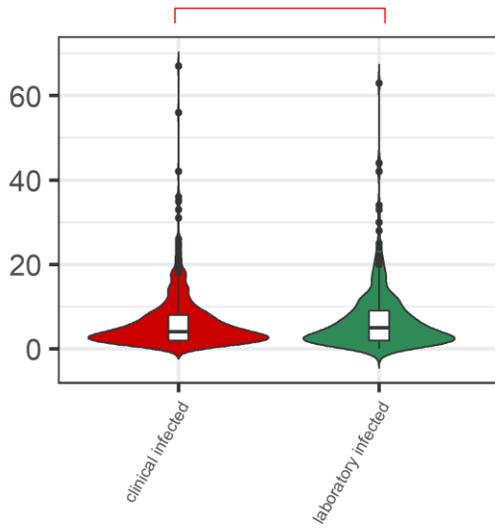
In M2 macrophages, none of the category is blocking the acidification of the phagolysosomes at 6 h p.i.. Whereas the category of clinical strains displayed acidification in 50% of cells, laboratory strains showed acidification in approx. 25% of macrophages.

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Lab adaptation

M2 – 6 h p.i.

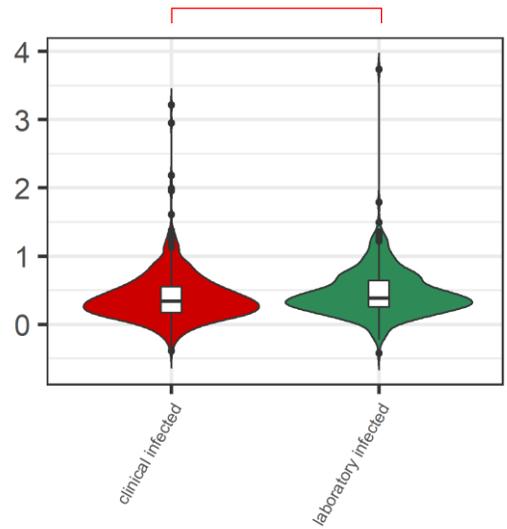
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.5888	0.1164	0
grouplaboratory infected	0.2835	0.1099	0.0122

Contrast	estimate	SE	p.value
clinical infected - laboratory infected	-0.2835	0.1099	0.0122

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	-0.1716	0.0503	7e-04
grouplaboratory infected	0.1868	0.0671	0.0071

Contrast	estimate	SE	p.value
clinical infected - laboratory infected	-0.1868	0.0671	0.0071

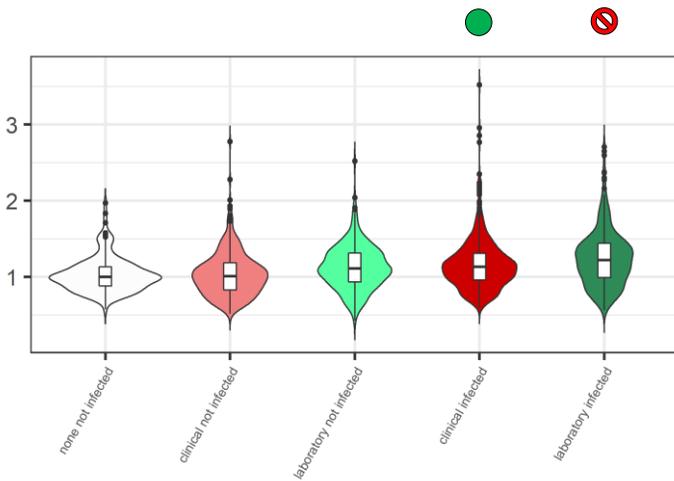
OBSERVATIONS

Colocalization with acidified compartments was higher for laboratory strains. Laboratory strains showed higher MOI values.

Grouping: Lab adaptation

M2 – 24 h p.i.

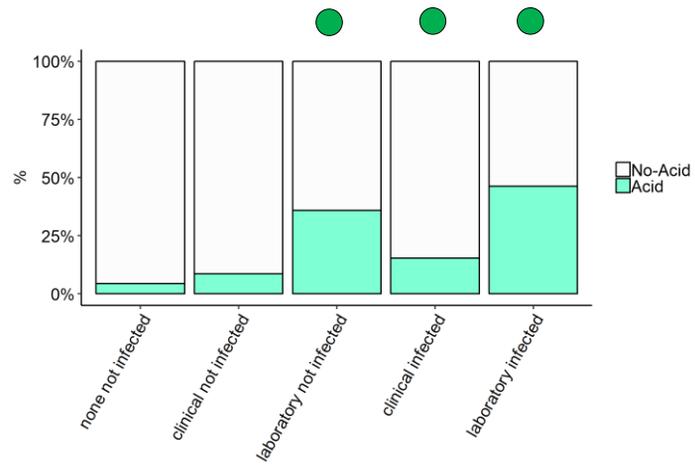
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	0.0083	0.0569	0.8839
groupclinical not infected	-0.0263	0.0293	0.3689
grouplaboratory not infected	-0.0328	0.0349	0.348
groupclinical infected	0.0732	0.0292	0.0123
grouplaboratory infected	0.0694	0.0344	0.0436

Contrast	estimate	SE	p.value
none not infected-clinical not infected	0.0263	0.0293	0.4918
none not infected-laboratory not infected	0.0328	0.0349	0.4918
none not infected-clinical infected	0.0732	0.0292	0.0329
none not infected-laboratory infected	-0.0694	0.0344	0.0872
clinical not infected-laboratory not infected	0.0064	0.0329	0.908
clinical infected-laboratory infected	0.0037	0.0323	0.908
clinical not infected-clinical infected	-0.0995	0.0117	0
laboratory not infected-laboratory infected	-0.1022	0.0156	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.5886	0.6313	0.0276	0
groupclinical not infected	0.9306	0.4839	2.536	0.0545
grouplaboratory not infected	1.5423	0.4771	4.6754	0.0012
groupclinical infected	1.7192	0.4723	5.5801	3e-04
grouplaboratory infected	2.3722	0.4727	10.7213	0

Contrast	estimate	SE	p.value
none not infected-clinical not infected	-0.9306	0.4839	0.0545
none not infected-laboratory not infected	-1.5423	0.4771	0.002
none not infected-clinical infected	-1.7192	0.4723	7e-04
none not infected-laboratory infected	-2.3722	0.4727	0
clinical not infected-laboratory not infected	-0.6117	0.295	0.0436
clinical infected-laboratory infected	-0.653	0.2656	0.0186
clinical not infected-clinical infected	-0.7886	0.2217	8e-04
laboratory not infected-laboratory infected	-0.8299	0.2063	2e-04

OBSERVATIONS

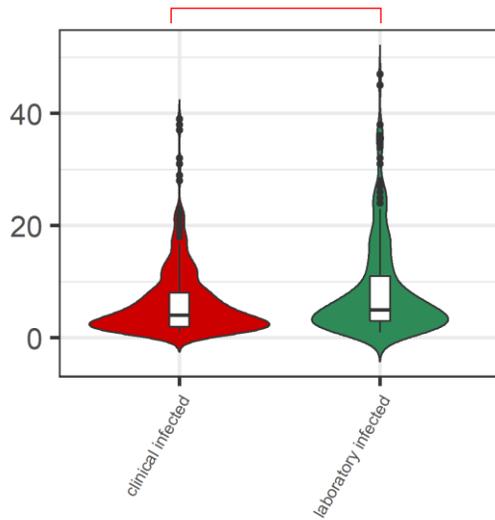
As the infection progresses, laboratory strains are blocking the acidification of the phagolysosomes. In this case, the percentage of macrophages displaying acidification was higher for laboratory strains (40%) compared to causing disease strains (<10%).

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Lab adaptation

M2 – 24 h p.i.

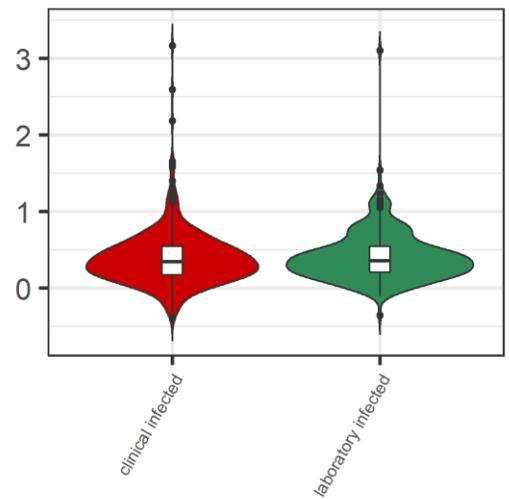
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.4563	0.1233	0
grouplaboratory infected	0.3249	0.1199	0.0086

Contrast	estimate	SE	p.value
clinical infected - laboratory infected	-0.3249	0.1199	0.0086

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	-0.1052	0.0513	0.0405
grouplaboratory infected	0.0934	0.0761	0.2246

Contrast	estimate	SE	p.value
clinical infected - laboratory infected	-0.0934	0.0761	0.2246

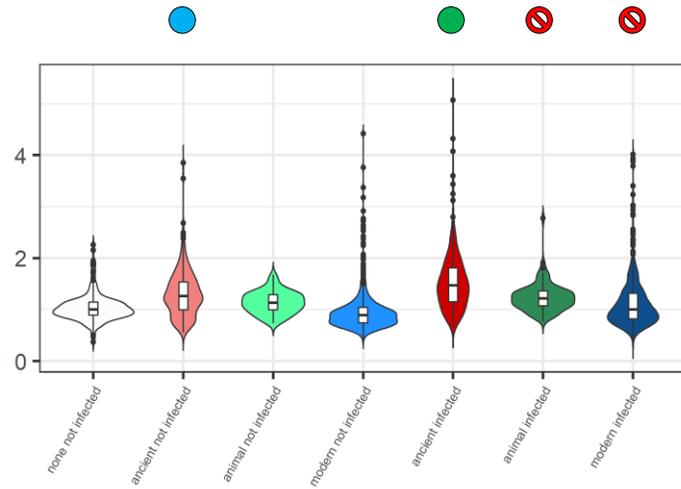
OBSERVATIONS

Laboratory strains showed higher MOI values. Colocalization with acidified compartments was similar among all the groups.

Grouping: Phylogeny

M1 – 6 h p.i.

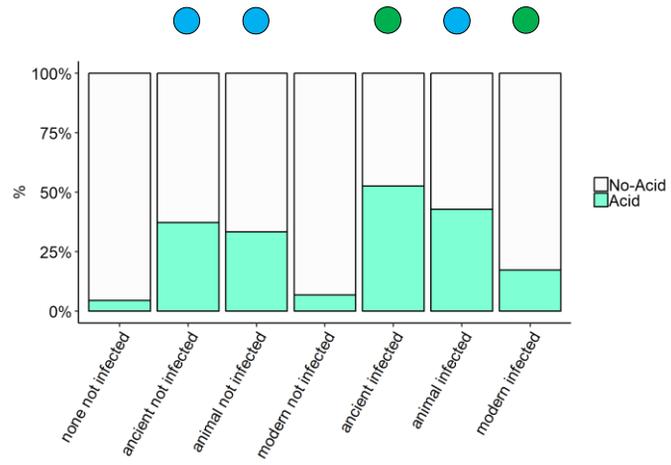
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	0.0032	0.0766	0.9672
groupancient not infected	0.1108	0.0359	0.0066
groupanimal not infected	-0.0011	0.1047	0.9914
groupmodern not infected	-0.0159	0.0513	0.7564
groupancient infected	0.261	0.0591	0
groupanimal infected	0.0641	0.1046	0.54
groupmodern infected	0.0963	0.0514	0.0612

Contrast	estimate	SE	p.value
none not infected-ancient not infected	-0.1108	0.059	0.102
none not infected-animal not infected	0.0011	0.1047	0.9914
none not infected-modern not infected	0.0159	0.0513	0.8824
none not infected-ancient infected	-0.261	0.0591	1e-04
none not infected-animal infected	-0.0641	0.1046	0.7363
none not infected-modern infected	-0.0963	0.0514	0.102
ancient not infected-animal not infected	0.1119	0.0996	0.3921
ancient not infected-modern not infected	0.1267	0.0576	0.0696
animal not infected-modern not infected	0.0148	0.1075	0.9542
ancient infected-animal infected	0.1969	0.0996	0.102
ancient infected-modern infected	0.1647	0.0577	0.0165
animal infected-modern infected	-0.0322	0.1074	0.8824
ancient not infected-ancient infected	-0.1502	0.0143	0
animal not infected-animal infected	-0.0653	0.0266	0.043
modern not infected-modern infected	-0.1122	0.0106	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.5696	0.8413	0.0282	0
groupancient not infected	-1.4974	0.64	4.4702	0.0193
groupanimal not infected	0.6781	0.9758	1.9702	0.4871
groupmodern not infected	0.176	0.6525	1.1924	0.7874
groupancient infected	2.6522	0.6409	14.1849	0
groupanimal infected	1.0873	0.9706	2.9661	0.2626
groupmodern infected	1.87	0.6288	6.4882	0.0029

Contrast	estimate	SE	p.value
none not infected-ancient not infected	-1.4974	0.64	0.055
none not infected-animal not infected	-0.6781	0.9758	0.562
none not infected-modern not infected	-0.176	0.6525	0.7874
none not infected-ancient infected	-2.6522	0.6409	2e-04
none not infected-animal infected	-1.0873	0.9706	0.3939
none not infected-modern infected	-1.87	0.6288	0.011
ancient not infected-animal not infected	0.8193	0.8766	0.4773
ancient not infected-modern not infected	1.3215	0.5769	0.055
animal not infected-modern not infected	0.5022	0.9926	0.6567
ancient infected-animal infected	1.5649	0.8776	0.1598
ancient infected-modern infected	0.7822	0.5522	0.2937
animal infected-modern infected	-0.7827	0.9736	0.5267
ancient not infected-ancient infected	-1.1548	0.2261	0
animal not infected-animal infected	-0.4091	0.3478	0.3939
modern not infected-modern infected	-1.694	0.286	0

OBSERVATIONS

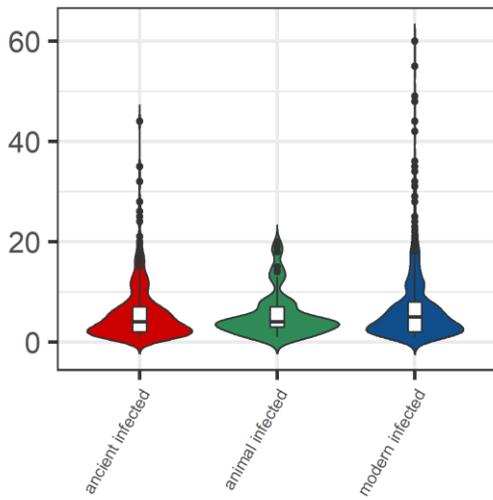
In M1 macrophages, animal and modern lineages are blocking the acidification of phagolysosomes at 6 h p.i. Ancient strains also induced acidification in a higher percentage (50%) of cells compared to the other groups.

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Phylogeny

M1 – 6 h p.i.

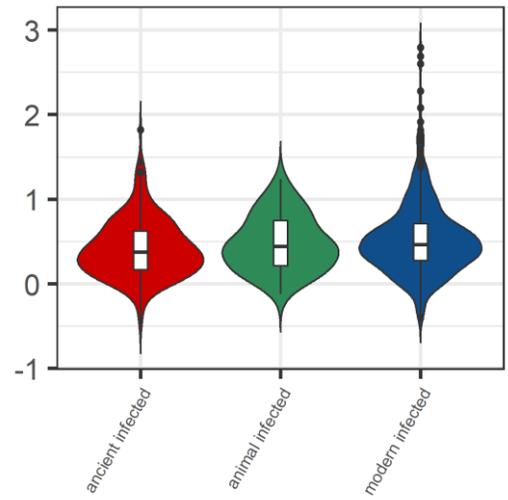
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.4071	0.1117	0
groupanimal infected	-0.0237	0.2083	0.9097
groupmodern infected	0.0615	0.1206	0.6117

Contrast	estimate	SE	p.value
ancient infected - animal infected	0.0237	0.2083	0.9097
ancient infected - modern infected	-0.0615	0.1206	0.9097
animal infected - modern infected	-0.0853	0.222	0.9097

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	-0.0986	0.0866	0.2548
groupanimal infected	0.1444	0.1671	0.3907
groupmodern infected	0.2167	0.0979	0.0306

Contrast	estimate	SE	p.value
ancient infected - animal infected	-0.1444	0.1671	0.5861
ancient infected - modern infected	-0.2167	0.0979	0.0917
animal infected - modern infected	-0.0723	0.1745	0.6801

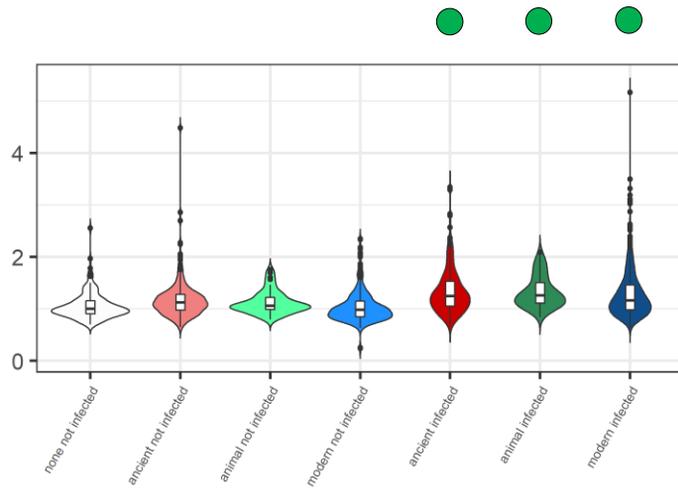
OBSERVATIONS

Colocalization with acidified compartments was similar among all the groups.

Grouping: Phylogeny

M1 – 24 h p.i.

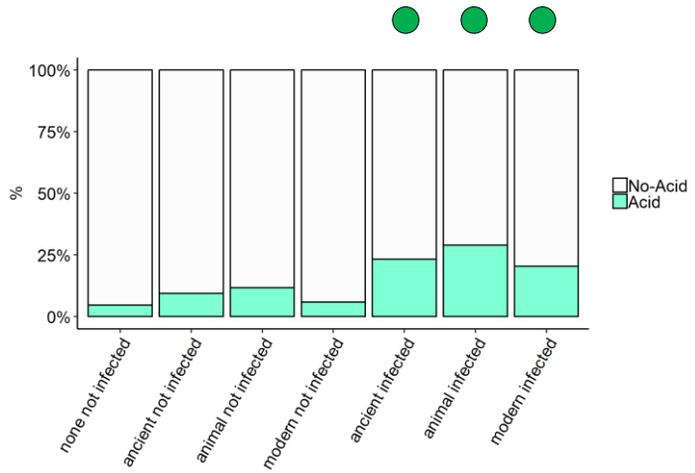
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	0.0087	0.0431	0.8401
groupancient not infected	0.0315	0.0331	0.3422
groupanimal not infected	0.0434	0.0571	0.4478
groupmodern not infected	0.0117	0.0303	0.6992
groupancient infected	0.142	0.0332	0
groupanimal infected	0.2007	0.0548	3e-04
groupmodern infected	0.1799	0.0299	0

Contrast	estimate	SE	p.value
none not infected-ancient not infected	-0.0315	0.0331	0.5703
none not infected-animal not infected	-0.0434	0.0571	0.6718
none not infected-modern not infected	-0.0117	0.0303	0.76
none not infected-ancient infected	-0.142	0.0332	1e-04
none not infected-animal infected	-0.2007	0.0548	6e-04
none not infected-modern infected	-0.1799	0.0299	0
ancient not infected-animal not infected	-0.0119	0.0538	0.8247
ancient not infected-modern not infected	0.0198	0.0313	0.7202
animal not infected-modern not infected	0.0317	0.0582	0.7328
ancient infected-animal infected	-0.0587	0.0515	0.4769
ancient infected-modern infected	-0.0379	0.031	0.4755
animal infected-modern infected	0.0208	0.0557	0.76
ancient not infected-ancient infected	-0.1105	0.0181	0
animal not infected-animal infected	-0.1573	0.036	0
modern not infected-modern infected	-0.1682	0.0146	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.1891	0.4578	0.0412	0
groupancient not infected	0.5115	0.4775	1.6678	0.284
groupanimal not infected	0.8163	0.6713	2.2622	0.224
groupmodern not infected	0.4505	0.4799	1.569	0.3479
groupancient infected	1.6627	0.4509	5.2737	2e-04
groupanimal infected	1.9547	0.569	7.0621	6e-04
groupmodern infected	1.8919	0.4403	6.6322	0

Contrast	estimate	SE	p.value
none not infected-ancient not infected	-0.5115	0.4775	0.5325
none not infected-animal not infected	-0.8163	0.6713	0.4799
none not infected-modern not infected	-0.4505	0.4799	0.5798
none not infected-ancient infected	-1.6627	0.4509	8e-04
none not infected-animal infected	-1.9547	0.569	0.0018
none not infected-modern infected	-1.8919	0.4403	1e-04
ancient not infected-animal not infected	-0.3048	0.5788	0.6905
ancient not infected-modern not infected	0.0611	0.3735	0.8945
animal not infected-modern not infected	0.3659	0.6218	0.6905
ancient infected-animal infected	-0.292	0.4286	0.676
ancient infected-modern infected	-0.2292	0.2814	0.6231
animal infected-modern infected	0.0628	0.4737	0.8945
ancient not infected-ancient infected	-1.1512	0.2683	1e-04
animal not infected-animal infected	-1.1384	0.5062	0.0613
modern not infected-modern infected	-1.4415	0.2592	0

OBSERVATIONS

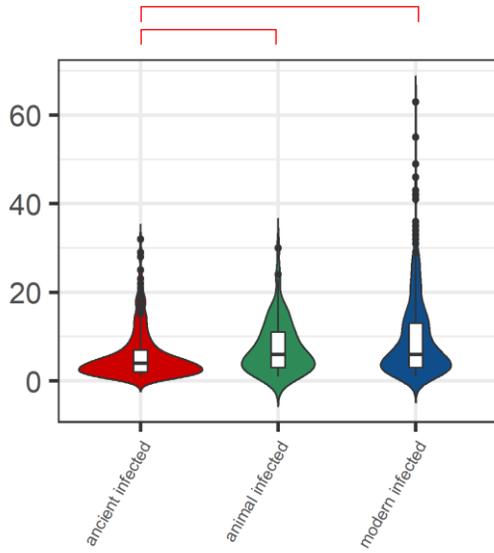
At 24 h p.i. none of the category is blocking the acidification of the phagolysosomes. However, despite an increase in the acidification of the phagolysosomes was observed, the number of macrophages displaying such acidification was found generally low (25%).

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, stastically not significant

Grouping: Phylogeny

M1 – 24 h p.i.

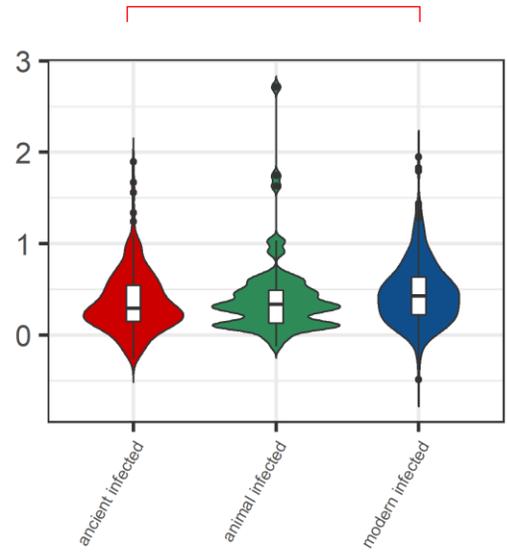
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.4813	0.1278	0
groupanimal infected	0.4699	0.1713	0.008
groupmodern infected	0.2946	0.1056	0.007

Contrast	estimate	SE	p.value
ancient infected - animal infected	-0.4699	0.1713	0.012
ancient infected - modern infected	-0.2946	0.1056	0.012
animal infected - modern infected	0.1752	0.1954	0.3733

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	-0.1795	0.0799	0.0251
groupanimal infected	0.0118	0.1634	0.9427
groupmodern infected	0.2982	0.0973	0.0032

Contrast	estimate	SE	p.value
ancient infected - animal infected	-0.0118	0.1634	0.9427
ancient infected - modern infected	-0.2982	0.0973	0.0097
animal infected - modern infected	-0.2864	0.1623	0.124

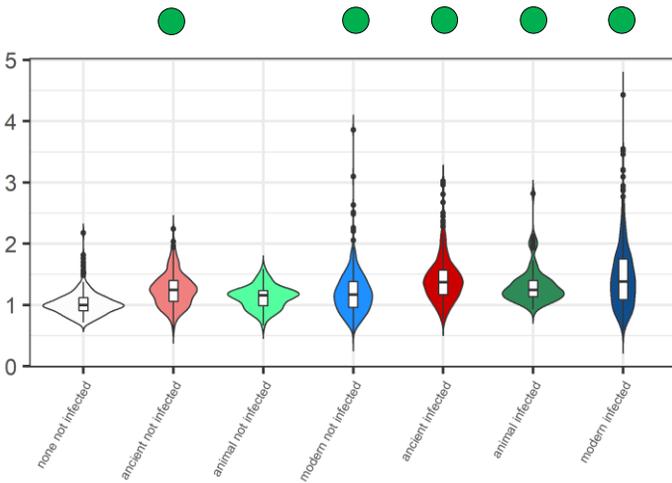
OBSERVATIONS

Colocalization with acidified compartments was found higher for modern lineages strains. Also, ancient lineages showed lower MOI compared to the other categories.

Grouping: Phylogeny

M2 – 6 h p.i.

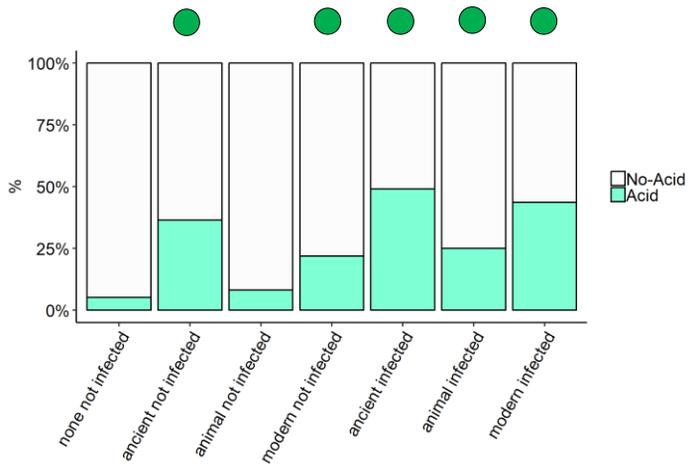
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	-0.0101	0.0525	0.8478
groupancient not infected	0.1614	0.0472	8e-04
groupanimal not infected	0.1094	0.0831	0.1883
groupmodern not infected	0.1363	0.0405	8e-04
groupancient infected	0.2705	0.0473	0
groupanimal infected	0.216	0.083	0.0093
groupmodern infected	0.2824	0.0405	0

Contrast	estimate	SE	p.value
none not infected-ancient not infected	-0.1614	0.0472	0.0016
none not infected-animal not infected	-0.1094	0.0831	0.3138
none not infected-modern not infected	-0.1363	0.0405	0.0017
none not infected-ancient infected	-0.2705	0.0473	0
none not infected-animal infected	-0.216	0.083	0.0174
none not infected-modern infected	-0.2824	0.0405	0
ancient not infected-animal not infected	0.052	0.0792	0.6389
ancient not infected-modern not infected	0.0252	0.0455	0.6697
ancient infected-animal infected	-0.0269	0.0848	0.794
ancient infected-modern infected	0.0544	0.079	0.6389
animal infected-modern infected	-0.0119	0.0456	0.794
ancient not infected-ancient infected	-0.0663	0.0846	0.6389
ancient not infected-animal infected	-0.109	0.0139	0
animal not infected-animal infected	-0.1066	0.0264	2e-04
modern not infected-modern infected	-0.1461	0.01	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.208	0.6462	0.0404	0
groupancient not infected	1.9331	0.572	6.9106	7e-04
groupanimal not infected	0.6619	0.9518	1.9385	0.4868
groupmodern not infected	1.5069	0.5249	4.5128	0.0041
groupancient infected	2.816	0.572	16.7104	0
groupanimal infected	2.0926	0.882	8.1062	0.0177
groupmodern infected	2.978	0.5202	19.6488	0

Contrast	estimate	SE	p.value
none not infected-ancient not infected	-1.9331	0.572	0.0022
none not infected-animal not infected	-0.6619	0.9518	0.5216
none not infected-modern not infected	-1.5069	0.5249	0.0102
none not infected-ancient infected	-2.816	0.572	0
none not infected-animal infected	-2.0926	0.882	0.0331
none not infected-modern infected	-2.978	0.5202	0
ancient not infected-animal not infected	1.2712	0.8495	0.2243
ancient not infected-modern not infected	0.4261	0.4678	0.4181
ancient infected-animal infected	-0.845	0.9196	0.4181
ancient infected-modern infected	0.7234	0.7688	0.4181
animal infected-modern infected	-0.162	0.4623	0.7261
ancient not infected-ancient infected	-0.8854	0.8444	0.4181
ancient not infected-animal infected	-0.883	0.2261	4e-04
animal not infected-animal infected	-1.4307	0.5082	0.0104
modern not infected-modern infected	-1.4711	0.173	0

OBSERVATIONS

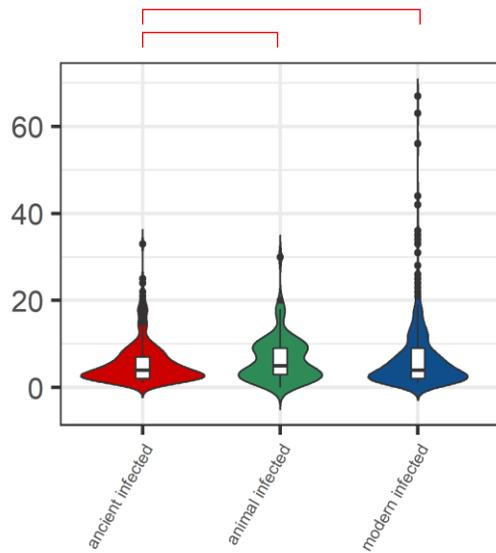
In M2 macrophages, none of the category is blocking the acidification of the phagolysosomes at 6 h p.i. and between 25% (animal) and 50% (ancient, modern) of macrophages showed acidification.

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Phylogeny

M2 – 6 h p.i.

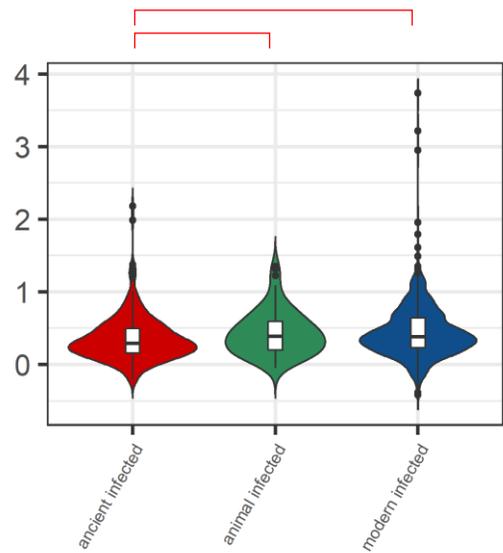
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.4813	0.1278	0
groupanimal infected	0.4699	0.1713	0.008
groupmodern infected	0.2946	0.1056	0.007

Contrast	estimate	SE	p.value
ancient infected - animal infected	-0.4699	0.1713	0.012
ancient infected - modern infected	-0.2946	0.1056	0.012
animal infected - modern infected	0.1752	0.1954	0.3733

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	-0.2813	0.0575	0
groupanimal infected	0.2535	0.1085	0.0227
groupmodern infected	0.2415	0.0643	4e-04

Contrast	estimate	SE	p.value
ancient infected - animal infected	-0.2535	0.1085	0.0341
ancient infected - modern infected	-0.2415	0.0643	0.0012
animal infected - modern infected	0.0119	0.1124	0.9156

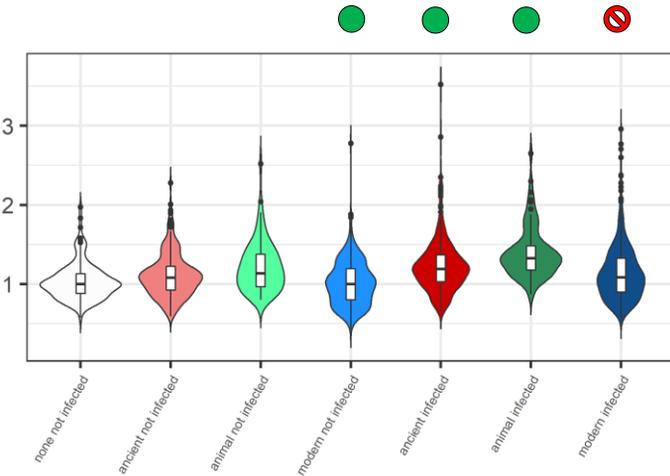
OBSERVATIONS

Ancient lineages showed lower MOI compared to the other categories. Colocalization with acidified compartments was found lower for ancient lineages strains.

Grouping: Phylogeny

M2 – 24 h.p.i.

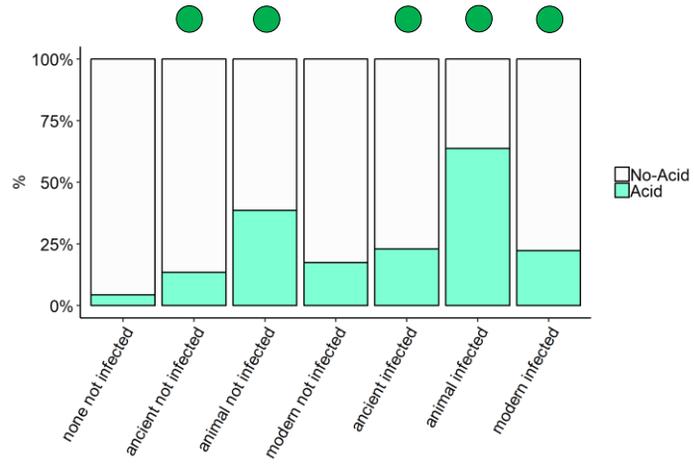
a) Phagolysosomal acidification



Par	Value	Std.Error	p-value
(Intercept)	0.0119	0.053	0.8229
groupancient not infected	0.0204	0.0317	0.519
groupanimal not infected	0.0679	0.0534	0.2033
groumodern not infected	-0.0721	0.0289	0.0126
groupancient infected	0.121	0.0317	1e-04
groupanimal infected	0.2187	0.0528	0
groumodern infected	0.0273	0.0285	0.3386

Contrast	estimate	SE	p-value
none not infected-ancient not infected	-0.0204	0.0317	0.519
none not infected-animal not infected	-0.0679	0.0534	0.2542
none not infected-modern not infected	0.0721	0.0289	0.0189
none not infected-ancient infected	-0.121	0.0317	4e-04
none not infected-animal infected	-0.2187	0.0528	1e-04
none not infected-modern infected	-0.0273	0.0285	0.3651
ancient not infected-animal not infected	-0.0475	0.0499	0.3651
ancient not infected-modern not infected	0.0925	0.0299	0.0038
animal not infected-modern not infected	0.14	0.0545	0.0172
ancient infected-animal infected	-0.0977	0.0492	0.0645
ancient infected-modern infected	0.0937	0.0297	0.0035
animal infected-modern infected	0.1914	0.0538	0.001
ancient not infected-ancient infected	-0.1006	0.0157	0
animal not infected-animal infected	-0.1508	0.0293	0
modern not infected-modern infected	-0.0994	0.0127	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.5319	0.6429	0.0292	0
groupancient not infected	1.3448	0.5089	3.8376	0.0082
groupanimal not infected	1.8666	0.5611	6.4663	9e-04
groumodern not infected	0.9158	0.5047	2.4987	0.0696
groupancient infected	2.0863	0.495	8.0549	0
groupanimal infected	2.8936	0.5554	18.0578	0
groumodern infected	1.6916	0.4911	5.4284	6e-04

Contrast	estimate	SE	p-value
none not infected-ancient not infected	-1.3448	0.5089	0.0137
none not infected-animal not infected	-1.8666	0.5611	0.0026
none not infected-modern not infected	-0.9158	0.5047	0.087
none not infected-ancient infected	-2.0863	0.495	2e-04
none not infected-animal infected	-2.8936	0.5554	0
none not infected-modern infected	-1.6916	0.4911	0.0026
ancient not infected-animal not infected	-0.5218	0.3587	0.1682
ancient not infected-modern not infected	0.4291	0.3725	0.2494
animal not infected-modern not infected	0.9508	0.4641	0.0552
ancient infected-animal infected	-0.8073	0.3397	0.0262
ancient infected-modern infected	0.3946	0.3171	0.2285
animal infected-modern infected	1.2019	0.4349	0.0107
ancient not infected-ancient infected	-0.7414	0.247	0.0058
animal not infected-animal infected	-1.027	0.3393	0.0058
modern not infected-modern infected	-0.7759	0.2306	0.0026

OBSERVATIONS

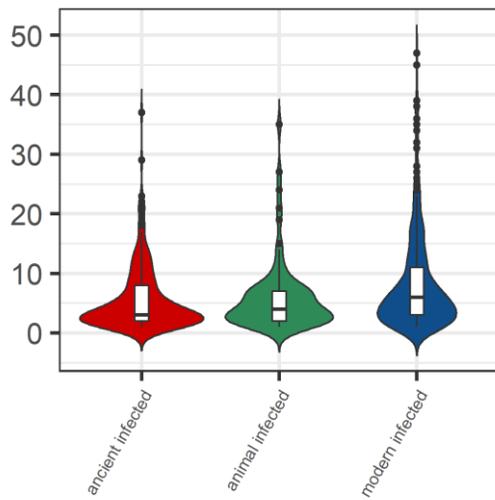
As the infection progresses, modern strains are blocking the acidification of the phagolysosomes. The percentage of acidified cells for these categories dropped below 25%, whereas animal strains induced acidification of phagolysosomes in more than 60% of macrophages.

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Phylogeny

M2 – 24 h p.i.

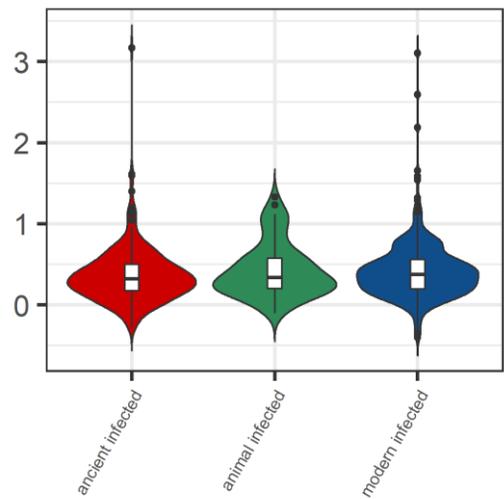
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.4013	0.1241	0
groupanimal infected	0.1285	0.1929	0.508
groupmodern infected	0.2535	0.1182	0.0359

Contrast	estimate	SE	p.value
ancient infected - animal infected	-0.1285	0.1929	0.5652
ancient infected - modern infected	-0.2535	0.1182	0.1076
animal infected - modern infected	-0.1251	0.2163	0.5652

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	-0.1895	0.061	0.002
groupanimal infected	0.2084	0.1193	0.0857
groupmodern infected	0.1631	0.0733	0.0298

Contrast	estimate	SE	p.value
ancient infected - animal infected	-0.2084	0.1193	0.1285
ancient infected - modern infected	-0.1631	0.0733	0.0895
animal infected - modern infected	0.0453	0.1231	0.7143

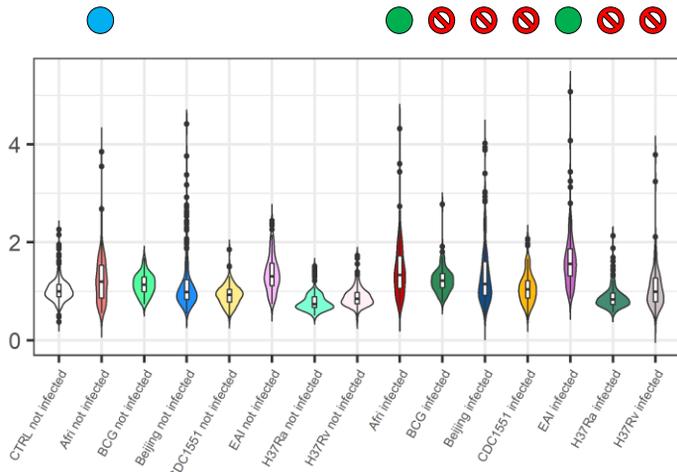
OBSERVATIONS

Differences in the MOI were cleared, even though modern strains showed higher MOI distribution. Colocalization with acidified compartments was similar among all the groups.

Grouping: Sample

M1 – 6 h p.i.

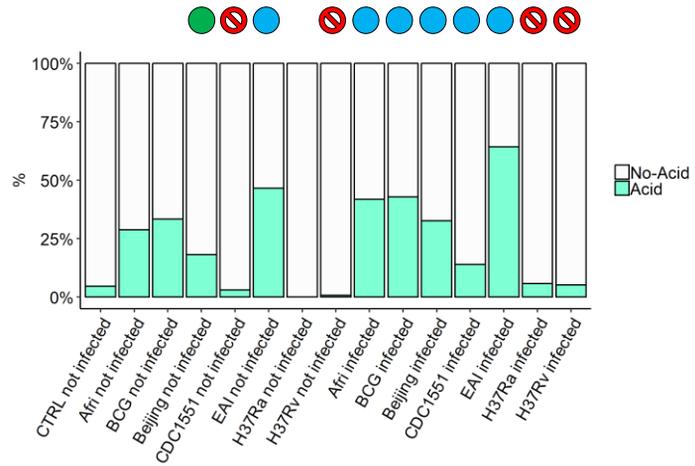
a) Phagolysosomal acidification



Par	Value	Std.Error	p-value
(Intercept)	0.0058	0.0668	0.9308
groupAfr1 not infected	0.1237	0.0695	0.0754
groupBCG not infected	0.0317	0.1052	0.7631
groupBeijing not infected	0.0952	0.0641	0.1379
groupCDC1551 not infected	-0.0414	0.0774	0.5923
groupEAI not infected	0.1613	0.075	0.0315
groupH37Ra not infected	-0.1934	0.0796	0.0152
groupH37Rv not infected	-0.0723	0.0801	0.3668
groupAfr1 infected	0.2767	0.0697	1e-04
groupBCG infected	0.097	0.1051	0.3562
groupBeijing infected	0.2043	0.064	0.0014
groupCDC1551 infected	0.1113	0.0776	0.1516
groupEAI infected	0.3083	0.0752	0
groupH37Ra infected	-0.112	0.08	0.1614
groupH37Rv infected	0.0413	0.0806	0.6083

Contrast	estimate	SE	p-value
none not infected-Afr1 not infected	-0.1295	0.079	0.2473
none not infected-BCG not infected	-0.0375	0.1119	0.7622
none not infected-Beijing not infected	-0.101	0.0743	0.3244
none not infected-CDC1551 not infected	0.0356	0.086	0.7407
none not infected-EAI not infected	-0.1671	0.084	0.1734
none not infected-H37Ra not infected	0.1876	0.0873	0.1516
none not infected-H37Rv not infected	0.0665	0.0877	0.5734
none not infected-Afr1 infected	-0.2825	0.0791	0.0403
none not infected-BCG infected	-0.1028	0.1118	0.5209
none not infected-Beijing infected	-0.2101	0.0742	0.0799
none not infected-CDC1551 infected	-0.1171	0.0862	0.3244
none not infected-EAI infected	-0.3141	0.0842	0.0356
none not infected-H37Ra infected	0.1062	0.0877	0.3793
none not infected-H37Rv infected	-0.0471	0.0882	0.6986
Afr1 not infected-BCG not infected	0.092	0.1167	0.5427
Afr1 not infected-Beijing not infected	0.0285	0.0724	0.7407
Afr1 not infected-CDC1551 not infected	0.1651	0.0858	0.1334
Afr1 not infected-EAI not infected	-0.0376	0.0853	0.7315
Afr1 not infected-H37Ra not infected	0.317	0.0932	0.0039
Afr1 not infected-H37Rv not infected	0.196	0.096	0.1135
BCG not infected-Beijing not infected	-0.0635	0.1143	0.6752
BCG not infected-CDC1551 not infected	0.0732	0.1243	0.6612
BCG not infected-EAI not infected	-0.1296	0.0989	0.3074
BCG not infected-H37Ra not infected	0.2251	0.1265	0.1585
BCG not infected-H37Rv not infected	0.104	0.127	0.5422
Beijing not infected-CDC1551 not infected	0.1366	0.0802	0.1734
Beijing not infected-EAI not infected	-0.0662	0.0827	0.5427
Beijing not infected-H37Ra not infected	0.2885	0.0891	0.0064
Beijing not infected-H37Rv not infected	0.1674	0.0895	0.1436
CDC1551 not infected-EAI not infected	-0.2028	0.098	0.1106
CDC1551 not infected-H37Ra not infected	0.1519	0.0968	0.2102
CDC1551 not infected-H37Rv not infected	0.0308	0.0968	0.7622
EAI not infected-H37Ra not infected	0.3547	0.1017	0.0035
EAI not infected-H37Rv not infected	0.2336	0.1025	0.0754
H37Ra not infected-H37Rv not infected	-0.1211	0.0898	0.2947
Afr1 infected-BCG infected	0.1798	0.1167	0.2162
Afr1 infected-Beijing infected	0.0725	0.0725	0.4445
Afr1 infected-CDC1551 infected	0.1654	0.0862	0.1334
Afr1 infected-EAI infected	-0.0316	0.0856	0.7478
Afr1 infected-H37Ra infected	0.3888	0.0936	4e-04
Afr1 infected-H37Rv infected	0.2354	0.0966	0.0551
BCG infected-Beijing infected	-0.1073	0.1141	0.4756
BCG infected-CDC1551 infected	-0.0143	0.1244	0.9083
BCG infected-EAI infected	-0.2113	0.099	0.0987
BCG infected-H37Ra infected	0.209	0.1267	0.1837
BCG infected-H37Rv infected	0.0557	0.1273	0.7315
Beijing infected-CDC1551 infected	0.093	0.0803	0.3621
Beijing infected-EAI infected	-0.1041	0.0827	0.3244

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.505	0.7631	0.03	0
groupAfr1 not infected	1.8572	0.7306	6.4057	0.011
groupBCG not infected	0.8416	0.9631	2.32	0.3822
groupBeijing not infected	1.1595	0.7357	3.1882	0.115
groupCDC1551 not infected	0.1986	0.9662	1.2197	0.8371
groupEAI not infected	1.7238	0.7466	5.6058	0.021
groupH37Ra not infected	-14.8469	583.2223	0	0.9797
groupH37Rv not infected	-1.2751	1.3749	0.2794	0.3537
groupAfr1 infected	2.998	0.7254	20.046	0
groupBCG infected	1.25	0.9578	3.4903	0.1919
groupBeijing infected	2.6757	0.7166	14.5219	2e-04
groupCDC1551 infected	1.9659	0.8611	7.1411	0.0224
groupEAI infected	2.8677	0.7499	17.5963	1e-04
groupH37Ra infected	0.8593	0.9051	2.3614	0.3424
groupH37Rv infected	0.723	1.0227	2.0606	0.4796

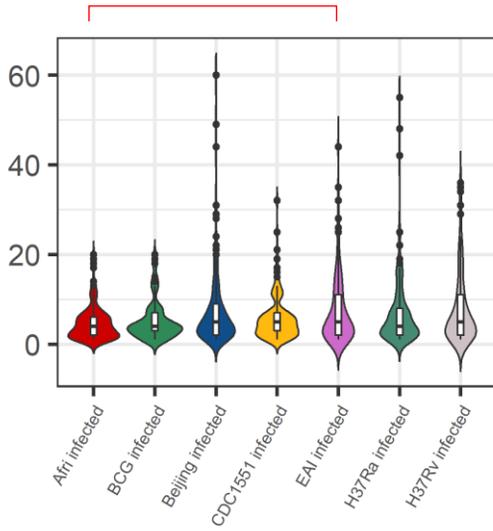
Contrast	estimate	SE	p-value
none not infected-Afr1 not infected	1.6478	0.7801	0.1456
none not infected-BCG not infected	2.6634	1.0411	0.0663
none not infected-Beijing not infected	2.3455	0.777	0.0239
none not infected-CDC1551 not infected	3.3063	0.9888	0.0104
none not infected-EAI not infected	1.7812	0.8266	0.1403
none not infected-H37Ra not infected	18.3519	583.2224	0.9814
none not infected-H37Rv not infected	4.7801	1.3959	0.0097
none not infected-Afr1 infected	0.5069	0.7747	0.718
none not infected-BCG infected	2.255	1.0362	0.1403
none not infected-Beijing infected	0.8293	0.7559	0.5029
none not infected-CDC1551 infected	1.5391	0.8854	0.2071
none not infected-EAI infected	0.6373	0.828	0.6468
none not infected-H37Ra infected	2.6457	0.9386	0.038
none not infected-H37Rv infected	2.782	1.0503	0.0566
Afr1 not infected-BCG not infected	1.0156	1.035	0.5274
Afr1 not infected-Beijing not infected	0.6977	0.6778	0.5029
Afr1 not infected-CDC1551 not infected	1.6586	0.9211	0.1917
Afr1 not infected-EAI not infected	0.1334	0.7717	0.9814
Afr1 not infected-H37Ra not infected	16.7041	583.2224	0.9814
Afr1 not infected-H37Rv not infected	3.1323	1.4441	0.1403
BCG not infected-Beijing not infected	-0.3179	1.0413	0.939
BCG not infected-CDC1551 not infected	0.6429	1.2377	0.8089
BCG not infected-EAI not infected	-0.8822	0.837	0.5029
BCG not infected-H37Ra not infected	15.6885	583.2229	0.9814
BCG not infected-H37Rv not infected	2.1167	1.6078	0.3948
Beijing not infected-CDC1551 not infected	0.9608	0.9201	0.5029
Beijing not infected-EAI not infected	-0.5643	0.7826	0.6742
Beijing not infected-H37Ra not infected	16.0064	583.2224	0.9814
Beijing not infected-H37Rv not infected	2.4346	1.4883	0.2112
CDC1551 not infected-EAI not infected	-1.5252	1.043	0.3232
CDC1551 not infected-H37Ra not infected	15.0455	583.2227	0.9814
CDC1551 not infected-H37Rv not infected	1.4737	1.5552	0.5275
EAI not infected-H37Ra not infected	16.5707	583.2225	0.9814
EAI not infected-H37Rv not infected	2.9989	1.4773	0.157
H37Ra not infected-H37Rv not infected	-13.5718	583.2232	0.9814
Afr1 infected-BCG infected	1.748	1.0262	0.2112
Afr1 infected-Beijing infected	0.3224	0.6516	0.8147
Afr1 infected-CDC1551 infected	1.0322	0.8074	0.4087
Afr1 infected-EAI infected	0.1303	0.7644	0.9814
Afr1 infected-H37Ra infected	2.1388	0.9561	0.1403
Afr1 infected-H37Rv infected	2.275	1.109	0.157
BCG infected-Beijing infected	-1.4257	1.0247	0.3566
BCG infected-CDC1551 infected	-0.7159	1.1531	0.7323
BCG infected-EAI infected	-1.6177	0.8452	0.1752
BCG infected-H37Ra infected	0.3907	1.2151	0.939
BCG infected-H37Rv infected	0.527	1.3151	0.8854
Beijing infected-CDC1551 infected	0.7098	0.7852	0.549
Beijing infected-EAI infected	-0.192	0.767	0.972

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Sample

M1 – 6 h p.i.

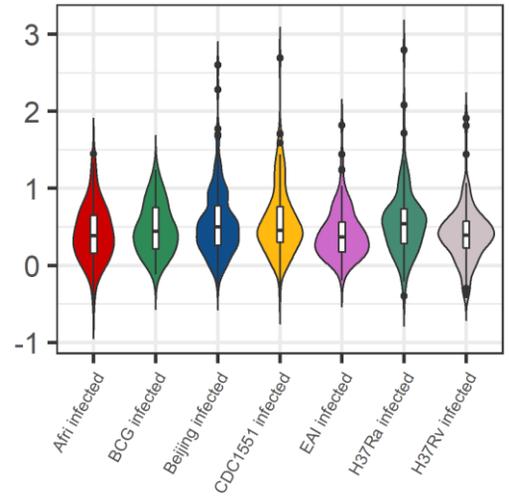
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.1618	0.1532	0
groupBCG infected	0.5123	0.2551	0.0493
groupBeijing infected	0.2666	0.146	0.0729
groupCDC1551 infected	0.0771	0.1751	0.6612
groupEAI infected	0.6082	0.183	0.0015
groupH37Ra infected	0.3031	0.1976	0.1306
groupH37Rv infected	0.2976	0.2059	0.1539

Contrast	estimate	SE	p.value
Afri infected - BCG infected	-0.5123	0.2551	0.3061
Afri infected - Beijing infected	-0.2666	0.146	0.3061
Afri infected - CDC1551 infected	-0.0771	0.1751	0.7713
Afri infected - EAI infected	-0.6082	0.183	0.0324
Afri infected - H37Ra infected	-0.3031	0.1976	0.3867
Afri infected - H37Rv infected	-0.2976	0.2059	0.3867
BCG infected - Beijing infected	0.2457	0.2511	0.498
BCG infected - CDC1551 infected	0.4352	0.2743	0.3867
BCG infected - EAI infected	-0.0959	0.2033	0.7713
BCG infected - H37Ra infected	0.2092	0.2835	0.6083
BCG infected - H37Rv infected	0.2148	0.2855	0.6083
Beijing infected - CDC1551 infected	0.1895	0.1614	0.4502
Beijing infected - EAI infected	-0.3416	0.1787	0.3061
Beijing infected - H37Ra infected	-0.0364	0.1881	0.9146
Beijing infected - H37Rv infected	-0.0309	0.1897	0.9146
CDC1551 infected - EAI infected	-0.5311	0.2133	0.1645
CDC1551 infected - H37Ra infected	-0.226	0.2009	0.4502
CDC1551 infected - H37Rv infected	-0.2204	0.2016	0.4502
EAI infected - H37Ra infected	0.3051	0.227	0.3867
EAI infected - H37Rv infected	0.3107	0.2299	0.3867
H37Ra infected - H37Rv infected	0.0055	0.1847	0.9763

d) Colocalization Index



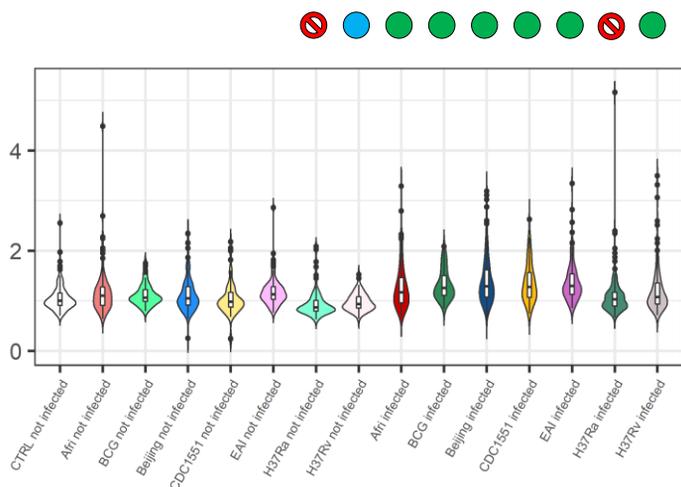
Par	Value	Std.Error	p.value
(Intercept)	-0.009	0.1222	0.9412
groupBCG infected	0.0397	0.2109	0.8513
groupBeijing infected	0.2322	0.1282	0.0752
groupCDC1551 infected	0.1699	0.1498	0.2613
groupEAI infected	-0.0855	0.1555	0.5845
groupH37Ra infected	0.2281	0.1663	0.1756
groupH37Rv infected	-0.2072	0.1721	0.2335

Contrast	estimate	SE	p.value
Afri infected - BCG infected	-0.0397	0.2109	0.8939
Afri infected - Beijing infected	-0.2322	0.1282	0.3157
Afri infected - CDC1551 infected	-0.1699	0.1498	0.5486
Afri infected - EAI infected	0.0855	0.1555	0.722
Afri infected - H37Ra infected	-0.2281	0.1663	0.4609
Afri infected - H37Rv infected	0.2072	0.1721	0.5449
BCG infected - Beijing infected	-0.1925	0.2068	0.6223
BCG infected - CDC1551 infected	-0.1302	0.2242	0.722
BCG infected - EAI infected	0.1252	0.173	0.7081
BCG infected - H37Ra infected	-0.1883	0.231	0.6756
BCG infected - H37Rv infected	0.2469	0.2313	0.5539
Beijing infected - CDC1551 infected	0.0623	0.1381	0.7623
Beijing infected - EAI infected	0.3177	0.151	0.2084
Beijing infected - H37Ra infected	0.0042	0.1592	0.9791
Beijing infected - H37Rv infected	0.0623	0.1381	0.7623
CDC1551 infected - EAI infected	0.4395	0.1583	0.0962
CDC1551 infected - H37Ra infected	0.2554	0.1773	0.4609
CDC1551 infected - H37Rv infected	-0.0581	0.1698	0.8106
EAI infected - H37Ra infected	0.3771	0.1691	0.2076
EAI infected - H37Rv infected	-0.3135	0.1874	0.349
H37Ra infected - H37Rv infected	0.4353	0.1614	0.0962

Grouping: Sample

M1 – 24 h.p.i.

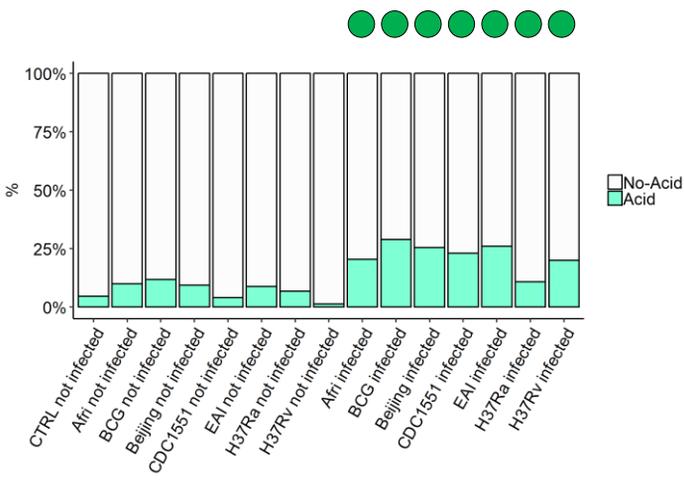
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	0.0184	0.0353	0.6021
groupAfrI not infected	0.0221	0.0389	0.5709
groupBCG not infected	0.0757	0.0572	0.1858
groupBeijing not infected	0.0485	0.0368	0.1871
groupCDC1551 not infected	0.0033	0.0444	0.9402
groupEAI not infected	0.0742	0.0406	0.0675
groupH37Ra not infected	-0.0957	0.0466	0.0403
groupH37Rv not infected	-0.0249	0.0462	0.5902
groupAfrI infected	0.1127	0.0389	0.0038
groupBCG infected	0.2329	0.0548	0
groupBeijing infected	0.2026	0.0358	0
groupCDC1551 infected	0.2551	0.0435	0
groupEAI infected	0.1992	0.0408	0
groupH37Ra infected	0.0394	0.0448	0.3793
groupH37Rv infected	0.1119	0.045	0.0129

Contrast	estimate	SE	p.value
none not infected-AfrI not infected	-0.0405	0.0408	0.4642
none not infected-BCG not infected	-0.0941	0.0591	0.2506
none not infected-Beijing not infected	-0.067	0.0387	0.2413
none not infected-CDC1551 not infected	-0.0218	0.0459	0.7302
none not infected-EAI not infected	-0.0927	0.0429	0.1486
none not infected-H37Ra not infected	0.0773	0.047	0.2436
none not infected-H37Rv not infected	0.0065	0.0467	0.9384
none not infected-AfrI infected	-0.1312	0.0407	0.0438
none not infected-BCG infected	-0.2514	0.0568	0.0128
none not infected-Beijing infected	-0.2211	0.0378	0.0039
none not infected-CDC1551 infected	-0.2735	0.045	0.0039
none not infected-EAI infected	-0.2176	0.0431	0.0077
none not infected-H37Ra infected	-0.0578	0.0452	0.354
none not infected-H37Rv infected	-0.1303	0.0454	0.0631
AfrI not infected-BCG not infected	-0.0536	0.0626	0.4942
AfrI not infected-Beijing not infected	-0.0265	0.0399	0.6188
AfrI not infected-CDC1551 not infected	0.0187	0.0479	0.7557
AfrI not infected-EAI not infected	-0.0522	0.0453	0.3578
AfrI not infected-H37Ra not infected	0.1178	0.052	0.0631
AfrI not infected-H37Rv not infected	0.047	0.0529	0.4823
BCG not infected-Beijing not infected	0.0271	0.0618	0.7302
BCG not infected-CDC1551 not infected	0.0723	0.0677	0.3947
BCG not infected-EAI not infected	0.0014	0.0535	0.9868
BCG not infected-H37Ra not infected	0.1714	0.0694	0.0431
BCG not infected-H37Rv not infected	0.1006	0.0693	0.2436
Beijing not infected-CDC1551 not infected	0.0452	0.0453	0.4275
Beijing not infected-EAI not infected	-0.0257	0.0445	0.6703
Beijing not infected-H37Ra not infected	0.1443	0.0502	0.0144
Beijing not infected-H37Rv not infected	0.0734	0.0501	0.2436
CDC1551 not infected-EAI not infected	-0.0709	0.0532	0.2814
CDC1551 not infected-H37Ra not infected	0.099	0.0548	0.1486
CDC1551 not infected-H37Rv not infected	0.0282	0.0544	0.6959
EAI not infected-H37Ra not infected	0.17	0.0558	0.0114
EAI not infected-H37Rv not infected	0.0991	0.0557	0.1535
H37Ra not infected-H37Rv not infected	-0.0708	0.0509	0.2588
AfrI infected-BCG infected	-0.1202	0.0605	0.1186
AfrI infected-Beijing infected	-0.0899	0.0392	0.0625
AfrI infected-CDC1551 infected	-0.1424	0.0471	0.0115
AfrI infected-EAI infected	-0.0864	0.0455	0.1375
AfrI infected-H37Ra infected	0.0733	0.0504	0.2436
AfrI infected-H37Rv infected	9e-04	0.0518	0.9868
BCG infected-Beijing infected	0.0303	0.059	0.6959
BCG infected-CDC1551 infected	-0.0222	0.0651	0.7835
BCG infected-EAI infected	0.0338	0.0513	0.6188
BCG infected-H37Ra infected	0.1935	0.0663	0.0132
BCG infected-H37Rv infected	0.1211	0.0665	0.1486
Beijing infected-CDC1551 infected	-0.0525	0.0438	0.3465
Beijing infected-EAI infected	0.0035	0.0438	0.9676

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.1573	0.4472	0.0425	0.007
groupAfrI not infected	0.6976	0.5452	2.0088	0.2007
groupBCG not infected	0.8584	0.683	2.3594	0.2088
groupBeijing not infected	0.6945	0.5348	2.0028	0.1941
groupCDC1551 not infected	0.0581	0.7516	1.0598	0.9384
groupEAI not infected	0.4412	0.5449	1.5546	0.4181
groupH37Ra not infected	0.9405	0.6736	2.5612	0.1626
groupH37Rv not infected	-1.0996	1.115	0.333	0.324
groupAfrI infected	1.6261	0.5024	5.0838	0.0012
groupBCG infected	1.9978	0.5825	7.3727	6e-04
groupBeijing infected	1.9321	0.4719	6.9042	0
groupCDC1551 infected	1.9848	0.5336	7.2773	2e-04
groupEAI infected	1.7757	0.496	5.9046	3e-04
groupH37Ra infected	1.422	0.5799	4.1452	0.0142
groupH37Rv infected	1.855	0.5356	6.3916	5e-04

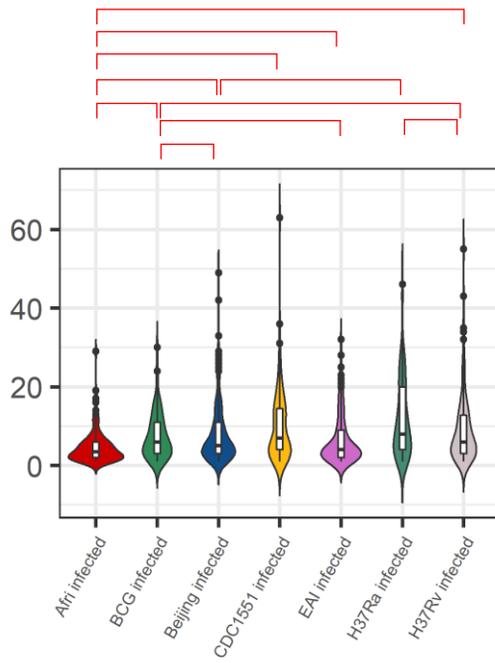
Contrast	estimate	SE	p.value
none not infected-AfrI not infected	2.4597	0.4095	0
none not infected-BCG not infected	2.2988	0.5951	6e-04
none not infected-Beijing not infected	2.4627	0.3939	0
none not infected-CDC1551 not infected	3.0991	0.6548	0
none not infected-EAI not infected	2.716	0.4226	0
none not infected-H37Ra not infected	2.2168	0.5547	5e-04
none not infected-H37Rv not infected	4.2569	1.0504	5e-04
none not infected-AfrI infected	1.5312	0.349	1e-04
none not infected-BCG infected	1.1595	0.4765	0.0524
none not infected-Beijing infected	1.2251	0.3016	5e-04
none not infected-CDC1551 infected	1.1725	0.3853	0.0098
none not infected-EAI infected	1.3815	0.3567	6e-04
none not infected-H37Ra infected	1.7353	0.4357	5e-04
none not infected-H37Rv infected	1.3023	0.3821	0.003
AfrI not infected-BCG not infected	-0.1609	0.6796	0.9145
AfrI not infected-Beijing not infected	0.003	0.4857	0.995
AfrI not infected-CDC1551 not infected	0.6394	0.7259	0.6934
AfrI not infected-EAI not infected	0.2563	0.5149	0.8655
AfrI not infected-H37Ra not infected	-0.2429	0.6584	0.8697
AfrI not infected-H37Rv not infected	1.7972	1.1098	0.2766
BCG not infected-Beijing not infected	0.1639	0.6755	0.9145
BCG not infected-CDC1551 not infected	0.8003	0.8647	0.6934
BCG not infected-EAI not infected	-0.0172	0.6115	0.9747
BCG not infected-H37Ra not infected	4.082	0.7989	0.9483
BCG not infected-H37Rv not infected	1.958	1.1967	0.2766
Beijing not infected-CDC1551 not infected	0.6364	0.71	0.6934
Beijing not infected-EAI not infected	0.2533	0.5127	0.8655
Beijing not infected-H37Ra not infected	-0.2459	0.6453	0.8697
Beijing not infected-H37Rv not infected	1.7942	1.0982	0.2766
CDC1551 not infected-EAI not infected	-0.3831	0.7501	0.8655
CDC1551 not infected-H37Ra not infected	-0.8823	0.8231	0.6585
CDC1551 not infected-H37Rv not infected	1.1577	1.2072	0.6934
EAI not infected-H37Ra not infected	-0.4992	0.6765	0.7635
EAI not infected-H37Rv not infected	1.5408	1.1186	0.4243
H37Ra not infected-H37Rv not infected	2.0401	1.1464	0.2255
AfrI infected-BCG infected	-0.3717	0.5393	0.7797
AfrI infected-Beijing infected	-0.3061	0.3608	0.6934
AfrI infected-CDC1551 infected	-0.3587	0.4494	0.7232
AfrI infected-EAI infected	-0.1497	0.4142	0.8697
AfrI infected-H37Ra infected	0.2041	0.5203	0.8697
AfrI infected-H37Rv infected	-0.2289	0.478	0.8655
BCG infected-Beijing infected	0.0657	0.5166	0.9438
BCG infected-CDC1551 infected	0.013	0.5839	0.995
BCG infected-EAI infected	0.2221	0.4403	0.8655
BCG infected-H37Ra infected	0.5758	0.6279	0.6934
BCG infected-H37Rv infected	0.1428	0.59	0.9145
Beijing infected-CDC1551 infected	-0.0526	0.397	0.9438
Beijing infected-EAI infected	0.1564	0.3882	0.8697

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Sample

M1 – 24 h p.i.

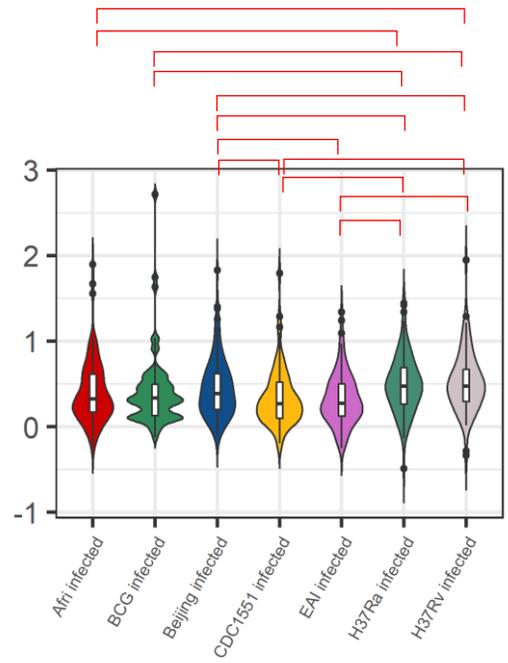
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.3459	0.1565	0
groupBCG infected	0.8519	0.2039	1e-04
groupBeijing infected	0.3098	0.1109	0.0071
groupCDC1551 infected	0.367	0.1412	0.0119
groupEAI infected	0.3906	0.1448	0.0091
groupH37Ra infected	0.7005	0.162	1e-04
groupH37Rv infected	0.1793	0.1664	0.2859

Contrast	estimate	SE	p.value
Afri infected - BCG infected	-0.8519	0.2039	0.0011
Afri infected - Beijing infected	-0.3098	0.1109	0.0247
Afri infected - CDC1551 infected	-0.367	0.1412	0.0256
Afri infected - EAI infected	-0.3906	0.1448	0.0249
Afri infected - H37Ra infected	-0.7005	0.162	0.0011
Afri infected - H37Rv infected	-0.1793	0.1664	0.3753
BCG infected - Beijing infected	0.5421	0.202	0.0249
BCG infected - CDC1551 infected	0.4849	0.2259	0.0688
BCG infected - EAI infected	0.4613	0.1516	0.0185
BCG infected - H37Ra infected	0.1514	0.2369	0.6128
BCG infected - H37Rv infected	0.6726	0.2381	0.0247
Beijing infected - CDC1551 infected	-0.0572	0.1262	0.6847
Beijing infected - EAI infected	-0.0808	0.1427	0.6337
Beijing infected - H37Ra infected	-0.3907	0.151	0.0256
Beijing infected - H37Rv infected	0.1305	0.1506	0.4812
CDC1551 infected - EAI infected	-0.0236	0.1761	0.8938
CDC1551 infected - H37Ra infected	-0.3335	0.1598	0.0723
CDC1551 infected - H37Rv infected	0.1877	0.1579	0.3591
EAI infected - H37Ra infected	-0.3099	0.1911	0.1781
EAI infected - H37Rv infected	0.2113	0.1926	0.3753
H37Ra infected - H37Rv infected	0.5212	0.1322	0.0015

d) Colocalization Index



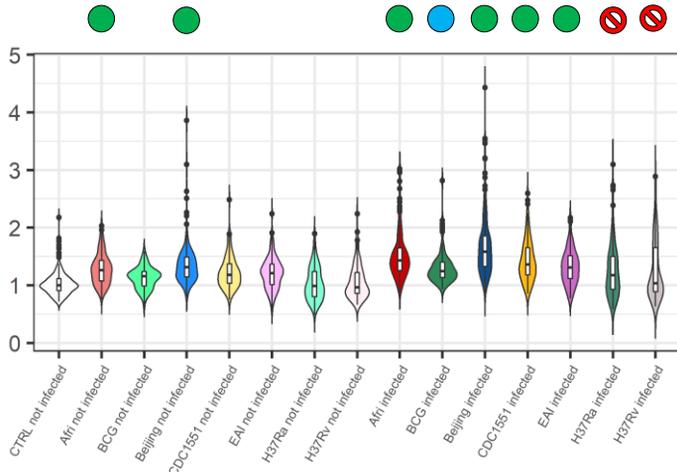
Par	Value	Std.Error	p.value
(Intercept)	-0.1611	0.1235	0.1926
groupBCG infected	-0.1594	0.1916	0.409
groupBeijing infected	0.131	0.1114	0.2443
groupCDC1551 infected	-0.1753	0.1453	0.2326
groupEAI infected	-0.2812	0.1399	0.0492
groupH37Ra infected	0.629	0.1562	2e-04
groupH37Rv infected	0.6642	0.1575	1e-04

Contrast	estimate	SE	p.value
Afri infected - BCG infected	0.1594	0.1916	0.482
Afri infected - Beijing infected	-0.131	0.1114	0.3206
Afri infected - CDC1551 infected	0.1753	0.1453	0.3206
Afri infected - EAI infected	0.2812	0.1399	0.0795
Afri infected - H37Ra infected	-0.629	0.1562	6e-04
Afri infected - H37Rv infected	-0.6642	0.1575	4e-04
BCG infected - Beijing infected	-0.2904	0.1889	0.1944
BCG infected - CDC1551 infected	0.0159	0.2133	0.9408
BCG infected - EAI infected	0.1218	0.1478	0.482
BCG infected - H37Ra infected	-0.7884	0.2173	0.0014
BCG infected - H37Rv infected	-0.8236	0.2164	0.001
Beijing infected - CDC1551 infected	0.3063	0.1345	0.0463
Beijing infected - EAI infected	0.4123	0.1368	0.0073
Beijing infected - H37Ra infected	-0.498	0.1486	0.003
Beijing infected - H37Rv infected	-0.5332	0.1463	0.0014
CDC1551 infected - EAI infected	0.1059	0.1717	0.5967
CDC1551 infected - H37Ra infected	-0.8043	0.1586	0
CDC1551 infected - H37Rv infected	-0.8395	0.1494	0
EAI infected - H37Ra infected	-0.9102	0.1782	0
EAI infected - H37Rv infected	-0.9454	0.1772	0
H37Ra infected - H37Rv infected	-0.0352	0.1294	0.8259

Grouping: Sample

M2 – 6 h p.i.

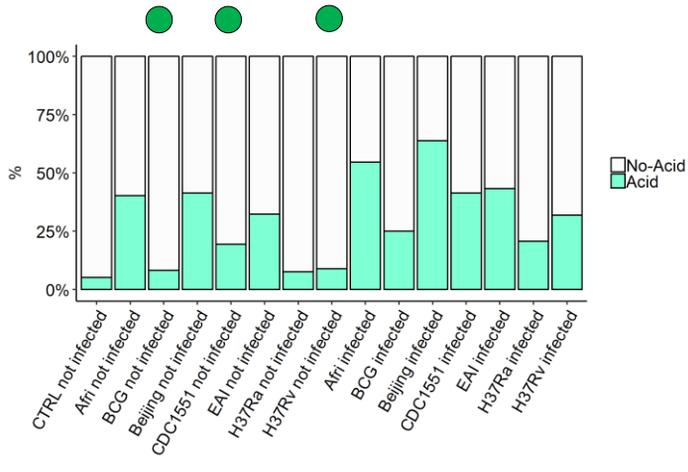
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	-0.0102	0.0427	0.8114
groupAfri not infected	0.2373	0.0532	0
groupBCG not infected	0.0963	0.0789	0.2224
groupBeijing not infected	0.2672	0.0491	0
groupCDC1551 not infected	0.1257	0.0587	0.0325
groupEAI not infected	0.1354	0.0569	0.0174
groupH37Ra not infected	0.0167	0.0593	0.7779
groupH37Rv not infected	0.0451	0.0599	0.4519
groupAfri infected	0.3544	0.0535	0
groupBCG infected	0.1968	0.0788	0.0125
groupBeijing infected	0.4182	0.0489	0
groupCDC1551 infected	0.2551	0.059	0
groupEAI infected	0.242	0.0569	0
groupH37Ra infected	0.178	0.0597	0.0029
groupH37Rv infected	0.1709	0.06	0.0045

Contrast	estimate	SE	p.value
none not infected-Afri not infected	-0.2271	0.0533	0.0147
none not infected-BCG not infected	-0.0861	0.079	0.4273
none not infected-Beijing not infected	-0.257	0.0493	0.0055
none not infected-CDC1551 not infected	-0.1155	0.059	0.1639
none not infected-EAI not infected	-0.1252	0.0569	0.1297
none not infected-H37Ra not infected	-0.0065	0.0597	0.9175
none not infected-H37Rv not infected	-0.0349	0.0603	0.6881
none not infected-Afri infected	0.3442	0.0536	0.0019
none not infected-BCG infected	-0.1866	0.0789	0.1048
none not infected-Beijing infected	-0.408	0.049	6e-04
none not infected-CDC1551 infected	-0.245	0.0592	0.0158
none not infected-EAI infected	0.2319	0.057	0.0158
none not infected-H37Ra infected	-0.1679	0.0601	0.0651
none not infected-H37Rv infected	-0.1607	0.0605	0.076
Afri not infected-BCG not infected	0.141	0.0873	0.1867
Afri not infected-Beijing not infected	-0.0299	0.0555	0.6881
Afri not infected-CDC1551 not infected	0.1116	0.0653	0.1621
Afri not infected-EAI not infected	0.1019	0.0646	0.1906
Afri not infected-H37Ra not infected	0.2205	0.0696	0.0066
Afri not infected-H37Rv not infected	0.1922	0.0719	0.0232
BCG not infected-Beijing not infected	-0.1709	0.0854	0.0988
BCG not infected-CDC1551 not infected	-0.0294	0.0926	0.8301
BCG not infected-EAI not infected	-0.0391	0.0754	0.6919
BCG not infected-H37Ra not infected	0.0796	0.0936	0.5107
BCG not infected-H37Rv not infected	0.0512	0.0941	0.6881
Beijing not infected-CDC1551 not infected	0.1415	0.0611	0.0522
Beijing not infected-EAI not infected	0.1318	0.0624	0.0784
Beijing not infected-H37Ra not infected	0.2504	0.0666	0.0012
Beijing not infected-H37Rv not infected	0.2221	0.0672	0.0047
CDC1551 not infected-EAI not infected	-0.0097	0.0734	0.9175
CDC1551 not infected-H37Ra not infected	0.109	0.0726	0.2106
CDC1551 not infected-H37Rv not infected	0.0806	0.0729	0.3846
EAI not infected-H37Ra not infected	0.1187	0.0752	0.1906
EAI not infected-H37Rv not infected	0.0903	0.076	0.3606
H37Ra not infected-H37Rv not infected	-0.0283	0.0686	0.7647
Afri infected-BCG infected	0.1576	0.0874	0.1409
Afri infected-Beijing infected	-0.0638	0.0556	0.369
Afri infected-CDC1551 infected	0.0993	0.0658	0.2106
Afri infected-EAI infected	0.1124	0.0649	0.1591
Afri infected-H37Ra infected	0.1764	0.0702	0.0317
Afri infected-H37Rv infected	0.1835	0.0722	0.0305
BCG infected-Beijing infected	0.2214	0.0851	0.0268
BCG infected-CDC1551 infected	-0.0583	0.0926	0.6667
BCG infected-EAI infected	-0.0452	0.0753	0.6773
BCG infected-H37Ra infected	0.0188	0.0937	0.8981
BCG infected-H37Rv infected	0.0259	0.094	0.8501
Beijing infected-CDC1551 infected	0.1631	0.0612	0.0232
Beijing infected-EAI infected	0.1762	0.0623	0.0158

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.1634	0.5967	0.0423	0
groupAfri not infected	2.4272	0.639	11.3268	1e-04
groupBCG not infected	0.4143	0.9521	1.5133	0.6634
groupBeijing not infected	2.3782	0.5865	10.7859	1e-04
groupCDC1551 not infected	1.0524	0.6893	2.8645	0.1268
groupEAI not infected	1.6846	0.6812	5.3902	0.0134
groupH37Ra not infected	1.351	0.7642	3.8612	0.0771
groupH37Rv not infected	-0.0155	0.7984	0.9846	0.9845
groupAfri infected	3.4862	0.6419	32.6619	0
groupBCG infected	1.8422	0.8825	6.3104	0.0369
groupBeijing infected	3.6119	0.5845	37.0353	0
groupCDC1551 infected	2.579	0.6714	13.1845	1e-04
groupEAI infected	2.3989	0.6793	11.0112	4e-04
groupH37Ra infected	2.6859	0.7168	14.6711	2e-04
groupH37Rv infected	2.3284	0.7232	10.2615	0.0013

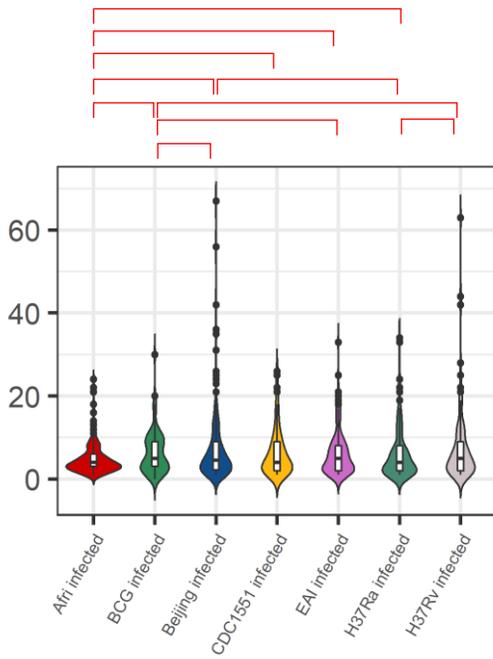
Contrast	estimate	SE	p.value
none not infected-Afri not infected	0.7362	0.6118	0.3897
none not infected-BCG not infected	2.749	0.9388	0.0239
none not infected-Beijing not infected	0.7851	0.5609	0.3384
none not infected-CDC1551 not infected	2.1109	0.6688	0.0153
none not infected-EAI not infected	1.4788	0.6612	0.114
none not infected-H37Ra not infected	1.8124	0.7267	0.0663
none not infected-H37Rv not infected	3.1789	0.7727	6e-04
none not infected-Afri infected	-0.3229	0.6143	0.7402
none not infected-BCG infected	1.3212	0.8681	0.2881
none not infected-Beijing infected	-0.4485	0.5584	0.5844
none not infected-CDC1551 infected	0.5843	0.6499	0.553
none not infected-EAI infected	0.7644	0.6589	0.4078
none not infected-H37Ra infected	0.4775	0.6767	0.6177
none not infected-H37Rv infected	0.835	0.6913	0.3897
Afri not infected-BCG not infected	2.0129	0.9882	0.1544
Afri not infected-Beijing not infected	0.0489	0.5653	0.9334
Afri not infected-CDC1551 not infected	1.3748	0.6868	0.1581
Afri not infected-EAI not infected	0.7426	0.6922	0.4501
Afri not infected-H37Ra not infected	1.0762	0.7976	0.3384
Afri not infected-H37Rv not infected	2.4427	0.8509	0.0258
BCG not infected-Beijing not infected	-1.9639	0.9638	0.1544
BCG not infected-CDC1551 not infected	-0.6381	1.0514	0.6853
BCG not infected-EAI not infected	-1.2703	0.8324	0.2881
BCG not infected-H37Ra not infected	-0.9567	1.1066	0.5799
BCG not infected-H37Rv not infected	0.4298	1.1354	0.8059
Beijing not infected-CDC1551 not infected	1.3258	0.631	0.1496
Beijing not infected-EAI not infected	0.6936	0.6611	0.4519
Beijing not infected-H37Ra not infected	1.0273	0.7566	0.3384
Beijing not infected-H37Rv not infected	2.3937	0.7908	0.0195
CDC1551 not infected-EAI not infected	-0.6322	0.7953	0.5844
CDC1551 not infected-H37Ra not infected	-0.2986	0.8218	0.8059
CDC1551 not infected-H37Rv not infected	1.0679	0.8434	0.3698
EAI not infected-H37Ra not infected	0.3336	0.873	0.8059
EAI not infected-H37Rv not infected	1.7001	0.91	0.1944
H37Ra not infected-H37Rv not infected	1.3665	0.8538	0.276
Afri infected-BCG infected	1.644	0.9248	0.2067
Afri infected-Beijing infected	-0.1257	0.5665	0.8657
Afri infected-CDC1551 infected	0.9072	0.6688	0.3384
Afri infected-EAI infected	1.0873	0.6953	0.2856
Afri infected-H37Ra infected	0.8003	0.7498	0.4501
Afri infected-H37Rv infected	1.1578	0.7825	0.302
BCG infected-Beijing infected	-1.7697	0.8937	0.1581
BCG infected-CDC1551 infected	-0.7368	0.9763	0.6004
BCG infected-EAI infected	-0.5567	0.7492	0.6004
BCG infected-H37Ra infected	-0.8437	1.0131	0.5799
BCG infected-H37Rv infected	-0.4862	1.0232	0.7544
Beijing infected-CDC1551 infected	1.0328	0.6103	0.2377
Beijing infected-EAI infected	1.213	0.6569	0.1945

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Sample

M2 – 6 h p.i.

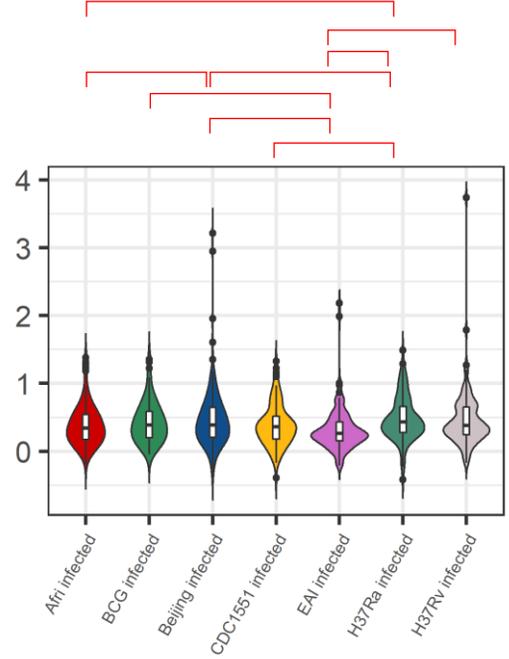
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.3459	0.1565	0
groupBCG infected	0.8519	0.2039	1e-04
groupBeijing infected	0.3098	0.1109	0.0071
groupCDC1551 infected	0.367	0.1412	0.0119
groupEAI infected	0.3906	0.1448	0.0091
groupH37Ra infected	0.7005	0.162	1e-04
groupH37Rv infected	0.1793	0.1664	0.2859

Contrast	estimate	SE	p.value
Afri infected - BCG infected	-0.8519	0.2039	0.0011
Afri infected - Beijing infected	-0.3098	0.1109	0.0247
Afri infected - CDC1551 infected	-0.367	0.1412	0.0256
Afri infected - EAI infected	-0.3906	0.1448	0.0249
Afri infected - H37Ra infected	-0.7005	0.162	0.0011
Afri infected - H37Rv infected	-0.1793	0.1664	0.3753
BCG infected - Beijing infected	0.5421	0.202	0.0249
BCG infected - CDC1551 infected	0.4849	0.2259	0.0688
BCG infected - EAI infected	0.4613	0.1516	0.0185
BCG infected - H37Ra infected	0.1514	0.2369	0.6128
BCG infected - H37Rv infected	0.6726	0.2381	0.0247
Beijing infected - CDC1551 infected	-0.0572	0.1262	0.6847
Beijing infected - EAI infected	-0.0808	0.1427	0.6337
Beijing infected - H37Ra infected	-0.3907	0.151	0.0256
Beijing infected - H37Rv infected	0.1305	0.1506	0.4812
CDC1551 infected - EAI infected	-0.0236	0.1761	0.8938
CDC1551 infected - H37Ra infected	-0.3335	0.1598	0.0723
CDC1551 infected - H37Rv infected	0.1877	0.1579	0.3591
EAI infected - H37Ra infected	-0.3099	0.1911	0.1781
EAI infected - H37Rv infected	0.2113	0.1926	0.3753
H37Ra infected - H37Rv infected	0.5212	0.1322	0.0015

d) Colocalization Index



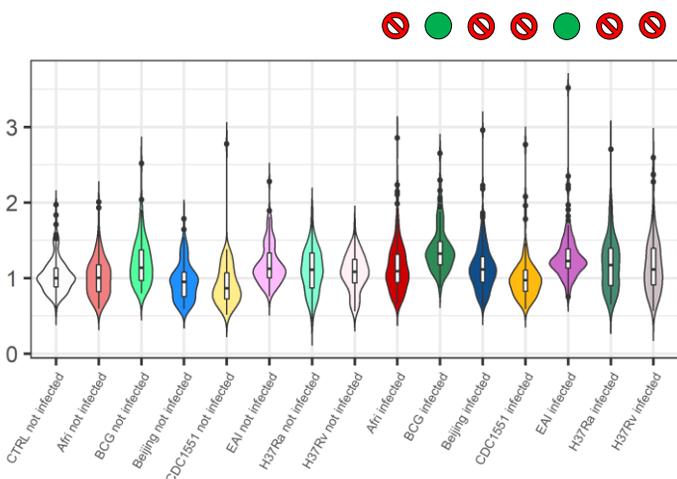
Par	Value	Std.Error	p.value
(Intercept)	-0.2367	0.0657	3e-04
groupBCG infected	0.1292	0.1071	0.2323
groupBeijing infected	0.2211	0.084	0.0108
groupCDC1551 infected	0.054	0.0961	0.5762
groupEAI infected	-0.1516	0.0943	0.1135
groupH37Ra infected	0.3735	0.0968	3e-04
groupH37Rv infected	0.1878	0.0981	0.0605

Contrast	estimate	SE	p.value
Afri infected - BCG infected	-0.1292	0.1071	0.3049
Afri infected - Beijing infected	-0.2211	0.084	0.036
Afri infected - CDC1551 infected	-0.054	0.0961	0.6318
Afri infected - EAI infected	0.1516	0.0943	0.1702
Afri infected - H37Ra infected	-0.3735	0.0968	0.002
Afri infected - H37Rv infected	-0.1878	0.0981	0.1163
BCG infected - Beijing infected	-0.0919	0.0994	0.4433
BCG infected - CDC1551 infected	0.0752	0.1098	0.5787
BCG infected - EAI infected	0.2808	0.1083	0.036
BCG infected - H37Ra infected	-0.2443	0.1104	0.0811
BCG infected - H37Rv infected	-0.0585	0.1116	0.6318
Beijing infected - CDC1551 infected	0.1671	0.0874	0.1163
Beijing infected - EAI infected	0.3727	0.0855	6e-04
Beijing infected - H37Ra infected	-0.1524	0.0882	0.1445
Beijing infected - H37Rv infected	0.0334	0.0896	0.7111
CDC1551 infected - EAI infected	0.2056	0.0974	0.0913
CDC1551 infected - H37Ra infected	-0.3195	0.0998	0.0093
CDC1551 infected - H37Rv infected	-0.1338	0.1011	0.2673
EAI infected - H37Ra infected	-0.5251	0.0981	0
EAI infected - H37Rv infected	-0.3394	0.0994	0.0062
H37Ra infected - H37Rv infected	0.1858	0.1018	0.1278

Grouping: Sample

M2 – 24 h p.i.

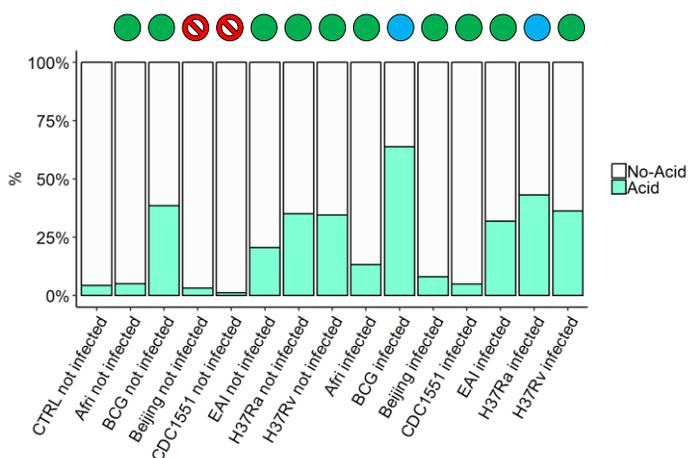
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	0.0139	0.0526	0.7917
groupAfrI not infected	-0.0115	0.0369	0.7548
groupBCG not infected	0.0972	0.054	0.0724
groupBeijing not infected	-0.0691	0.0351	0.0492
groupCDC1551 not infected	-0.0907	0.0411	0.0275
groupEAI not infected	0.0767	0.0392	0.0506
groupH37Ra not infected	-0.0554	0.044	0.2083
groupH37Rv not infected	-0.1223	0.0446	0.0061
groupAfrI infected	0.1065	0.0368	0.0039
groupBCG infected	0.2481	0.0534	0
groupBeijing infected	0.0653	0.034	0.055
groupCDC1551 infected	-0.023	0.0416	0.5806
groupEAI infected	0.1561	0.0393	1e-04
groupH37Ra infected	0.0528	0.0426	0.2149
groupH37Rv infected	-0.0563	0.0435	0.195

Contrast	estimate	SE	p.value
none not infected-AfrI not infected	-0.0024	0.056	0.9672
none not infected-BCG not infected	-0.1111	0.0691	0.2339
none not infected-Beijing not infected	0.0552	0.0549	0.4292
none not infected-CDC1551 not infected	0.0768	0.0588	0.3256
none not infected-EAI not infected	-0.0906	0.058	0.2376
none not infected-H37Ra not infected	0.0415	0.0602	0.5771
none not infected-H37Rv not infected	0.1084	0.0606	0.1932
none not infected-AfrI infected	-0.1203	0.056	0.1202
none not infected-BCG infected	-0.262	0.0687	0.0207
none not infected-Beijing infected	-0.0792	0.0541	0.2675
none not infected-CDC1551 infected	0.0091	0.0591	0.8966
none not infected-EAI infected	-0.17	0.0581	0.0498
none not infected-H37Ra infected	-0.0667	0.0592	0.3739
none not infected-H37Rv infected	0.0424	0.0598	0.5771
AfrI not infected-BCG not infected	-0.1087	0.0593	0.1202
AfrI not infected-Beijing not infected	0.0576	0.0376	0.202
AfrI not infected-CDC1551 not infected	0.0792	0.0441	0.1238
AfrI not infected-EAI not infected	-0.0882	0.0434	0.0835
AfrI not infected-H37Ra not infected	0.0438	0.0495	0.4553
AfrI not infected-H37Rv not infected	0.1108	0.0514	0.0664
BCG not infected-Beijing not infected	0.1663	0.0588	0.0158
BCG not infected-CDC1551 not infected	0.1879	0.064	0.0126
BCG not infected-EAI not infected	0.0205	0.049	0.7218
BCG not infected-H37Ra not infected	0.1525	0.0666	0.0498
BCG not infected-H37Rv not infected	0.2195	0.0672	0.0054
Beijing not infected-CDC1551 not infected	0.0216	0.0418	0.6575
Beijing not infected-EAI not infected	-0.1458	0.043	0.0041
Beijing not infected-H37Ra not infected	-0.0138	0.0478	0.8113
Beijing not infected-H37Rv not infected	0.0532	0.0488	0.3547
CDC1551 not infected-EAI not infected	-0.1674	0.0507	0.0051
CDC1551 not infected-H37Ra not infected	-0.0354	0.0509	0.5771
CDC1551 not infected-H37Rv not infected	0.0316	0.0515	0.5966
EAI not infected-H37Ra not infected	0.132	0.0542	0.0395
EAI not infected-H37Rv not infected	0.199	0.0551	0.0022
H37Ra not infected-H37Rv not infected	0.067	0.0474	0.2367
AfrI infected-BCG infected	-0.1417	0.0587	0.0395
AfrI infected-Beijing infected	0.0411	0.0367	0.3514
AfrI infected-CDC1551 infected	0.1294	0.0446	0.0131
AfrI infected-EAI infected	-0.0496	0.0434	0.3468
AfrI infected-H37Ra infected	0.0536	0.0484	0.3514
AfrI infected-H37Rv infected	0.1628	0.0504	0.0057
BCG infected-Beijing infected	0.1828	0.0576	0.0064
BCG infected-CDC1551 infected	0.2711	0.0638	3e-04
BCG infected-EAI infected	0.092	0.0484	0.1061
BCG infected-H37Ra infected	0.1953	0.0652	0.011
BCG infected-H37Rv infected	0.3045	0.066	1e-04
Beijing infected-CDC1551 infected	0.0883	0.0414	0.0674
Beijing infected-EAI infected	-0.0908	0.0422	0.0664

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.484	0.6133	0.0307	0
groupAfrI not infected	0.9109	0.6428	2.4866	0.1565
groupBCG not infected	1.7782	0.5663	5.9191	0.0017
groupBeijing not infected	0.5988	0.7014	1.82	0.3933
groupCDC1551 not infected	-0.2013	1.1277	0.8176	0.8583
groupEAI not infected	1.3244	0.5316	3.7599	0.0127
groupH37Ra not infected	1.4107	0.5892	4.0989	0.0166
groupH37Rv not infected	1.1886	0.5907	3.2824	0.0442
groupAfrI infected	1.9846	0.5506	7.2758	3e-04
groupBCG infected	2.805	0.5607	16.5272	0
groupBeijing infected	1.4598	0.5585	4.3052	0.009
groupCDC1551 infected	1.3262	0.7207	3.7668	0.0657
groupEAI infected	1.951	0.5188	7.0354	2e-04
groupH37Ra infected	2.3266	0.5764	10.243	1e-04
groupH37Rv infected	1.7108	0.5785	5.5335	0.0031

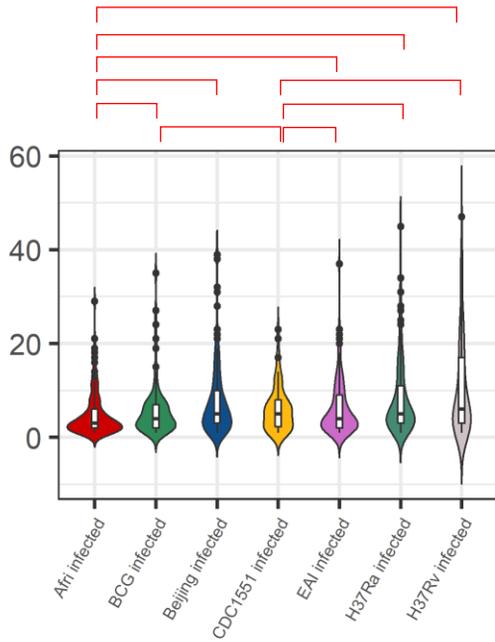
Contrast	estimate	SE	p.value
none not infected-AfrI not infected	2.5731	0.6172	0.001
none not infected-BCG not infected	1.7058	0.5856	0.0189
none not infected-Beijing not infected	2.8851	0.6739	0.001
none not infected-CDC1551 not infected	3.6853	1.1059	0.0077
none not infected-EAI not infected	2.1596	0.5412	0.0011
none not infected-H37Ra not infected	2.0732	0.5744	0.0032
none not infected-H37Rv not infected	2.2954	0.5766	0.0011
none not infected-AfrI infected	1.4994	0.5203	0.0192
none not infected-BCG infected	0.6789	0.5802	0.3944
none not infected-Beijing infected	2.0241	0.5265	0.0015
none not infected-CDC1551 infected	2.1577	0.6859	0.0116
none not infected-EAI infected	1.533	0.5264	0.0189
none not infected-H37Ra infected	1.1574	0.5541	0.1178
none not infected-H37Rv infected	1.7731	0.56	0.0116
AfrI not infected-BCG not infected	-0.8673	0.5955	0.2996
AfrI not infected-Beijing not infected	0.3121	0.6701	0.7089
AfrI not infected-CDC1551 not infected	1.1122	1.1161	0.4785
AfrI not infected-EAI not infected	-0.4135	0.543	0.5859
AfrI not infected-H37Ra not infected	-0.4998	0.6498	0.5859
AfrI not infected-H37Rv not infected	-0.2777	0.6554	0.7174
BCG not infected-Beijing not infected	1.1794	0.6683	0.2131
BCG not infected-CDC1551 not infected	1.9795	1.1224	0.2131
BCG not infected-EAI not infected	0.4538	0.3524	0.3666
BCG not infected-H37Ra not infected	0.3675	0.6323	0.6653
BCG not infected-H37Rv not infected	0.5896	0.6358	0.5065
Beijing not infected-CDC1551 not infected	0.8001	1.1422	0.6218
Beijing not infected-EAI not infected	-0.7256	0.6229	0.3944
Beijing not infected-H37Ra not infected	-0.8119	0.6971	0.3944
Beijing not infected-H37Rv not infected	-0.5898	0.701	0.5602
CDC1551 not infected-EAI not infected	-1.5257	1.0974	0.3237
CDC1551 not infected-H37Ra not infected	-1.6121	1.1087	0.2996
CDC1551 not infected-H37Rv not infected	-1.3899	1.1093	0.3679
EAI not infected-H37Ra not infected	-0.0863	0.5905	0.898
EAI not infected-H37Rv not infected	0.1358	0.5944	0.8569
H37Ra not infected-H37Rv not infected	0.2221	0.3913	0.6653
AfrI infected-BCG infected	-0.8205	0.4894	0.2459
AfrI infected-Beijing infected	0.5247	0.4148	0.3679
AfrI infected-CDC1551 infected	0.6583	0.6258	0.45
AfrI infected-EAI infected	0.0336	0.4105	0.9348
AfrI infected-H37Ra infected	-0.342	0.5317	0.6424
AfrI infected-H37Rv infected	0.2737	0.5451	0.7051
BCG infected-Beijing infected	1.3452	0.5093	0.0372
BCG infected-CDC1551 infected	1.4788	0.7104	0.1178
BCG infected-EAI infected	0.8541	0.3319	0.0423
BCG infected-H37Ra infected	0.4784	0.6097	0.5859
BCG infected-H37Rv infected	1.0942	0.6158	0.2131
Beijing infected-CDC1551 infected	0.1336	0.621	0.8569
Beijing infected-EAI infected	-0.4911	0.4352	0.4082

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Sample

M2 – 24 h p.i.

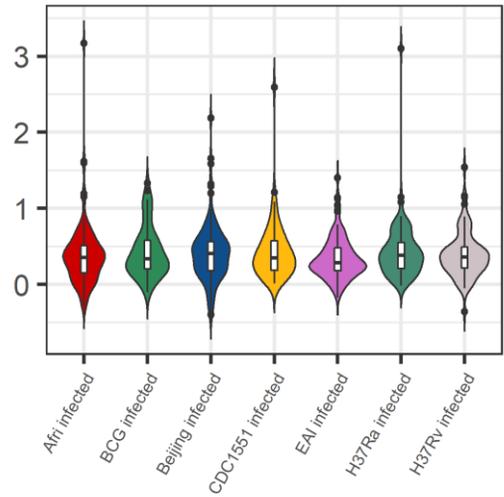
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.126	0.1674	0
groupBCG infected	0.7435	0.235	0.0025
groupBeijing infected	0.3785	0.1311	0.0055
groupCDC1551 infected	0.1261	0.1629	0.4418
groupEAI infected	0.6265	0.1691	5e-04
groupH37Ra infected	0.6752	0.186	6e-04
groupH37Rv infected	0.7453	0.1955	3e-04

Contrast	estimate	SE	p.value
Afri infected - BCG infected	-0.7435	0.235	0.0104
Afri infected - Beijing infected	-0.3785	0.1311	0.0164
Afri infected - CDC1551 infected	-0.1261	0.1629	0.6185
Afri infected - EAI infected	-0.6265	0.1691	0.0042
Afri infected - H37Ra infected	-0.6752	0.186	0.0042
Afri infected - H37Rv infected	-0.7453	0.1955	0.0042
BCG infected - Beijing infected	0.365	0.2331	0.1984
BCG infected - CDC1551 infected	0.6174	0.2601	0.0488
BCG infected - EAI infected	0.117	0.1746	0.6634
BCG infected - H37Ra infected	0.0684	0.271	0.8669
BCG infected - H37Rv infected	-0.0018	0.2748	0.9949
Beijing infected - CDC1551 infected	0.2524	0.1509	0.1744
Beijing infected - EAI infected	-0.248	0.1672	0.215
Beijing infected - H37Ra infected	-0.2966	0.1765	0.1744
Beijing infected - H37Rv infected	-0.3667	0.18	0.0968
CDC1551 infected - EAI infected	-0.5004	0.2047	0.0461
CDC1551 infected - H37Ra infected	-0.549	0.1879	0.0164
CDC1551 infected - H37Rv infected	-0.6192	0.1915	0.0104
EAI infected - H37Ra infected	-0.0486	0.2197	0.8669
EAI infected - H37Rv infected	-0.1187	0.2246	0.7399
H37Ra infected - H37Rv infected	-0.0701	0.1606	0.7748

d) Colocalization Index



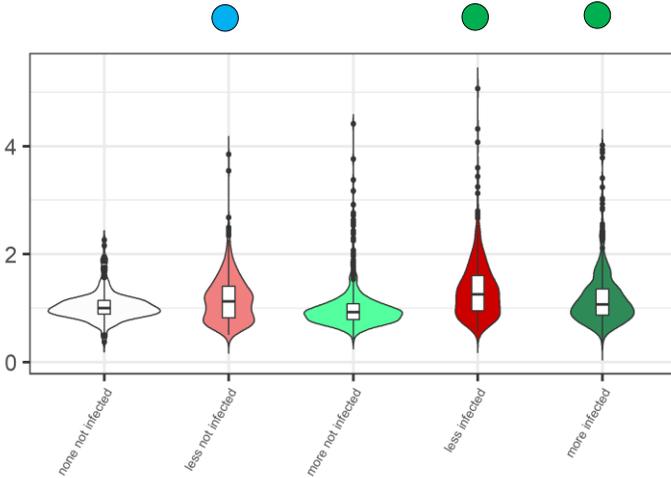
Par	Value	Std.Error	p.value
(Intercept)	-0.1201	0.0896	0.1804
groupBCG infected	0.1305	0.1528	0.3968
groupBeijing infected	0.1024	0.1093	0.3527
groupCDC1551 infected	0.1207	0.1284	0.3511
groupEAI infected	-0.1137	0.119	0.3433
groupH37Ra infected	0.121	0.1261	0.3414
groupH37Rv infected	0.0021	0.1336	0.9873

Contrast	estimate	SE	p.value
Afri infected - BCG infected	-0.1305	0.1528	0.6543
Afri infected - Beijing infected	-0.1024	0.1093	0.6543
Afri infected - CDC1551 infected	-0.1207	0.1284	0.6543
Afri infected - EAI infected	0.1137	0.119	0.6543
Afri infected - H37Ra infected	-0.121	0.1261	0.6543
Afri infected - H37Rv infected	-0.0021	0.1336	0.9985
BCG infected - Beijing infected	0.0281	0.1498	0.9985
BCG infected - CDC1551 infected	0.0098	0.1651	0.9985
BCG infected - EAI infected	0.2442	0.1298	0.4907
BCG infected - H37Ra infected	0.0095	0.1599	0.9985
BCG infected - H37Rv infected	0.1283	0.1637	0.6543
Beijing infected - CDC1551 infected	-0.0183	0.1235	0.9985
Beijing infected - EAI infected	0.2161	0.1154	0.4907
Beijing infected - H37Ra infected	-0.0185	0.1226	0.9985
Beijing infected - H37Rv infected	0.1003	0.1274	0.6543
CDC1551 infected - EAI infected	0.2344	0.1374	0.4907
CDC1551 infected - H37Ra infected	-3e-04	0.1352	0.9985
CDC1551 infected - H37Rv infected	0.1186	0.1404	0.6543
EAI infected - H37Ra infected	-0.2346	0.1318	0.4907
EAI infected - H37Rv infected	-0.1158	0.1365	0.6543
H37Ra infected - H37Rv infected	0.1188	0.1235	0.6543

Grouping: Virulence (absolute)

M1 – 6 h p.i.

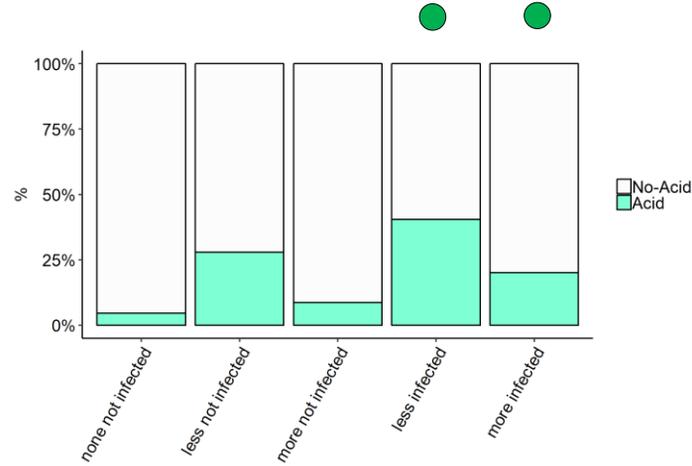
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	0.002	0.0846	0.9808
groupless not infected	0.0275	0.0535	0.6078
groupmore not infected	0.0253	0.0549	0.6455
groupless infected	0.146	0.0536	0.0065
groupmore infected	0.1435	0.055	0.0091

Contrast	estimate	SE	p.value
none not infected-less not infected	-0.0275	0.0535	0.8607
none not infected-more not infected	-0.0253	0.0549	0.8607
none not infected-less infected	-0.146	0.0536	0.0174
none not infected-more infected	-0.1435	0.055	0.0183
less not infected-more not infected	0.0022	0.0526	0.9666
less infected-more infected	0.0024	0.0527	0.9666
less not infected-less infected	-0.1185	0.0111	0
more not infected-more infected	-0.1183	0.0119	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.6062	0.8803	0.0272	0
groupless not infected	1.0881	0.6079	2.9687	0.0735
groupmore not infected	0.3838	0.6742	1.4679	0.5692
groupless infected	2.0723	0.6054	7.9427	6e-04
groupmore infected	2.024	0.6518	7.5685	0.0019

Contrast	estimate	SE	p.value
none not infected-less not infected	-1.0881	0.6079	0.1175
none not infected-more not infected	-0.3838	0.6742	0.6505
none not infected-less infected	-2.0723	0.6054	0.0017
none not infected-more infected	-2.024	0.6518	0.0038
less not infected-more not infected	0.7043	0.5425	0.2589
less infected-more infected	0.0483	0.5109	0.9248
less not infected-less infected	-0.9841	0.1835	0
more not infected-more infected	-1.6402	0.2963	0

OBSERVATIONS

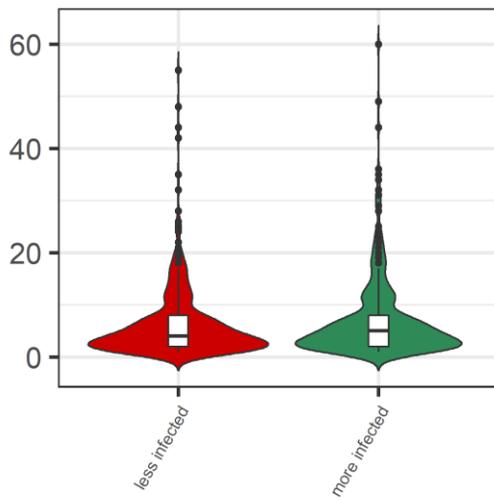
In M1 macrophages, none of the category is blocking the acidification of the phagolysosomes irrespective from the time point considered. The percentage of acidified macrophage was anyway found higher (40%) in the category less virulent at 6 h p.i.

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Virulence (absolute)

M1 – 6 h p.i.

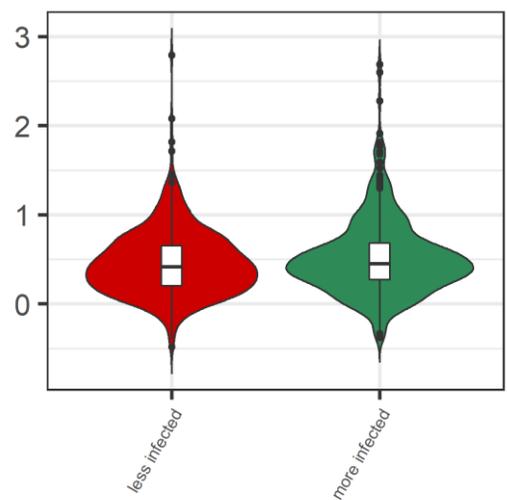
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.4301	0.0942	0
groupmore infected	0.0276	0.1075	0.7984

Contrast	estimate	SE	p.value
less infected - more infected	-0.0276	0.1075	0.7984

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	0.0118	0.079	0.881
groupmore infected	0.0669	0.0917	0.4683

Contrast	estimate	SE	p.value
less infected - more infected	-0.0669	0.0917	0.4683

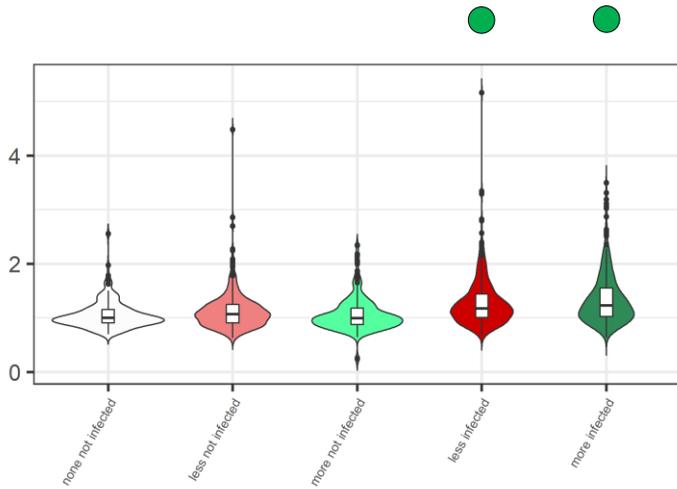
OBSERVATIONS

No statistically relevant differences in effective MOI were observed among the categories in M1 macrophages, irrespectively from the time point considered. Colocalization with acidified compartments was similar among all the groups.

Grouping: Virulence (absolute)

M1 – 24 h.p.i.

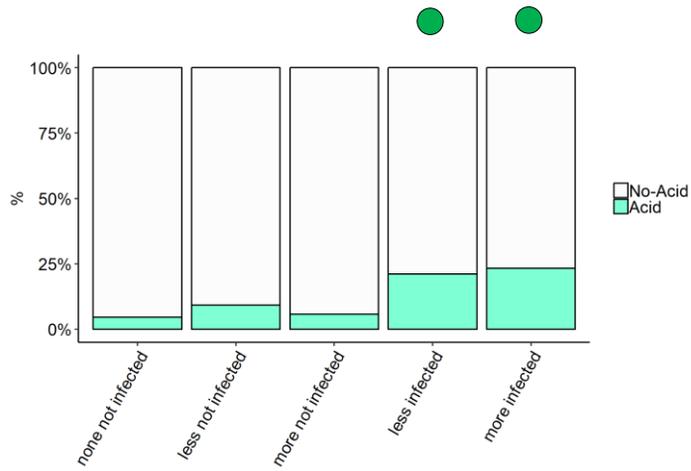
a) Phagolysosomal acidification



Par	Value	Std.Error	p-value
(Intercept)	0.0084	0.0437	0.848
groupless not infected	0.0103	0.0292	0.7237
groupmore not infected	0.0295	0.0303	0.3296
groupless infected	0.1331	0.029	0
groupmore infected	0.2074	0.0299	0

Contrast	estimate	SE	p-value
none not infected-less not infected	-0.0103	0.0292	0.7237
none not infected-more not infected	-0.0295	0.0303	0.4395
none not infected-less infected	-0.1331	0.029	0
none not infected-more infected	-0.2074	0.0299	0
less not infected-more not infected	-0.0192	0.0272	0.5482
less infected-more infected	-0.0743	0.0265	0.0082
less not infected-less infected	-0.1228	0.0142	0
more not infected-more infected	-0.1779	0.0165	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.1891	0.4561	0.0412	0
groupless not infected	0.6196	0.4567	1.8581	0.1749
groupmore not infected	0.3275	0.498	1.3875	0.5107
groupless infected	1.6859	0.436	5.3974	1e-04
groupmore infected	1.9742	0.4445	7.201	0

Contrast	estimate	SE	p-value
none not infected-less not infected	-0.6196	0.4567	0.2799
none not infected-more not infected	-0.3275	0.498	0.5107
none not infected-less infected	-1.6859	0.436	2e-04
none not infected-more infected	-1.9742	0.4445	0
less not infected-more not infected	0.292	0.3643	0.4831
less infected-more infected	-0.2883	0.2523	0.3376
less not infected-less infected	-1.0664	0.2188	0
more not infected-more infected	-1.6467	0.2934	0

OBSERVATIONS

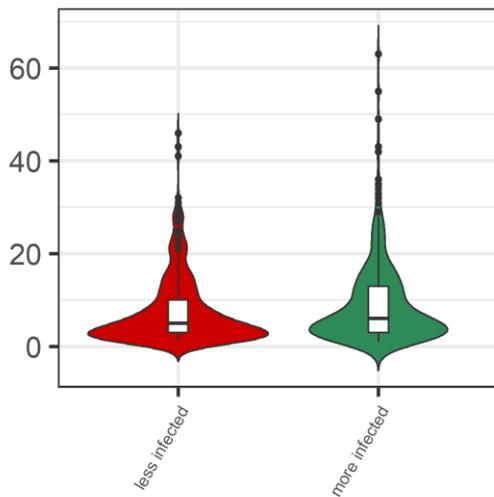
In M1 macrophages, none of the category is blocking the acidification of the phagolysosomes irrespective from the time point considered. The percentage of acidified macrophage decreased (25%) in the category less virulent and was found similar to the one observed for more virulent strains.

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Virulence (absolute)

M1 – 24 h p.i.

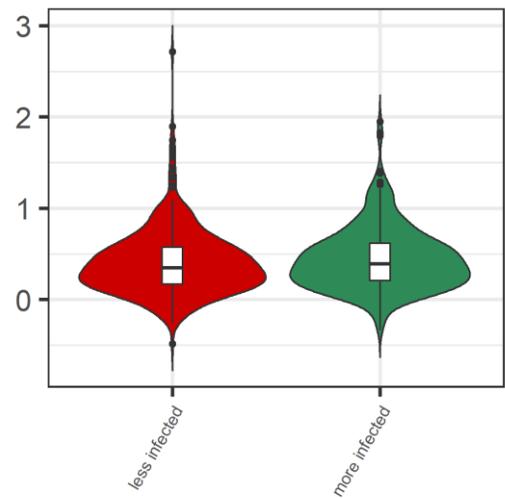
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.7065	0.1352	0
groupmore infected	-0.0243	0.0952	0.7994

Contrast	estimate	SE	p.value
less infected - more infected	0.0243	0.0952	0.7994

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	-0.0793	0.0691	0.2518
groupmore infected	0.146	0.0958	0.1326

Contrast	estimate	SE	p.value
less infected - more infected	-0.146	0.0958	0.1326

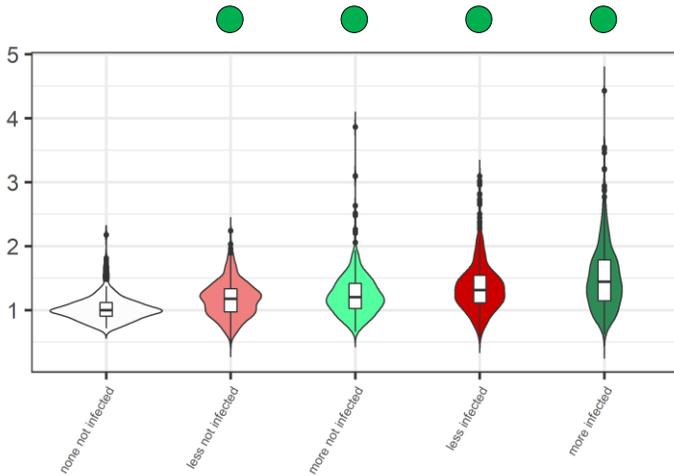
OBSERVATIONS

No statistically relevant differences in effective MOI were observed among the categories in M1 macrophages, irrespectively from the time point considered. Colocalization with acidified compartments was similar among all the groups.

Grouping: Virulence (absolute)

M2 – 6 h p.i.

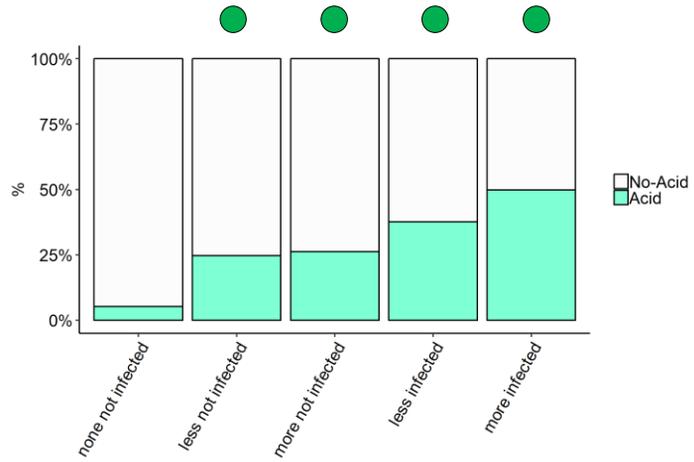
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	-0.0062	0.0521	0.9055
groupless not infected	0.1224	0.0411	0.003
groupmore not infected	0.1602	0.0421	1e-04
groupless infected	0.2403	0.0412	0
groupmore infected	0.2978	0.0421	0

Contrast	estimate	SE	p.value
none not infected-less not infected	-0.1224	0.0411	0.0039
none not infected-more not infected	-0.1602	0.0421	2e-04
none not infected-less infected	-0.2403	0.0412	0
none not infected-more infected	-0.2978	0.0421	0
less not infected-more not infected	-0.0377	0.0402	0.3484
less infected-more infected	-0.0576	0.0403	0.1752
less not infected-less infected	-0.1178	0.0107	0
more not infected-more infected	-0.1377	0.0113	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.2188	0.6551	0.04	0
groupless not infected	1.7554	0.5414	5.7858	0.0012
groupmore not infected	1.4685	0.5411	4.3427	0.0067
groupless infected	2.8003	0.5384	16.4504	0
groupmore infected	2.9692	0.5354	19.4768	0

Contrast	estimate	SE	p.value
none not infected-less not infected	-1.7554	0.5414	0.0019
none not infected-more not infected	-1.4685	0.5411	0.0089
none not infected-less infected	-2.8003	0.5384	0
none not infected-more infected	-2.9692	0.5354	0
less not infected-more not infected	0.2869	0.4268	0.5731
less infected-more infected	-0.1689	0.4134	0.6829
less not infected-less infected	-1.0449	0.1839	0
more not infected-more infected	-1.5007	0.1895	0

OBSERVATIONS

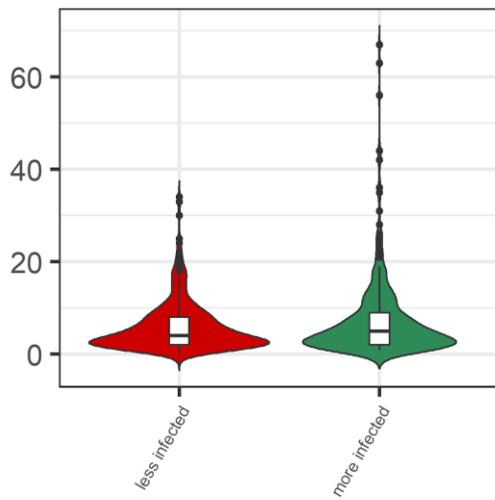
In M2 macrophages, none of the category is blocking the acidification of the phagolysosomes at 6 h p.i. and between 30% (less virulent) and 50% (more virulent) of cells are showing acidification. A bystander effect in non-infected M2a macrophages was observed for both less and more virulent strains.

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, stastically not significant

Grouping: Virulence (absolute)

M2 – 6 h p.i.

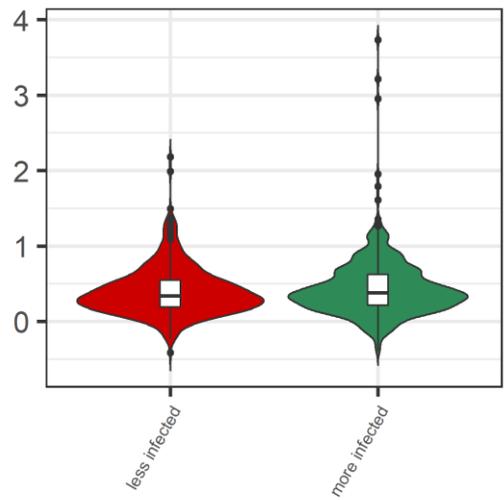
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.7065	0.1352	0
groupmore infected	-0.0243	0.0952	0.7994

Contrast	estimate	SE	p.value
less infected - more infected	0.0243	0.0952	0.7994

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	-0.1285	0.0572	0.0249
groupmore infected	0.04	0.0641	0.5354

Contrast	estimate	SE	p.value
less infected - more infected	-0.04	0.0641	0.5354

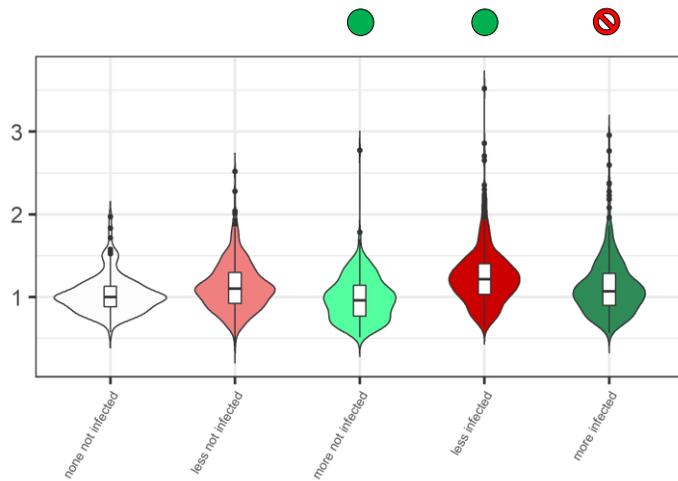
OBSERVATIONS

No statistically relevant differences in effective MOI were observed among the categories in M2 macrophages, irrespectively from the time point considered. Colocalization with acidified compartments was similar among all the groups.

Grouping: Virulence (absolute)

M2 – 24 h.p.i.

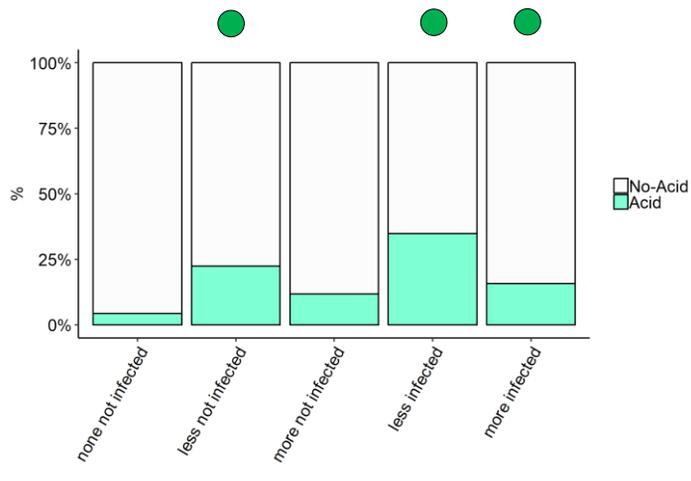
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	0.0091	0.0541	0.8666
groupless not infected	0.0102	0.0281	0.7169
groupmore not infected	-0.0721	0.0293	0.014
groupless infected	0.1174	0.028	0
groupmore infected	0.0238	0.029	0.4125

Contrast	estimate	SE	p.value
none not infected-less not infected	-0.0102	0.0281	0.7169
none not infected-more not infected	0.0721	0.0293	0.0186
none not infected-less infected	-0.1174	0.028	1e-04
none not infected-more infected	-0.0238	0.029	0.4714
less not infected-more not infected	0.0823	0.0261	0.0027
less infected-more infected	0.0936	0.0257	6e-04
less not infected-less infected	-0.1072	0.0123	0
more not infected-more infected	-0.0958	0.0146	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.585	0.6451	0.0277	0
groupless not infected	1.3491	0.4714	3.8539	0.0042
groupmore not infected	0.9519	0.5148	2.5906	0.0644
groupless infected	2.2141	0.4669	9.1531	0
groupmore infected	1.6312	0.4988	5.1099	0.0011

Contrast	estimate	SE	p.value
none not infected-less not infected	-1.3491	0.4714	0.0084
none not infected-more not infected	-0.9519	0.5148	0.0737
none not infected-less infected	-2.2141	0.4669	0
none not infected-more infected	-1.6312	0.4988	0.0029
less not infected-more not infected	0.3972	0.3048	0.1926
less infected-more infected	0.5829	0.2669	0.0386
less not infected-less infected	-0.865	0.176	0
more not infected-more infected	-0.6793	0.2867	0.0285

OBSERVATIONS

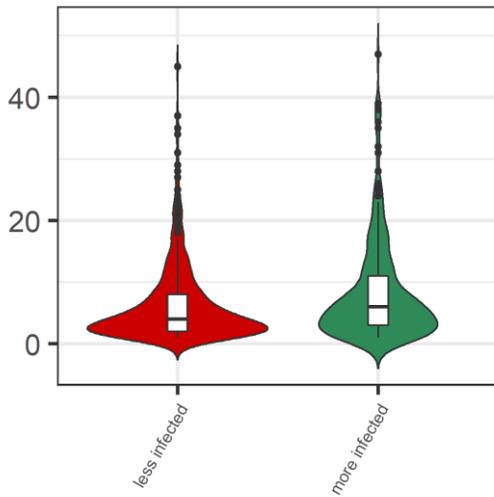
However, as the infection progresses, more virulent strains are blocking the acidification of the phagolysosomes. The percentage of acidified cells remained constant for less virulent strains, whereas it dropped below 25% for more virulent strains.

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Virulence (absolute)

M2 – 24 h p.i.

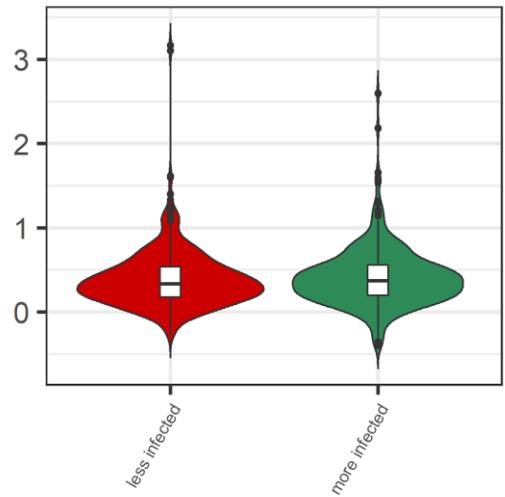
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.5323	0.1172	0
groupmore infected	0.0789	0.1043	0.4523

Contrast	estimate	SE	p.value
less infected - more infected	-0.0789	0.1043	0.4523

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	-0.1034	0.0503	0.0402
groupmore infected	0.0671	0.0705	0.3444

Contrast	estimate	SE	p.value
less infected - more infected	-0.0671	0.0705	0.3444

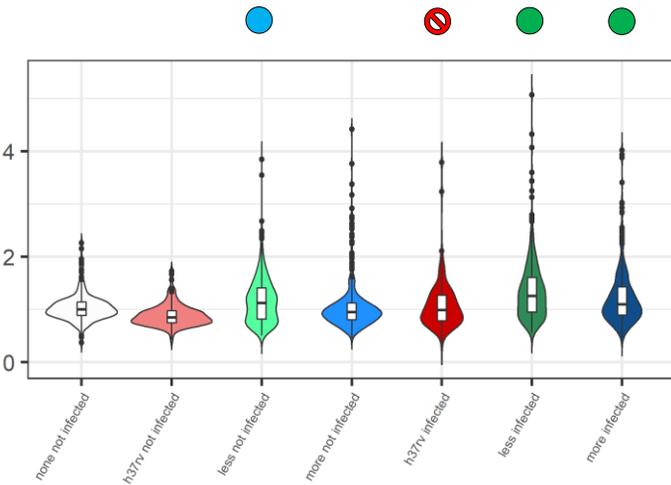
OBSERVATIONS

No statistically relevant differences in effective MOI were observed among the categories in M2 macrophages, irrespectively from the time point considered. Colocalization with acidified compartments was similar among all the groups.

Grouping: Virulence (relative)

M1 – 6 h p.i.

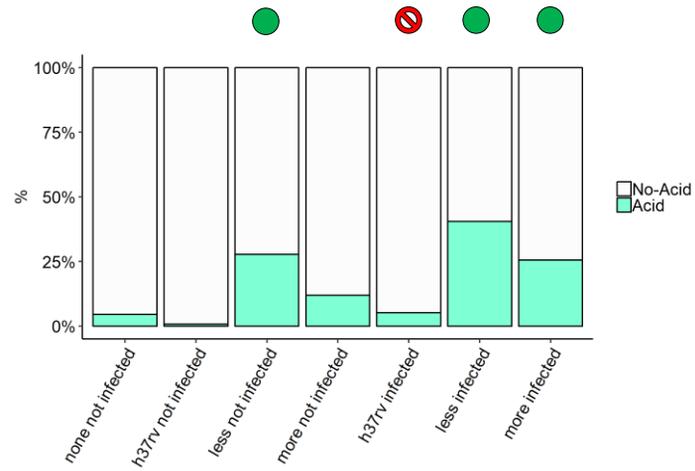
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	0.0024	0.0838	0.9773
group $h37rv$ not infected	0.0038	0.0834	0.9633
groupless not infected	0.0285	0.0539	0.5975
groupmore not infected	0.0336	0.0605	0.5786
group $h37rv$ infected	0.1172	0.0839	0.1625
groupless infected	0.147	0.054	0.0065
groupmore infected	0.1569	0.0605	0.0096

Contrast	estimate	SE	p.value
none not infected- $h37rv$ not infected	-0.0038	0.0834	0.9633
none not infected-less not infected	-0.0285	0.0539	0.9596
none not infected-more not infected	-0.0336	0.0605	0.9596
none not infected- $h37rv$ infected	-0.1172	0.0839	0.4063
none not infected-less infected	-0.147	0.054	0.0245
none not infected-more infected	-0.1569	0.0605	0.0288
$h37rv$ not infected-less not infected	-0.0246	0.0834	0.9596
$h37rv$ not infected-more not infected	-0.0298	0.0871	0.9596
less not infected-more not infected	-0.0052	0.0576	0.9633
$h37rv$ infected-less infected	-0.0298	0.0839	0.9596
$h37rv$ infected-more infected	-0.0397	0.0875	0.9596
less infected-more infected	-0.0099	0.0576	0.9633
$h37rv$ not infected- $h37rv$ infected	-0.1134	0.0232	0
less not infected-less infected	-0.1185	0.0111	0
more not infected-more infected	-0.1233	0.0139	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.5774	0.8537	0.0279	0
group $h37rv$ not infected	-0.5903	1.3921	0.5542	0.6715
groupless not infected	1.115	0.6074	3.0497	0.0664
groupmore not infected	0.5864	0.7119	1.7976	0.4101
group $h37rv$ infected	1.4056	1.0433	4.078	0.1779
groupless infected	2.0973	0.6048	8.144	5e-04
groupmore infected	2.2031	0.6926	9.0534	0.0015

Contrast	estimate	SE	p-value
none not infected- $h37rv$ not infected	0.5903	1.3921	0.7195
none not infected-less not infected	-1.115	0.6074	0.177
none not infected-more not infected	-0.5864	0.7119	0.5552
none not infected- $h37rv$ infected	-1.4056	1.0433	0.3812
none not infected-less infected	-2.0973	0.6048	0.0026
none not infected-more infected	-2.2031	0.6926	0.0055
$h37rv$ not infected-less not infected	-1.7053	1.3492	0.3867
$h37rv$ not infected-more not infected	-1.1767	1.4085	0.5552
less not infected-more not infected	0.5286	0.5785	0.5552
$h37rv$ infected-less infected	-0.6917	0.9818	0.5552
$h37rv$ infected-more infected	-0.7975	1.0485	0.5552
less infected-more infected	-0.1059	0.5515	0.8478
$h37rv$ not infected- $h37rv$ infected	-1.9959	1.1047	0.177
less not infected-less infected	-0.9823	0.1833	0
more not infected-more infected	-1.6167	0.3103	0

OBSERVATIONS

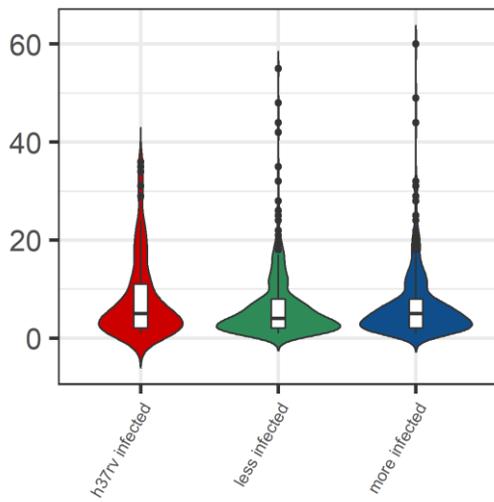
In M1 macrophages, H37Rv is blocking the phagolysosomal acidification at 6 h p.i. The percentage of acidified macrophase was anyway found higher (40%) in the category less virulent at 6 h p.i.

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Virulence (relative)

M1 – 6 h p.i.

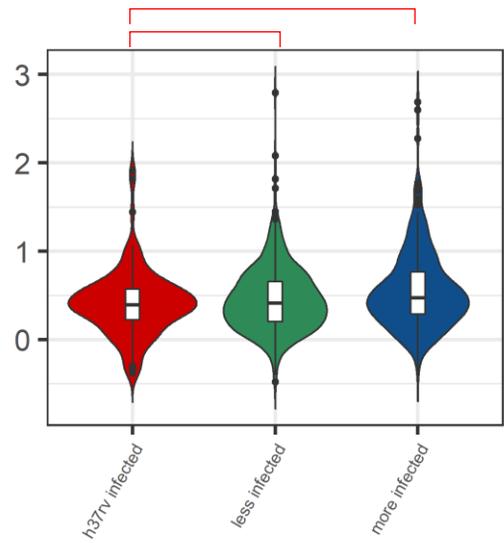
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.4927	0.1598	0
groupless infected	-0.0639	0.1702	0.7087
groupmore infected	-0.0496	0.1761	0.7793

Contrast	estimate	SE	p.value
h37rv infected - less infected	0.0639	0.1702	0.9031
h37rv infected - more infected	0.0496	0.1761	0.9031
less infected - more infected	-0.0143	0.1171	0.9031

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	-0.274	0.1405	0.0516
groupless infected	0.3106	0.1384	0.0283
groupmore infected	0.4833	0.1413	0.0011

Contrast	estimate	SE	p.value
h37rv infected - less infected	-0.3106	0.1384	0.0425
h37rv infected - more infected	-0.4833	0.1413	0.0033
less infected - more infected	-0.1727	0.0945	0.0724

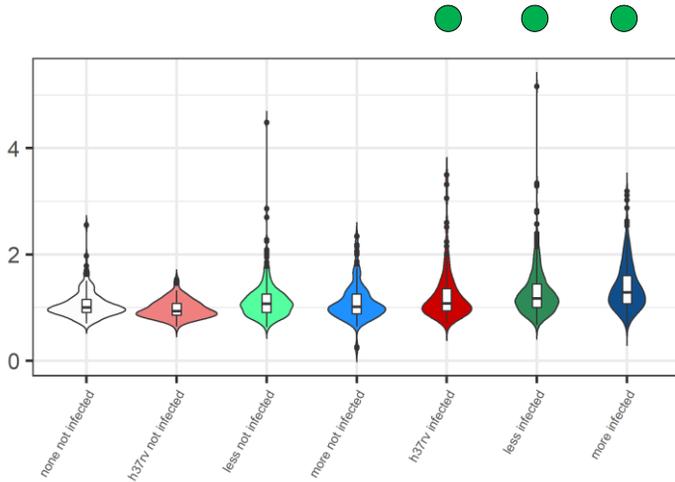
OBSERVATIONS

Colocalization with acidified compartments was found higher for more virulent strains at 6 h p.i.

Grouping: Virulence (relative)

M1 – 24 h p.i.

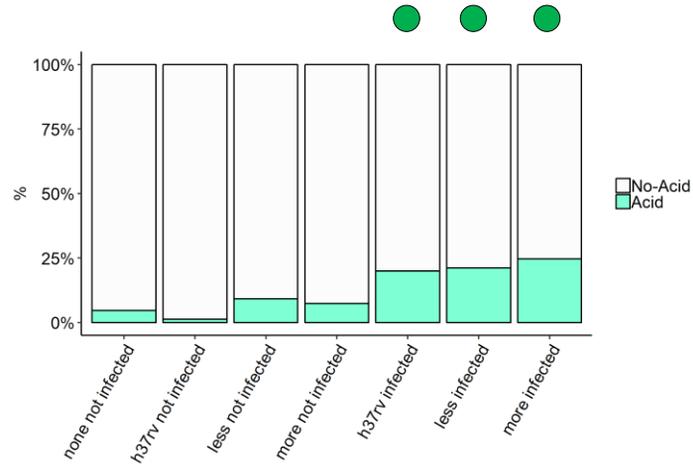
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	0.0095	0.0424	0.8227
grouph37rv not infected	0.0183	0.0448	0.6826
groupless not infected	0.0111	0.0292	0.7038
groupmore not infected	0.0323	0.0328	0.3242
grouph37rv infected	0.155	0.0436	4e-04
groupless infected	0.1336	0.029	0
groupmore infected	0.2277	0.0321	0

Contrast	estimate	SE	p.value
none not infected-h37rv not infected	-0.0183	0.0448	0.812
none not infected-less not infected	-0.0111	0.0292	0.812
none not infected-more not infected	-0.0323	0.0328	0.5404
none not infected-h37rv infected	-0.155	0.0436	0.001
none not infected-less infected	-0.1336	0.029	0
none not infected-more infected	-0.2277	0.0321	0
h37rv not infected-less not infected	0.0072	0.0432	0.8672
h37rv not infected-more not infected	-0.014	0.0456	0.8131
less not infected-more not infected	-0.0212	0.0297	0.712
h37rv infected-less infected	0.0213	0.0416	0.812
h37rv infected-more infected	-0.0728	0.0436	0.1791
less infected-more infected	-0.0941	0.0289	0.0024
h37rv not infected-h37rv infected	-0.1367	0.03	0
less not infected-less infected	-0.1225	0.0142	0
more not infected-more infected	-0.1954	0.0197	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.183	0.4538	0.0415	0
grouph37rv not infected	-1.0349	1.1117	0.3552	0.3519
groupless not infected	0.6288	0.457	1.8753	0.1689
groupmore not infected	0.5519	0.5093	1.7366	0.2785
grouph37rv infected	1.9228	0.5288	6.84	3e-04
groupless infected	1.6912	0.4361	5.426	1e-04
groupmore infected	1.9824	0.4561	7.2598	0

Contrast	estimate	SE	p.value
none not infected-h37rv not infected	1.0349	1.1117	0.4398
none not infected-less not infected	-0.6288	0.457	0.2815
none not infected-more not infected	-0.5519	0.5093	0.3836
none not infected-h37rv infected	-1.9228	0.5288	8e-04
none not infected-less infected	-1.6912	0.4361	4e-04
none not infected-more infected	-1.9824	0.4561	1e-04
h37rv not infected-less not infected	-1.6637	1.0623	0.2514
h37rv not infected-more not infected	-1.5869	1.0787	0.2649
less not infected-more not infected	0.0768	0.3765	0.8784
h37rv infected-less infected	0.2316	0.3862	0.6331
h37rv infected-more infected	-0.0596	0.3893	0.8784
less infected-more infected	-0.2911	0.2702	0.3836
h37rv not infected-h37rv infected	-2.9577	1.0388	0.011
less not infected-less infected	-1.0625	0.2188	0
more not infected-more infected	-1.4304	0.3144	0

OBSERVATIONS

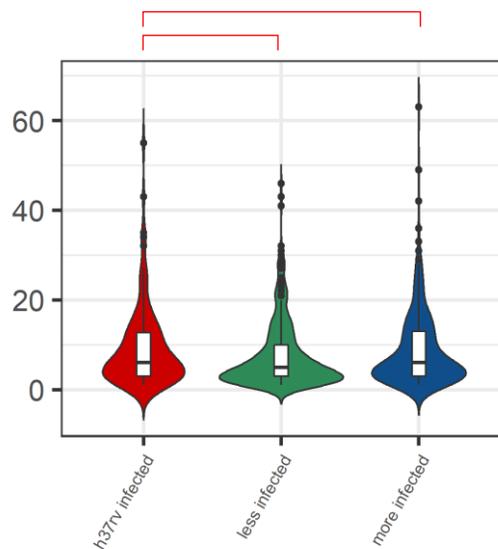
At 24 h p.i. none of the category is blocking the acidification of the phagolysosomes. The percentage of acidified macrophase was found below 25%.

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, stastically not significant

Grouping: Virulence (relative)

M1 – 24 h p.i.

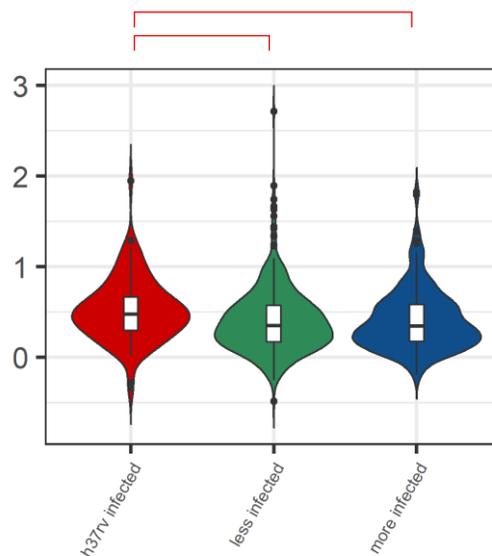
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.4019	0.1849	0
groupless infected	0.3265	0.1363	0.0197
groupmore infected	0.3724	0.1425	0.0113

Contrast	estimate	SE	p.value
h37rv infected - less infected	-0.3265	0.1363	0.0295
h37rv infected - more infected	-0.3724	0.1425	0.0295
less infected - more infected	-0.0459	0.0961	0.6351

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	0.3599	0.1222	0.0033
groupless infected	-0.4429	0.1358	0.0018
groupmore infected	-0.4322	0.1442	0.0039

Contrast	estimate	SE	p.value
h37rv infected - less infected	0.4429	0.1358	0.0054
h37rv infected - more infected	0.4322	0.1442	0.0059
less infected - more infected	-0.0107	0.0999	0.9152

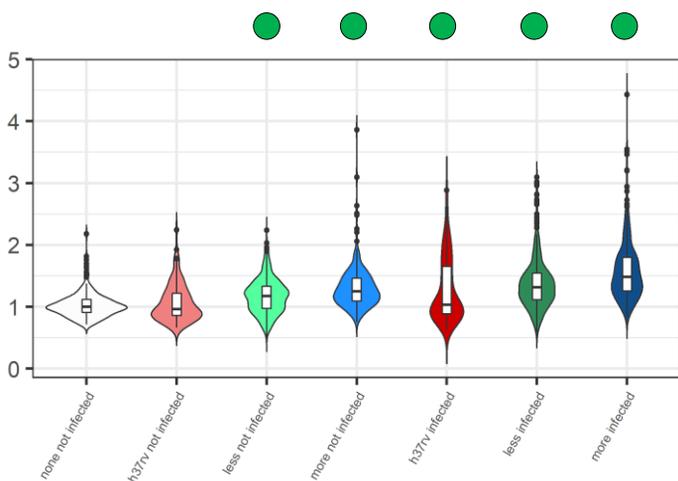
OBSERVATIONS

Colocalization was found higher for H37Rv at 24 h p.i. H37Rv showed higher MOI at 24 h p.i.

Grouping: Virulence (relative)

M2 – 6 h p.i.

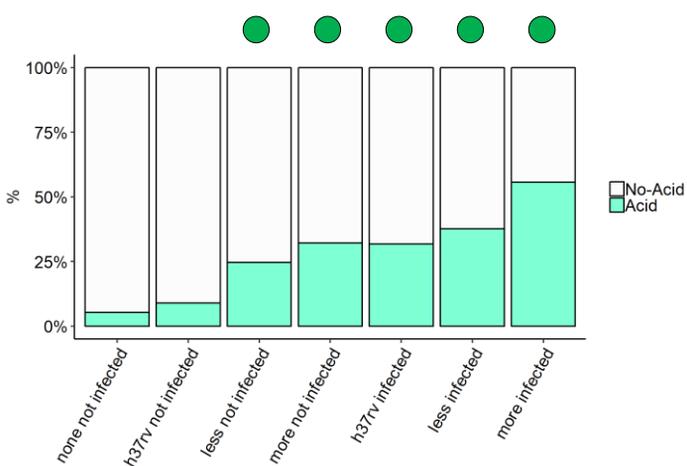
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	-0.0092	0.0468	0.8444
grouph37rv not infected	0.0701	0.062	0.2582
groupless not infected	0.1306	0.0407	0.0013
groupmore not infected	0.2014	0.0457	0
grouph37rv infected	0.196	0.0622	0.0016
groupless infected	0.249	0.0408	0
groupmore infected	0.3473	0.0456	0

Contrast	estimate	SE	p.value
none not infected-h37rv not infected	-0.0701	0.062	0.298
none not infected-less not infected	-0.1306	0.0407	0.0029
none not infected-more not infected	-0.2014	0.0457	0
none not infected-h37rv infected	-0.196	0.0622	0.0031
none not infected-less infected	-0.249	0.0408	0
none not infected-more infected	-0.3473	0.0456	0
h37rv not infected-less not infected	-0.0605	0.0621	0.3534
h37rv not infected-more not infected	-0.1313	0.0648	0.0584
less not infected-more not infected	-0.0708	0.043	0.1247
h37rv infected-less infected	-0.053	0.0623	0.3947
h37rv infected-more infected	-0.1512	0.0648	0.033
less infected-more infected	-0.0982	0.043	0.0338
h37rv not infected-h37rv infected	-0.1259	0.0224	0
less not infected-less infected	-0.1184	0.0108	0
more not infected-more infected	-0.1459	0.0132	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	sc	OR	pvalue
(Intercept)	-3.1983	0.6306	0.0408	0
grouph37rv not infected	0.0529	0.8209	1.0543	0.9486
groupless not infected	1.7791	0.5405	5.9247	0.001
groupmore not infected	1.806	0.5651	6.0859	0.0014
grouph37rv infected	2.4272	0.7421	11.3272	0.0011
groupless infected	2.8201	0.5373	16.7779	0
groupmore infected	3.1457	0.5618	23.2369	0

Contrast	estimate	SE	p.value
none not infected-h37rv not infected	-0.0529	0.8209	0.9526
none not infected-less not infected	-1.7791	0.5405	0.0023
none not infected-more not infected	-1.806	0.5651	0.0026
none not infected-h37rv infected	-2.4272	0.7421	0.0023
none not infected-less infected	-2.8201	0.5373	0
none not infected-more infected	-3.1457	0.5618	0
h37rv not infected-less not infected	-1.7262	0.7637	0.0357
h37rv not infected-more not infected	-1.7531	0.77	0.0357
less not infected-more not infected	-0.0268	0.4515	0.9526
h37rv infected-less infected	-0.3929	0.6699	0.6433
h37rv infected-more infected	-0.7185	0.6881	0.4041
less infected-more infected	-0.3257	0.4428	0.5775
h37rv not infected-h37rv infected	-2.3743	0.5099	0
less not infected-less infected	-1.0409	0.1835	0
more not infected-more infected	-1.3398	0.2066	0

OBSERVATIONS

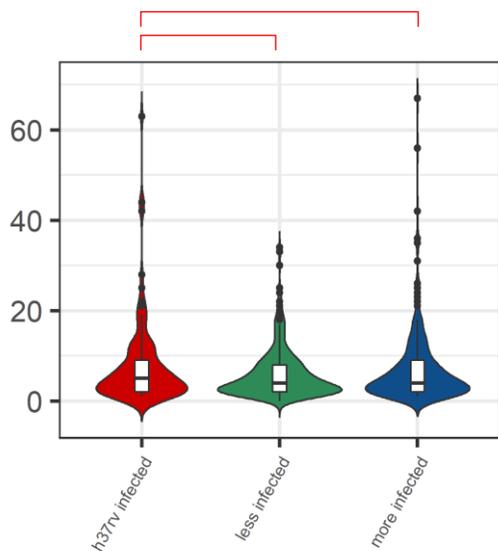
In M2 macrophages, none of the category is blocking the acidification of the phagolysosomes at 6 h p.i. and between 30% (less virulent and H37Rv) and 50% (more virulent) of cells are showing acidification. A bystander effect in non-infected M2a macrophages was consistently observed for less virulent strains.

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Virulence (relative)

M2 – 6 h p.i.

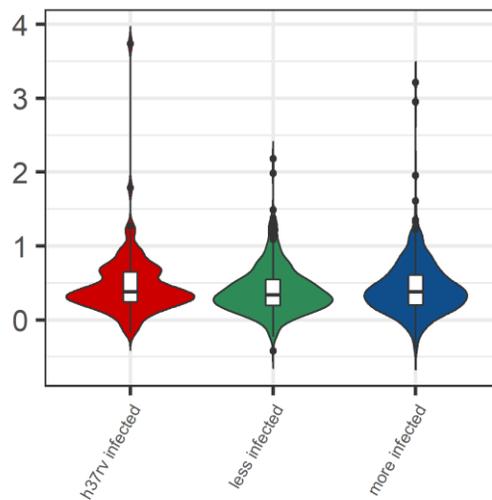
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.4019	0.1849	0
groupless infected	0.3265	0.1363	0.0197
groupmore infected	0.3724	0.1425	0.0113

Contrast	estimate	SE	p.value
h37rv infected - less infected	-0.3265	0.1363	0.0295
h37rv infected - more infected	-0.3724	0.1425	0.0295
less infected - more infected	-0.0459	0.0961	0.6351

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	-0.1544	0.0986	0.1178
groupless infected	0.035	0.1015	0.7316
groupmore infected	0.0949	0.1037	0.3634

Contrast	estimate	SE	p.value
h37rv infected - less infected	-0.035	0.1015	0.7316
h37rv infected - more infected	-0.0949	0.1037	0.5812
less infected - more infected	-0.06	0.0689	0.5812

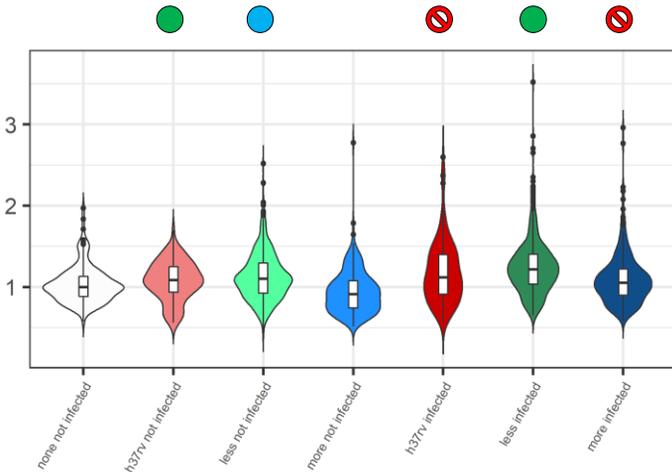
OBSERVATIONS

H37Rv showed higher MOI. Colocalization with acidified compartments was similar among all the groups.

Grouping: Virulence (relative)

M2 – 24 h p.i.

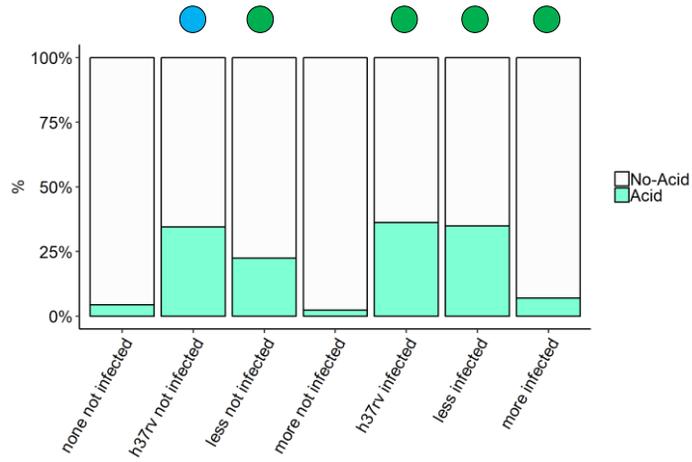
a) Phagolysosomal acidification



Par	Value	Std.Error	p.value
(Intercept)	0.0101	0.0549	0.8537
grouph37rv not infected	-0.0953	0.0434	0.0281
groupless not infected	0.0111	0.028	0.6931
groupmore not infected	-0.0639	0.0316	0.0429
grouph37rv infected	-0.0298	0.0423	0.4809
groupless infected	0.1181	0.0279	0
groupmore infected	0.0422	0.0311	0.1752

Contrast	estimate	SE	p.value
none not infected-h37rv not infected	0.0953	0.0434	0.0469
none not infected-less not infected	-0.0111	0.028	0.6931
none not infected-more not infected	0.0639	0.0316	0.0644
none not infected-h37rv infected	0.0298	0.0423	0.5153
none not infected-less infected	-0.1181	0.0279	1e-04
none not infected-more infected	-0.0422	0.0311	0.219
h37rv not infected-less not infected	-0.1064	0.0418	0.0238
h37rv not infected-more not infected	-0.0313	0.0441	0.5153
less not infected-more not infected	0.075	0.0284	0.0207
h37rv infected-less infected	-0.1479	0.0406	0.001
h37rv infected-more infected	-0.0721	0.0426	0.1238
less infected-more infected	0.0759	0.0278	0.0196
h37rv not infected-h37rv infected	-0.0655	0.0264	0.0248
less not infected-less infected	-0.107	0.0123	0
more not infected-more infected	-0.1062	0.0176	0

b) Percentage of cells showing acidified phagolysosomes



Par	Estimate	se	OR	pvalue
(Intercept)	-3.5499	0.636	0.0287	0
grouph37rv not infected	1.1603	0.5574	3.1908	0.0374
groupless not infected	1.3347	0.4707	3.7989	0.0046
groupmore not infected	0.5412	0.6621	1.718	0.4137
grouph37rv infected	1.6871	0.5497	5.4039	0.0021
groupless infected	2.1921	0.4664	8.9544	0
groupmore infected	1.5756	0.5424	4.8338	0.0037

Contrast	estimate	SE	p.value
none not infected-h37rv not infected	-1.1603	0.5574	0.0934
none not infected-less not infected	-1.3347	0.4707	0.0137
none not infected-more not infected	-0.5412	0.6621	0.4773
none not infected-h37rv infected	-1.6871	0.5497	0.0107
none not infected-less infected	-2.1921	0.4664	0
none not infected-more infected	-1.5756	0.5424	0.0137
h37rv not infected-less not infected	-0.1744	0.3789	0.6913
h37rv not infected-more not infected	0.6191	0.6029	0.3806
less not infected-more not infected	0.7935	0.513	0.2032
h37rv infected-less infected	-0.505	0.3672	0.2305
h37rv infected-more infected	0.1115	0.4481	0.8036
less infected-more infected	0.6165	0.3323	0.1192
h37rv not infected-h37rv infected	-0.5268	0.36	0.2151
less not infected-less infected	-0.8574	0.1754	0
more not infected-more infected	-1.0345	0.525	0.1045

OBSERVATIONS

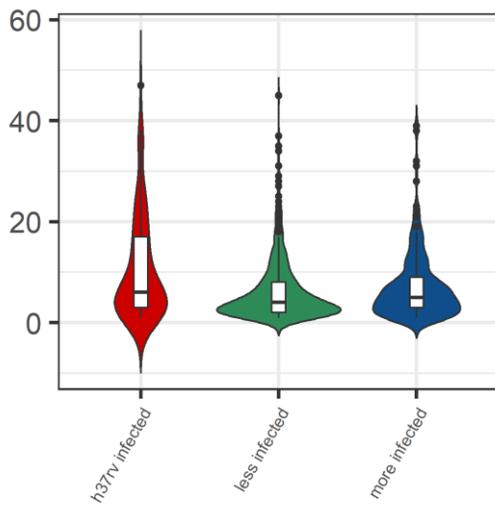
As the infection progresses, H37Rv and more virulent strains are blocking the acidification of the phagolysosomes. The percentage of acidified cells remained constant for less virulent strains and H37Rv, whereas it dropped below 10% for more virulent strains. A bystander effect in non-infected M2a macrophages was consistently observed for less virulent strains.

- Acidification of phagolysosomes observed
- ⊘ Acidification of phagolysosomes not observed (blocked)
- Acidification trend observed, statically not significant

Grouping: Virulence (relative)

M2 – 24 h p.i.

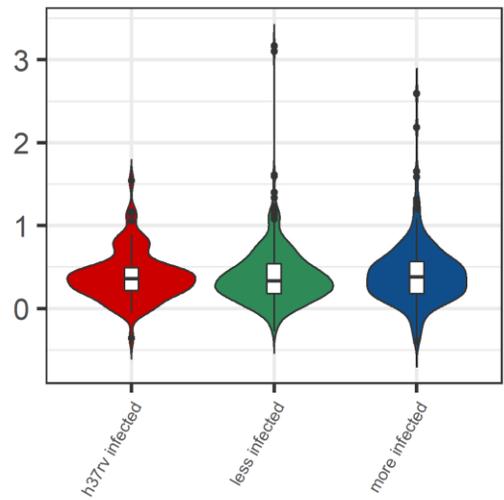
c) Effective MOI



Par	Value	Std.Error	p.value
(Intercept)	1.8249	0.1664	0
groupless infected	-0.3148	0.1656	0.062
groupmore infected	-0.2826	0.1734	0.1082

Contrast	estimate	SE	p.value
h37rv infected - less infected	0.3148	0.1656	0.1623
h37rv infected - more infected	0.2826	0.1734	0.1623
less infected - more infected	-0.0322	0.1146	0.7798

d) Colocalization Index



Par	Value	Std.Error	p.value
(Intercept)	-0.1339	0.1023	0.1911
groupless infected	0.0343	0.1102	0.7563
groupmore infected	0.1336	0.1187	0.2646

Contrast	estimate	SE	p.value
h37rv infected - less infected	-0.0343	0.1102	0.7563
h37rv infected - more infected	-0.1336	0.1187	0.397
less infected - more infected	-0.0993	0.0797	0.397

OBSERVATIONS

H37Rv showed higher MOI. Colocalization with acidified compartments was similar among all the groups.

Autophagy

CYTO-ID	
Wells/sample	2+2
Acquisition positions/well	2
Channels	Hoeschest, CYTO-ID, dsRed
Z-stacks	1.48 μm x4

	NOT infected		Infected		<i>TOT</i>	
	N of cells	Replicates	N of cells	Replicates	Not inf	Inf
M1	CTRL	121	5	<i>na</i>	<i>na</i>	
	RapaChlo	95	5	<i>na</i>	<i>na</i>	
	DMSO	67	5	<i>na</i>	<i>na</i>	
	H37Rv	48	3	80	3	
	H37Ra	49	2	51	2	609
	CDC1551					
	Beijing	114	5	141	5	
	EAI					
	Afri	115	5	131	5	
M2	CTRL	108	5	<i>na</i>	<i>na</i>	
	RapaChlo	87	5	<i>na</i>	<i>na</i>	
	DMSO	66	5	<i>na</i>	<i>na</i>	
	H37Rv	64	3	70	3	
	H37Ra	46	2	55	2	572
	CDC1551					
	Beijing	85	5	133	5	
	EAI					
	Afri	116	5	118	5	
<i>TOT</i>		<i>1181</i>		<i>779</i>		
						1960

Acquisition:

obj. 63x, oil NA 1.40

z-stacks: 1.48 μm x 4 stacks

Res. 16-bit format: 1024x1024

Laser 405 (for Hoechst) = 40%; gain: 935, offset: 0%

Laser 488 (for CYTO-ID) = 20% (Argon power: 30%); gain: 780, offset: 0%

Laser 453 (for dsRed) = 18%; gain: 905, offset: 0%

Confocal: AP 22 FD 6 O INT 0 FIM 100%

Hoechst (Ex/Em 350/461)

CYTO-ID (Ex/Em 499/548)

dsRed (Ex/Em 584-557)

Analysis:

24 h p.i.; N of autophagosomes, size of autophagosomes

CYTO-ID (Ex/Em 499/548)

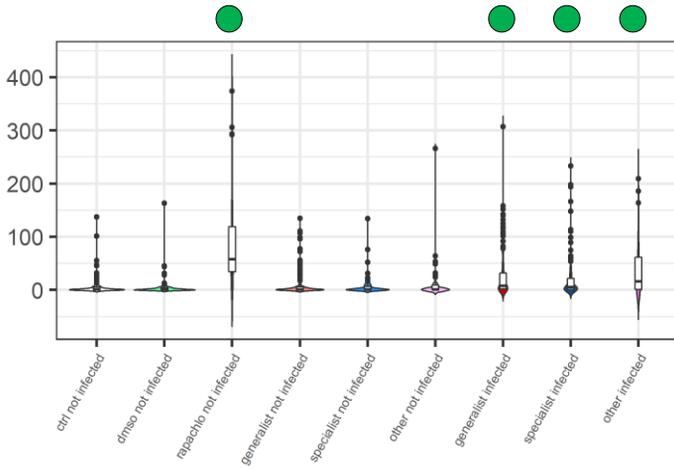
threshold (Default, Red; no flags on Dark background and Stack histogram) = 160

size = 5-infinity pixel units circularity = 0-1 (Show: nothing; Flags on Add to Manager, Display results)

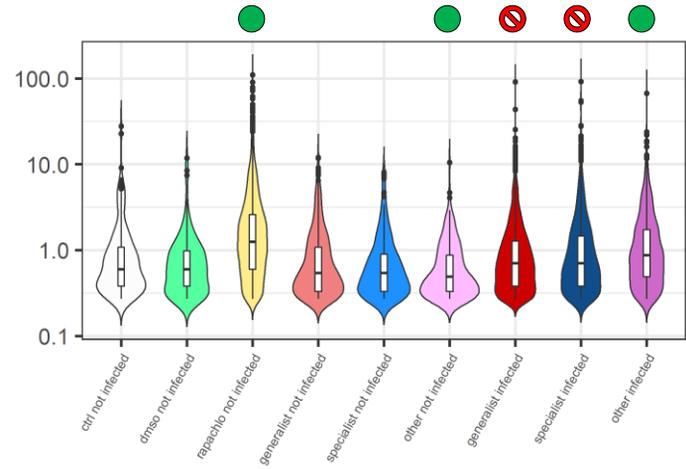
Grouping: Geo-host adaptation

M1 – 24 h.p.i.

a) Number of autophagosomes



b) Area of autophagosomes



Conditional model	Estimate	SE	p-value
(Intercept)	1.7344	0.3377	0
groupdms0 not infected	-0.0036	0.4416	0.9935
grouprapachlo not infected	2.4301	0.3888	0
groupgeneralist not infected	0.5395	0.3541	0.1276
groupspecialist not infected	0.0695	0.3828	0.8559
grouph0ther not infected	0.461	0.5017	0.3581
groupgeneralist infected	1.4475	0.3533	0
groupspecialist infected	1.1038	0.3813	0.0038
grouph0ther infected	1.4338	0.5005	0

Zero-inflation model	Estimate	SE	p-value
(Intercept)	-0.7759	0.379	0.0406
groupdms0 not infected	0.1688	0.5801	0.7711
grouprapachlo not infected	2.4906	0.6896	3e-04
groupgeneralist not infected	0.2177	0.4579	0.6345
groupspecialist not infected	0.3014	0.4855	0.5348
grouph0ther not infected	0.2938	0.6353	0.6438
groupgeneralist infected	-1.2833	0.468	0.0061
groupspecialist infected	-0.8256	0.4963	0.0962
grouph0ther infected	-0.7705	0.6669	0.2479

Contrast	estimate	SE	p.value
ctrl not infected-dms0 not infected	0.0036	0.4416	0.9935
ctrl not infected-rapachlo not infected	-2.4301	0.3888	0
ctrl not infected-generalist not infected	-0.5395	0.3541	0.2133
ctrl not infected-specialist not infected	-0.0695	0.3828	0.9305
ctrl not infected-other not infected	-0.461	0.5017	0.4886
ctrl not infected-generalist infected	-1.4475	0.3533	2e-04
ctrl not infected-specialist infected	-1.1038	0.3813	0.0089
ctrl not infected-other infected	-1.4338	0.5005	0.0091
dms0 not infected-rapachlo not infected	-2.4337	0.4259	0
dms0 not infected-generalist not infected	-0.5431	0.3959	0.2557
dms0 not infected-specialist not infected	-0.0731	0.4208	0.9305
dms0 not infected-other not infected	-0.4646	0.5279	0.4944
dms0 not infected-generalist infected	-1.4511	0.3952	7e-04
dms0 not infected-specialist infected	-1.1074	0.4194	0.0158
dms0 not infected-other infected	-1.4374	0.5268	0.0129
rapachlo not infected-generalist not infected	1.8905	0.3318	0
rapachlo not infected-specialist not infected	2.3605	0.3632	0
rapachlo not infected-other not infected	1.969	0.4894	2e-04
rapachlo not infected-generalist infected	0.9825	0.3309	0.0076
rapachlo not infected-specialist infected	1.3262	0.3616	7e-04
rapachlo not infected-other infected	0.9962	0.4882	0.0734
generalist not infected-specialist not infected	0.47	0.3216	0.2277
generalist not infected-other not infected	0.0785	0.4738	0.9305
specialist not infected-other not infected	-0.3915	0.4875	0.5276
generalist infected-specialist infected	0.3437	0.3188	0.4018
generalist infected-other infected	0.0137	0.4719	0.9935
specialist infected-other infected	-0.33	0.485	0.5957
generalist not infected-generalist infected	-0.908	0.0295	0
specialist not infected-specialist infected	-1.0343	0.0429	0
other not infected-other infected	-0.9728	0.0457	0

Par	Value	Std.Error	p.value
(Intercept)	-0.2579	0.1042	0.0133
groupdms0 not infected	-0.0571	0.1545	0.7136
grouprapachlo not infected	0.497	0.1119	1e-04
groupgeneralist not infected	-0.0992	0.1161	0.3931
groupspecialist not infected	-0.2413	0.1295	0.0625
grouph0ther not infected	-0.3661	0.1437	0.0108
groupgeneralist infected	0.0123	0.1082	0.9097
groupspecialist infected	0.0023	0.1169	0.9846
grouph0ther infected	0.3443	0.1269	0.0067

Contrast	estimate	SE	p.value
ctrl not infected-dms0 not infected	0.0571	0.1545	0.8234
ctrl not infected-rapachlo not infected	-0.497	0.1119	2e-04
ctrl not infected-generalist not infected	0.0992	0.1161	0.5127
ctrl not infected-specialist not infected	0.2413	0.1295	0.1041
ctrl not infected-other not infected	0.3661	0.1437	0.0232
ctrl not infected-generalist infected	-0.0123	0.1082	0.9411
ctrl not infected-specialist infected	-0.0023	0.1169	0.9846
ctrl not infected-other infected	-0.3443	0.1269	0.0166
dms0 not infected-rapachlo not infected	-0.5541	0.1356	6e-04
dms0 not infected-generalist not infected	0.0421	0.1386	0.8474
dms0 not infected-specialist not infected	0.1842	0.1501	0.3227
dms0 not infected-other not infected	0.309	0.1616	0.1041
dms0 not infected-generalist infected	-0.0693	0.132	0.7524
dms0 not infected-specialist infected	-0.0593	0.1394	0.8069
dms0 not infected-other infected	-0.4013	0.147	0.0206
rapachlo not infected-generalist not infected	0.5962	0.0864	0
rapachlo not infected-specialist not infected	0.7383	0.1038	0
rapachlo not infected-other not infected	0.8631	0.1226	0
rapachlo not infected-generalist infected	0.4848	0.0755	0
rapachlo not infected-specialist infected	0.4948	0.0873	0
rapachlo not infected-other infected	0.1527	0.1022	0.2241
generalist not infected-specialist not infected	0.1421	0.108	0.2823
generalist not infected-other not infected	0.2669	0.1199	0.0487
specialist not infected-other not infected	0.1248	0.1384	0.501
generalist infected-specialist infected	0.01	0.0822	0.9411
generalist infected-other infected	-0.332	0.0872	5e-04
specialist infected-other infected	-0.342	0.107	0.0038
generalist not infected-generalist infected	-0.1114	0.0487	0.0441
specialist not infected-specialist infected	-0.2435	0.0704	0.0016
other not infected-other infected	-0.7104	0.0768	0

OBSERVATIONS

In M1 macrophages, only the group "other" showed some degree of autophagy; indeed, whereas also generalist and specialist groups are showing an increase in the number of autophagosomes, their size does not show statistically increased size, thus suggesting poor maturation and progression in the autophagic flux.

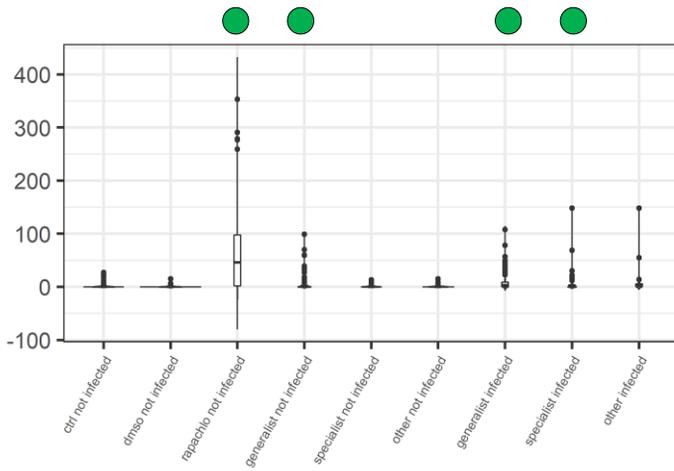
● Autophagosomes observed

⊘ Autophagosomes not observed (blocked)

Grouping: Geo-host adaptation

M2 – 24 h p.i.

a) Number of autophagosomes

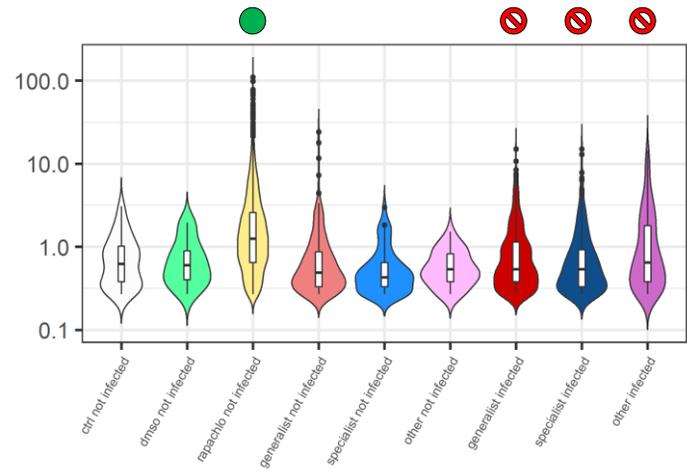


Conditional model	Estimate	SE	p-value
(Intercept)	0.903	0.345	0.0089
groupdms0 not infected	-0.4088	0.4775	0.3919
grouprapachlo not infected	2.9446	0.389	0
groupgeneralist not infected	0.9518	0.3649	0.0091
groupspecialist not infected	-0.2611	0.3975	0.5114
grouphother not infected	-0.4535	0.5153	0.3788
groupgeneralist infected	1.3009	0.3636	3e-04
groupspecialist infected	0.8508	0.3901	0.0292
grouphother infected	0.638	0.5016	0.2034

Zero-inflation model	Estimate	SE	p-value
(Intercept)	0.0905	0.3961	0.8193
groupdms0 not infected	0.6714	0.5867	0.2524
grouprapachlo not infected	-2.111	0.5777	3e-04
groupgeneralist not infected	-0.1886	0.4582	0.6806
groupspecialist not infected	-0.1775	0.4964	0.7207
grouphother not infected	-0.3559	0.6681	0.5942
groupgeneralist infected	-1.4032	0.4586	0.0022
groupspecialist infected	-0.663	0.4796	0.1669
grouphother infected	-1.3147	0.6501	0.0431

Contrast	estimate	SE	p-value
ctrl not infected-dms0 not infected	0.4088	0.4775	0.4705
ctrl not infected-rapachlo not infected	-2.9446	0.389	0
ctrl not infected-generalist not infected	-0.9518	0.3649	0.0154
ctrl not infected-specialist not infected	0.2611	0.3975	0.5902
ctrl not infected-other not infected	0.4535	0.5153	0.4705
ctrl not infected-generalist infected	-1.3009	0.3636	8e-04
ctrl not infected-specialist infected	-0.8508	0.3901	0.0465
ctrl not infected-other infected	-0.638	0.5016	0.2658
dms0 not infected-rapachlo not infected	-3.3534	0.4289	0
dms0 not infected-generalist not infected	-1.3606	0.4288	0.0031
dms0 not infected-specialist not infected	-0.1478	0.4584	0.7731
dms0 not infected-other not infected	0.0447	0.5682	0.9373
dms0 not infected-generalist infected	-1.7097	0.4277	3e-04
dms0 not infected-specialist infected	-1.2596	0.452	0.0096
dms0 not infected-other infected	-1.0468	0.5558	0.09
rapachlo not infected-generalist not infected	1.9928	0.3227	0
rapachlo not infected-specialist not infected	3.2057	0.3617	0
rapachlo not infected-other not infected	3.3981	0.4938	0
rapachlo not infected-generalist infected	1.6437	0.3211	0
rapachlo not infected-specialist infected	2.0938	0.3533	0
rapachlo not infected-other infected	2.3067	0.4796	0
generalist not infected-specialist not infected	1.2129	0.3292	6e-04
generalist not infected-other not infected	1.4053	0.484	0.0071
generalist infected-specialist not infected	0.1925	0.5027	0.752
generalist infected-specialist infected	0.4501	0.3182	0.2149
generalist infected-other not infected	0.6629	0.4685	0.2149
generalist infected-other infected	0.2128	0.4827	0.7326
specialist not infected-generalist not infected	-0.3491	0.046	0
specialist not infected-specialist not infected	-1.118	0.0942	0
specialist not infected-specialist infected	-1.0915	0.1381	0

b) Area of autophagosomes



Par	Value	Std.Error	p-value
(Intercept)	-0.5251	0.1368	1e-04
groupdms0 not infected	-0.0041	0.2487	0.987
grouprapachlo not infected	0.7515	0.1483	0
groupgeneralist not infected	-0.0939	0.1564	0.5485
groupspecialist not infected	-0.3087	0.1945	0.1125
grouphother not infected	-0.2501	0.2381	0.2936
groupgeneralist infected	0.0301	0.1472	0.8381
groupspecialist infected	-0.1507	0.162	0.3522
grouphother infected	0.1919	0.1829	0.2941

Contrast	estimate	SE	p-value
ctrl not infected-dms0 not infected	0.0041	0.2487	0.987
ctrl not infected-rapachlo not infected	-0.7515	0.1483	0
ctrl not infected-generalist not infected	0.0939	0.1564	0.6582
ctrl not infected-specialist not infected	0.3087	0.1945	0.2597
ctrl not infected-other not infected	0.2501	0.2381	0.4644
ctrl not infected-generalist infected	-0.0301	0.1472	0.8979
ctrl not infected-specialist infected	0.1507	0.162	0.5283
ctrl not infected-other infected	-0.1919	0.1829	0.4644
dms0 not infected-rapachlo not infected	-0.7555	0.2157	0.0017
dms0 not infected-generalist not infected	0.0898	0.222	0.7913
dms0 not infected-specialist not infected	0.3046	0.2501	0.4292
dms0 not infected-other not infected	0.246	0.2852	0.5548
dms0 not infected-generalist infected	-0.0341	0.2155	0.9043
dms0 not infected-specialist infected	0.1467	0.2257	0.6448
dms0 not infected-other infected	-0.1959	0.241	0.5675
rapachlo not infected-generalist not infected	0.8453	0.0968	0
rapachlo not infected-specialist not infected	1.0602	0.1506	0
rapachlo not infected-other not infected	1.0015	0.2039	0
rapachlo not infected-generalist infected	0.7214	0.0809	0
rapachlo not infected-specialist infected	0.9022	0.1052	0
rapachlo not infected-other infected	0.5596	0.1353	2e-04
generalist not infected-specialist not infected	0.2148	0.1589	0.3778
generalist not infected-other not infected	0.1562	0.2101	0.5964
specialist not infected-other not infected	-0.0586	0.2396	0.8964
generalist infected-specialist not infected	0.1808	0.1039	0.223
generalist infected-other not infected	-0.1618	0.1345	0.4292
specialist infected-other not infected	-0.3426	0.1501	0.0676
generalist not infected-generalist infected	-0.1239	0.0739	0.2344
specialist not infected-specialist infected	-0.158	0.1441	0.4644
other not infected-other infected	-0.442	0.193	0.0676

OBSERVATIONS

In M2 macrophages, despite a certain increase in the number of autophagosomes, their size does not show statistically increased size, thus suggesting poor maturation and progression in the autophagic flux. None of the group is inducing autophagy.

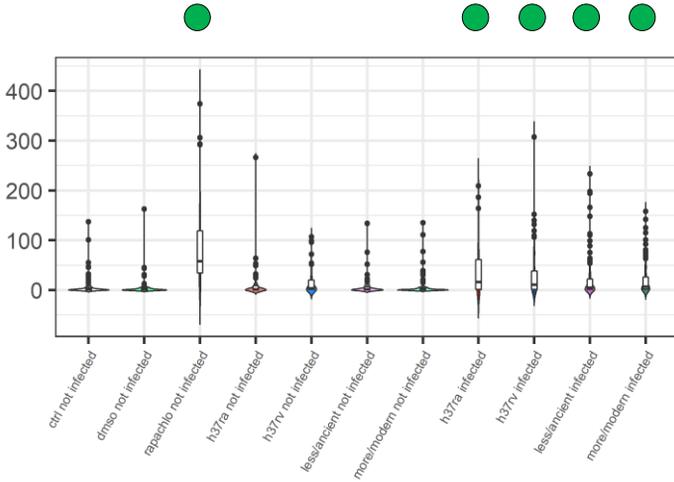
● Autophagosomes observed

⊘ Autophagosomes not observed (blocked)

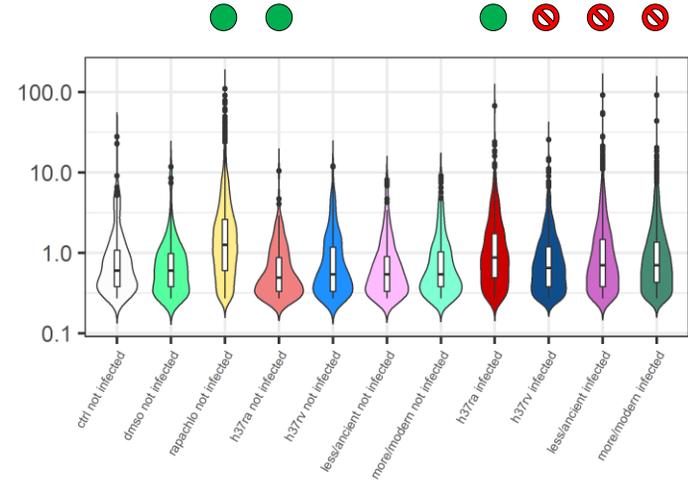
Grouping: Basic

M1 – 24 h.p.i.

a) Number of autophagosomes



b) Area of autophagosomes



Conditional model	Estimate	SE	p-value
(Intercept)	1.7318	0.3391	0
groupdmsmo not infected	-0.0094	0.4321	0.9826
grouprapachlo not infected	2.4283	0.3804	0
grouph37ra not infected	0.4201	0.492	0.3931
grouph37rv not infected	0.8728	0.4393	0.047
groupless/ancient not infected	0.072	0.3747	0.8476
groupmore/modern not infected	0.3621	0.3744	0.3336
grouph37ra infected	1.393	0.4908	0.0045
grouph37rv infected	1.7815	0.438	0
groupless/ancient infected	1.1064	0.3731	0.003
groupmore/modern infected	1.2688	0.3729	7e-04

Zero-inflation model	Estimate	SE	p-value
(Intercept)	-0.7554	0.3686	0.0404
groupdmsmo not infected	0.1755	0.5553	0.752
grouprapachlo not infected	-2.4925	0.6732	2e-04
grouph37ra not infected	0.323	0.6114	0.5973
grouph37rv not infected	-0.459	0.6239	0.462
groupless/ancient not infected	0.2864	0.4651	0.5381
groupmore/modern not infected	0.4171	0.4636	0.3683
grouph37ra infected	-0.7319	0.6441	0.2558
grouph37rv infected	-1.1875	0.5809	0.0409
groupless/ancient infected	-0.8359	0.4764	0.0794
groupmore/modern infected	-1.3416	0.4919	0.0064

Contrast	estimate	SE	p-value
ctrl not infected-dmsmo not infected	0.0094	0.4321	0.9826
ctrl not infected-rapachlo not infected	-2.4283	0.3804	0
ctrl not infected-h37ra not infected	-0.4201	0.492	0.5211
ctrl not infected-h37rv not infected	-0.8728	0.4393	0.0923
ctrl not infected-less/ancient not infected	-0.072	0.3747	0.889
ctrl not infected-more/modern not infected	-0.3621	0.3744	0.4949
ctrl not infected-h37ra infected	-1.393	0.4908	0.0111
ctrl not infected-h37rv infected	-1.7815	0.438	2e-04
ctrl not infected-less/ancient infected	-1.1064	0.3731	0.0078
ctrl not infected-more/modern infected	-1.2688	0.3729	0.0021
dmsmo not infected-rapachlo not infected	-2.4377	0.4167	0
dmsmo not infected-h37ra not infected	-0.4296	0.5176	0.5211
dmsmo not infected-h37rv not infected	-0.8822	0.4736	0.1125
dmsmo not infected-less/ancient not infected	-0.0815	0.4118	0.889
dmsmo not infected-more/modern not infected	-0.3715	0.4115	0.5211
dmsmo not infected-h37ra infected	-1.4024	0.5165	0.0145
dmsmo not infected-h37rv infected	-1.7909	0.4723	6e-04
dmsmo not infected-less/ancient infected	-1.1158	0.4104	0.0145
dmsmo not infected-more/modern infected	-1.2783	0.4101	0.0051
rapachlo not infected-h37ra not infected	2.0082	0.4803	2e-04
rapachlo not infected-h37rv not infected	1.5555	0.4204	8e-04
rapachlo not infected-less/ancient not infected	2.3563	0.3553	0
rapachlo not infected-more/modern not infected	2.0663	0.3548	0
rapachlo not infected-h37ra infected	1.0353	0.4791	0.0633
rapachlo not infected-h37rv infected	0.6468	0.419	0.2035
rapachlo not infected-less/ancient infected	1.3219	0.3536	7e-04
rapachlo not infected-more/modern infected	1.1595	0.3532	0.003
h37ra not infected-h37rv not infected	-0.4526	0.5515	0.5211
h37ra not infected-less/ancient not infected	0.3481	0.4787	0.5729
h37ra not infected-more/modern not infected	0.0581	0.4785	0.9249
h37rv not infected-less/ancient not infected	0.8007	0.4108	0.0964
h37rv not infected-more/modern not infected	0.5107	0.4104	0.328
less/ancient not infected-more/modern not infected	-0.29	0.3471	0.5211
h37ra infected-h37rv infected	-0.3885	0.5494	0.5729
h37ra infected-less/ancient infected	0.2866	0.4763	0.6362
h37ra infected-more/modern infected	0.1242	0.476	0.8757
h37rv infected-less/ancient infected	0.6751	0.4079	0.1689
h37rv infected-more/modern infected	0.5127	0.4075	0.328
less/ancient infected-more/modern infected	-0.1625	0.3437	0.7203

Par	Value	Std. Error	p-value
(Intercept)	-0.2574	0.1046	0.0138
groupdmsmo not infected	-0.0567	0.1556	0.717
grouprapachlo not infected	0.4966	0.1129	1e-04
grouph37ra not infected	-0.3725	0.1464	0.011
grouph37rv not infected	-0.0446	0.1376	0.7462
groupless/ancient not infected	-0.2415	0.1305	0.0642
groupmore/modern not infected	-0.172	0.1312	0.1898
grouph37ra infected	0.3378	0.1302	0.0095
grouph37rv infected	-0.006	0.1246	0.9618
groupless/ancient infected	0.0021	0.1179	0.986
groupmore/modern infected	0.0308	0.1161	0.7908

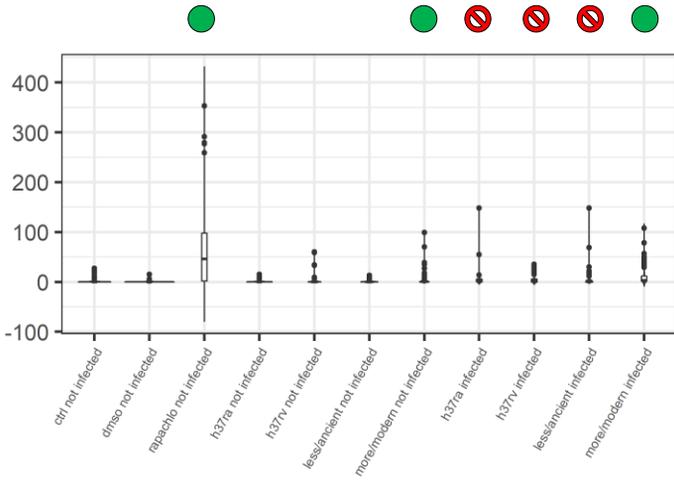
Contrast	estimate	SE	p-value
ctrl not infected-dmsmo not infected	0.0567	0.1556	0.855
ctrl not infected-rapachlo not infected	-0.4966	0.1129	3e-04
ctrl not infected-h37ra not infected	0.3725	0.1464	0.0276
ctrl not infected-h37rv not infected	0.0446	0.1376	0.855
ctrl not infected-less/ancient not infected	0.2415	0.1305	0.1315
ctrl not infected-more/modern not infected	0.172	0.1312	0.3265
ctrl not infected-h37ra infected	-0.3378	0.1302	0.0255
ctrl not infected-h37rv infected	-0.006	0.1246	0.9847
ctrl not infected-less/ancient infected	-0.0021	0.1179	0.986
ctrl not infected-more/modern infected	-0.0308	0.1161	0.872
dmsmo not infected-rapachlo not infected	-0.5533	0.1366	8e-04
dmsmo not infected-h37ra not infected	0.3157	0.1642	0.1305
dmsmo not infected-h37rv not infected	-0.0122	0.1571	0.9843
dmsmo not infected-less/ancient not infected	0.1847	0.151	0.3764
dmsmo not infected-more/modern not infected	0.1153	0.1516	0.6688
dmsmo not infected-h37ra infected	-0.3945	0.15	0.0276
dmsmo not infected-h37rv infected	-0.0508	0.1458	0.855
dmsmo not infected-less/ancient infected	-0.0588	0.1404	0.855
dmsmo not infected-more/modern infected	-0.0875	0.1387	0.7612
rapachlo not infected-h37ra not infected	0.8691	0.1255	0
rapachlo not infected-h37rv not infected	0.5411	0.1133	1e-04
rapachlo not infected-less/ancient not infected	0.738	0.1046	0
rapachlo not infected-more/modern not infected	0.6686	0.1056	0
rapachlo not infected-h37ra infected	0.1588	0.1059	0.2629
rapachlo not infected-h37rv infected	0.5026	0.0971	0
rapachlo not infected-less/ancient infected	0.4945	0.0884	0
rapachlo not infected-more/modern infected	0.4658	0.0861	0
h37ra not infected-h37rv not infected	-0.3279	0.1325	0.0302
h37ra not infected-less/ancient not infected	-0.131	0.1409	0.5416
h37ra not infected-more/modern not infected	-0.2005	0.1417	0.2813
h37rv not infected-less/ancient not infected	0.1969	0.1303	0.2558
h37rv not infected-more/modern not infected	0.1274	0.1301	0.5214
less/ancient not infected-more/modern not infected	-0.0695	0.124	0.7732
h37ra infected-h37rv infected	0.3438	0.0947	0.0011
h37ra infected-less/ancient infected	0.3357	0.1106	0.0079
h37rv infected-more/modern infected	0.307	0.1083	0.0141
less/ancient infected-more/modern infected	-0.008	0.1023	0.9843
less/ancient infected-less/ancient infected	-0.0368	0.0994	0.855
less/ancient infected-more/modern infected	-0.0287	0.0923	0.855
h37ra not infected-h37ra infected	-0.7103	0.0768	0
h37rv not infected-h37rv infected	-0.0386	0.0653	0.7693
less/ancient not infected-less/ancient infected	-0.2435	0.0704	0.002
more/modern not infected-more/modern infected	-0.2028	0.0729	0.0155

- Autophagosomes observed
- ⊘ Autophagosomes not observed (blocked)

Grouping: Basic

M2 – 24 h.p.i.

a) Number of autophagosomes

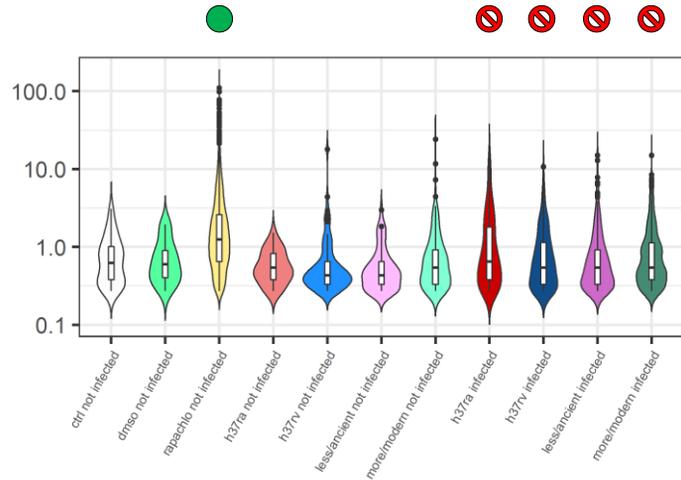


Conditional model	Estimate	SE	p-value
(Intercept)	0.9132	0.3393	0.0071
groupdmsmo not infected	-0.4106	0.4739	0.3863
grouprapapachlo not infected	2.9377	0.3858	0
grouph37ra not infected	-0.4356	0.5111	0.394
grouph37rv not infected	0.8393	0.4565	0.066
groupless/ancient not infected	-0.2701	0.3944	0.4934
groupmore/modern not infected	1.0085	0.3868	0.0091
grouph37ra infected	0.6558	0.4973	0.1873
grouph37rv infected	0.9766	0.4541	0.0315
groupless/ancient infected	0.8416	0.3869	0.0296
groupmore/modern infected	1.4501	0.3847	2e-04

Zero-inflation model	Estimate	SE	p-value
(Intercept)	0.092	0.3961	0.8163
groupdmsmo not infected	0.674	0.5865	0.2505
grouprapapachlo not infected	-2.1108	0.5782	5e-04
grouph37ra not infected	-0.3638	0.6681	0.5861
grouph37rv not infected	-0.0641	0.5851	0.9127
groupless/ancient not infected	-0.1785	0.4967	0.7193
groupmore/modern not infected	-0.2535	0.4915	0.606
grouph37ra infected	-1.3229	0.6502	0.0419
grouph37rv infected	-1.3892	0.5963	0.0198
groupless/ancient infected	-0.6637	0.4802	0.1669
groupmore/modern infected	-1.4196	0.4873	0.0036

Contrast	estimate	SE	p-value
ctrl not infected-dmsmo not infected	0.4106	0.4739	0.4843
ctrl not infected-rapapachlo not infected	-2.9377	0.3858	0
ctrl not infected-h37ra not infected	-0.4356	0.5111	0.4843
ctrl not infected-h37rv not infected	-0.8393	0.4565	0.1019
ctrl not infected-less/ancient not infected	0.2701	0.3944	0.5895
ctrl not infected-more/modern not infected	-1.0085	0.3868	0.0181
ctrl not infected-h37ra infected	-0.6558	0.4973	0.2521
ctrl not infected-h37rv infected	-0.9766	0.4541	0.0525
ctrl not infected-less/ancient infected	-0.8416	0.3869	0.0514
ctrl not infected-more/modern infected	-1.4501	0.3847	5e-04
dmsmo not infected-rapapachlo not infected	-3.3484	0.4261	0
dmsmo not infected-h37ra not infected	-0.025	0.5638	0.9647
dmsmo not infected-h37rv not infected	-1.2499	0.5062	0.0256
dmsmo not infected-less/ancient not infected	-0.1405	0.4552	0.7757
dmsmo not infected-more/modern not infected	-1.4192	0.4487	0.0041
dmsmo not infected-h37ra infected	-1.0664	0.5514	0.0851
dmsmo not infected-h37rv infected	-1.3873	0.504	0.013
dmsmo not infected-less/ancient infected	-1.2522	0.4487	0.0121
dmsmo not infected-more/modern infected	-1.8607	0.4468	0.0025
rapapachlo not infected-h37ra not infected	3.3734	0.4897	0
rapapachlo not infected-h37rv not infected	2.0985	0.4209	0
rapapachlo not infected-less/ancient not infected	3.2079	0.3588	0
rapapachlo not infected-more/modern not infected	1.9292	0.3498	0
rapapachlo not infected-h37ra infected	2.282	0.4754	0
rapapachlo not infected-h37rv infected	1.9611	0.4181	0
rapapachlo not infected-less/ancient infected	2.0962	0.3503	0
rapapachlo not infected-more/modern infected	1.4877	0.3474	1e-04
h37ra not infected-h37rv not infected	-1.2749	0.5642	0.0431
h37ra not infected-less/ancient not infected	-0.1655	0.4986	0.7757
h37ra not infected-more/modern not infected	-1.4441	0.4925	0.0082
h37rv not infected-less/ancient not infected	1.1094	0.4239	0.0181
h37rv not infected-more/modern not infected	-0.1693	0.4156	0.7694
less/ancient not infected-more/modern not infected	-1.2786	0.3573	0.001
h37ra infected-h37rv infected	-0.3208	0.5498	0.6504
h37ra infected-less/ancient infected	-0.1858	0.4784	0.7694
h37ra infected-more/modern infected	-0.7943	0.4766	0.1332
h37rv infected-less/ancient infected	0.135	0.4137	0.7757
h37rv infected-more/modern infected	-0.4734	0.4106	0.3247
less/ancient infected-more/modern infected	-0.6085	0.3463	0.1174

b) Area of autophagosomes



Par	Value	Std.Error	p-value
(Intercept)	-0.525	0.138	1e-04
groupdmsmo not infected	-0.0035	0.251	0.989
grouprapapachlo not infected	0.7501	0.151	0
grouph37ra not infected	-0.2585	0.2404	0.2823
grouph37rv not infected	-0.2842	0.1956	0.1464
groupless/ancient not infected	-0.3039	0.1967	0.1224
groupmore/modern not infected	-0.0075	0.1677	0.9645
grouph37ra infected	0.1817	0.1861	0.3288
grouph37rv infected	0.0488	0.1728	0.7775
groupless/ancient infected	-0.1504	0.1645	0.3606
groupmore/modern infected	0.0289	0.1543	0.8513

Contrast	estimate	SE	p-value
ctrl not infected-dmsmo not infected	0.0035	0.251	0.989
ctrl not infected-rapapachlo not infected	0.7501	0.151	0
ctrl not infected-h37ra not infected	0.2585	0.2404	0.4951
ctrl not infected-h37rv not infected	0.2842	0.1956	0.3325
ctrl not infected-less/ancient not infected	0.3039	0.1967	0.3096
ctrl not infected-more/modern not infected	0.0075	0.1677	0.989
ctrl not infected-h37ra infected	-0.1817	0.1861	0.5437
ctrl not infected-h37rv infected	-0.0488	0.1728	0.989
ctrl not infected-less/ancient infected	0.1504	0.1645	0.5743
ctrl not infected-more/modern infected	-0.0289	0.1543	0.989
dmsmo not infected-rapapachlo not infected	-0.7535	0.2179	0.0024
dmsmo not infected-h37ra not infected	0.255	0.2877	0.5764
dmsmo not infected-h37rv not infected	0.2807	0.2514	0.4951
dmsmo not infected-less/ancient not infected	0.3004	0.2523	0.4951
dmsmo not infected-more/modern not infected	0.004	0.2304	0.989
dmsmo not infected-h37ra infected	-0.1852	0.2441	0.6421
dmsmo not infected-h37rv infected	-0.0523	0.2341	0.989
dmsmo not infected-less/ancient infected	0.1469	0.228	0.7206
dmsmo not infected-more/modern infected	-0.0324	0.2208	0.989
rapapachlo not infected-h37ra not infected	1.0086	0.2063	0
rapapachlo not infected-h37rv not infected	1.0342	0.1517	0
rapapachlo not infected-less/ancient not infected	1.054	0.153	0
rapapachlo not infected-more/modern not infected	0.7575	0.1134	0
rapapachlo not infected-h37ra infected	0.5683	0.1391	2e-04
rapapachlo not infected-h37rv infected	0.7012	0.1208	0
rapapachlo not infected-less/ancient infected	0.9004	0.1085	0
rapapachlo not infected-more/modern infected	0.7211	0.0925	0
h37ra not infected-h37rv not infected	0.0257	0.2409	0.989
h37ra not infected-less/ancient not infected	0.0454	0.2417	0.989
h37ra not infected-more/modern not infected	-0.251	0.2188	0.4951
h37rv not infected-less/ancient not infected	0.0197	0.1972	0.989
h37rv not infected-more/modern not infected	-0.2767	0.1684	0.2876
less/ancient not infected-more/modern not infected	-0.2964	0.1696	0.2474
h37ra infected-h37rv infected	0.1329	0.1626	0.6135
h37ra infected-less/ancient infected	0.3321	0.1536	0.1015
h37ra infected-more/modern infected	0.1528	0.1428	0.4951
h37rv infected-less/ancient infected	0.1992	0.1373	0.3325
h37rv infected-more/modern infected	0.0199	0.125	0.989
less/ancient infected-more/modern infected	-0.1793	0.1132	0.3041
h37ra not infected-h37ra infected	-0.4402	0.1931	0.0813
h37rv not infected-h37rv infected	-0.333	0.1384	0.0631
less/ancient not infected-less/ancient infected	-0.1535	0.1444	0.4951
more/modern not infected-more/modern infected	-0.0364	0.088	0.9124

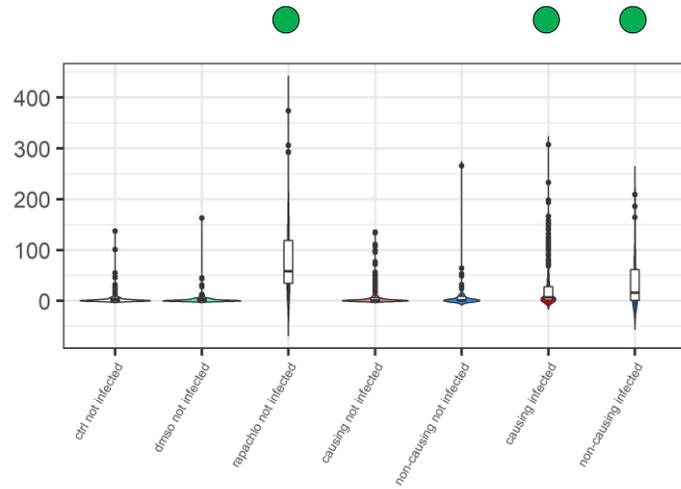
● Autophagosomes observed

⊘ Autophagosomes not observed (blocked)

Grouping: Disease

M1 – 24 h.p.i.

a) Number of autophagosomes

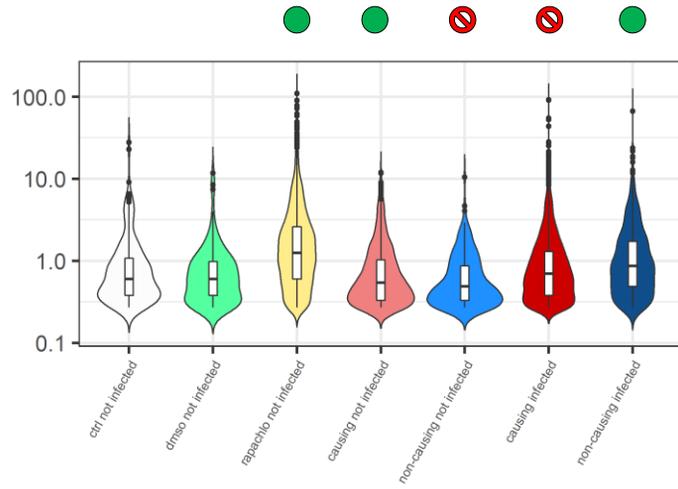


Conditional model	Estimate	SE	p-value
(Intercept)	1.7349	0.3388	0
groupdmsop not infected	-0.0012	0.4486	0.9978
grouprapachlo not infected	2.4316	0.3951	0
groupcausing not infected	0.3608	0.3353	0.282
groupnon-causing not infected	0.4802	0.5093	0.3458
groupcausing infected	1.3104	0.3347	1e-04
groupnon-causing infected	1.453	0.5081	0.0042

Zero-inflation model	Estimate	SE	p-value
(Intercept)	-0.7823	0.3792	0.0391
groupdmsop not infected	0.1665	0.5845	0.7757
grouprapachlo not infected	-2.4847	0.6921	3e-04
groupcausing not infected	0.2565	0.4255	0.5466
groupnon-causing not infected	0.2787	0.6387	0.6625
groupcausing infected	-1.0903	0.4317	0.0116
groupnon-causing infected	-0.7849	0.6697	0.2412

Contrast	estimate	SE	p-value
ctrl not infected-dmsop not infected	0.0012	0.4486	0.9978
ctrl not infected-rapachlo not infected	-2.4316	0.3951	0
ctrl not infected-causing not infected	-0.3608	0.3353	0.4125
ctrl not infected-non-causing not infected	-0.4802	0.5093	0.4383
ctrl not infected-causing infected	-1.3104	0.3347	3e-04
ctrl not infected-non-causing infected	-1.453	0.5081	0.0082
dmsop not infected-rapachlo not infected	-2.4329	0.4327	0
dmsop not infected-causing not infected	-0.362	0.3801	0.4383
dmsop not infected-non-causing not infected	-0.4814	0.536	0.4386
dmsop not infected-causing infected	-1.3116	0.3796	0.0012
dmsop not infected-non-causing infected	-1.4542	0.5349	0.0115
rapachlo not infected-causing not infected	2.0709	0.3116	0
rapachlo not infected-non-causing not infected	1.9515	0.4969	3e-04
rapachlo not infected-causing infected	1.1213	0.311	8e-04
rapachlo not infected-non-causing infected	0.9786	0.4957	0.077
causing not infected-non-causing not infected	-0.1194	0.4592	0.8391
causing infected-non-causing not infected	-0.1426	0.4575	0.8391
causing not infected-causing infected	-0.9496	0.0243	0
non-causing not infected-non-causing infected	-0.9728	0.0457	0

b) Area of autophagosomes



Par	Value	Std.Error	p-value
(Intercept)	-0.2578	0.1037	0.0129
groupdmsop not infected	-0.0574	0.154	0.7111
grouprapachlo not infected	0.4975	0.1114	1e-04
groupcausing not infected	-0.148	0.1091	0.1748
groupnon-causing not infected	-0.3676	0.1429	0.0101
groupcausing infected	0.0067	0.1038	0.9488
groupnon-causing infected	0.3425	0.1259	0.0065

Contrast	estimate	SE	p-value
ctrl not infected-dmsop not infected	0.0574	0.154	0.7507
ctrl not infected-rapachlo not infected	-0.4975	0.1114	2e-04
ctrl not infected-causing not infected	0.148	0.1091	0.2214
ctrl not infected-non-causing not infected	0.3676	0.1429	0.0174
ctrl not infected-causing infected	-0.0067	0.1038	0.9488
ctrl not infected-non-causing infected	-0.3425	0.1259	0.0138
dmsop not infected-rapachlo not infected	-0.5549	0.1351	4e-04
dmsop not infected-causing not infected	0.0906	0.1327	0.5916
dmsop not infected-non-causing not infected	0.3103	0.1609	0.0877
dmsop not infected-causing infected	-0.0641	0.1284	0.6932
dmsop not infected-non-causing infected	-0.3999	0.1462	0.0167
rapachlo not infected-causing not infected	0.6455	0.0769	0
rapachlo not infected-non-causing not infected	0.8651	0.1218	0
rapachlo not infected-causing infected	0.4908	0.0692	0
rapachlo not infected-non-causing infected	0.155	0.1012	0.1796
causing not infected-non-causing not infected	0.2196	0.1152	0.0877
causing infected-non-causing not infected	-0.3358	0.0852	3e-04
causing not infected-causing infected	-0.1547	0.04	3e-04
non-causing not infected-non-causing infected	-0.7101	0.0768	0

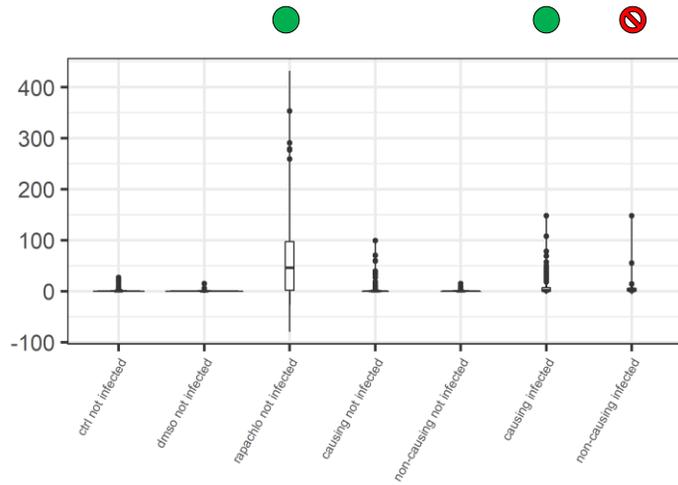
● Autophagosomes observed

⊘ Autophagosomes not observed (blocked)

Grouping: Disease

M2 – 24 h.p.i.

a) Number of autophagosomes

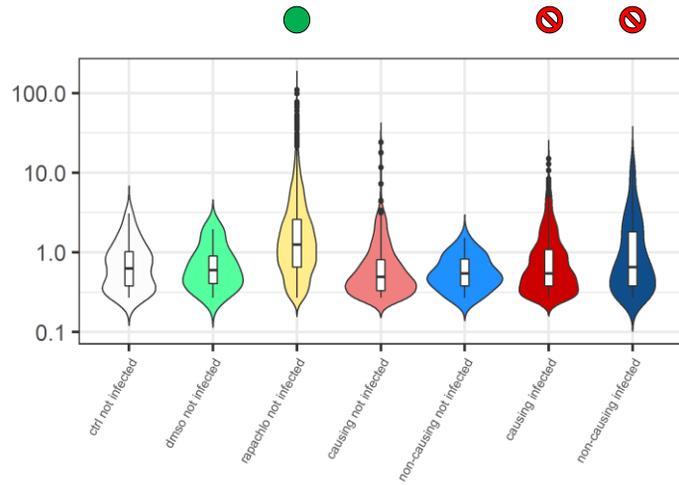


Conditional model	Estimate	SE	p-value
(Intercept)	0.9022	0.3512	0.0102
groupdms0 not infected	-0.4225	0.4959	0.3942
grouprapachlo not infected	2.9525	0.4065	0
groupcausing not infected	0.5802	0.3565	0.1036
groupnon-causing not infected	-0.4175	0.5371	0.437
groupcausing infected	1.1025	0.3554	0.0019
groupnon-causing infected	0.6735	0.524	0.1988

Zero-inflation model	Estimate	SE	p-value
(Intercept)	0.0662	0.4045	0.8701
groupdms0 not infected	0.6852	0.6036	0.2563
grouprapachlo not infected	-2.1235	0.5937	3e-04
groupcausing not infected	-0.0818	0.4352	0.8509
groupnon-causing not infected	-0.3562	0.6839	0.6025
groupcausing infected	-1.0945	0.4354	0.012
groupnon-causing infected	-1.3206	0.6667	0.0476

Contrast	estimate	SE	p-value
ctrl not infected-dms0 not infected	0.4225	0.4959	0.4408
ctrl not infected-rapachlo not infected	-2.9525	0.4065	0
ctrl not infected-causing not infected	-0.5802	0.3565	0.141
ctrl not infected-non-causing not infected	0.4175	0.5371	0.4615
ctrl not infected-causing infected	-1.1025	0.3554	0.0038
ctrl not infected-non-causing infected	-0.6735	0.524	0.2522
dms0 not infected-rapachlo not infected	-3.375	0.4437	0
dms0 not infected-causing not infected	-1.0027	0.4246	0.0318
dms0 not infected-non-causing not infected	-0.005	0.5899	0.9933
dms0 not infected-causing infected	-1.525	0.4236	7e-04
dms0 not infected-non-causing infected	-1.096	0.5781	0.0852
rapachlo not infected-causing not infected	2.3723	0.3112	0
rapachlo not infected-non-causing not infected	3.37	0.5146	0
rapachlo not infected-causing infected	1.85	0.3099	0
rapachlo not infected-non-causing infected	2.2791	0.501	0
causing not infected-non-causing not infected	0.9977	0.4825	0.0616
causing infected-non-causing infected	0.429	0.4672	0.4259
causing not infected-causing infected	-0.5222	0.041	0
non-causing not infected-non-causing infected	-1.091	0.1381	0

b) Area of autophagosomes



Par	Value	Std.Error	p-value
(Intercept)	-0.5249	0.1379	1e-04
groupdms0 not infected	-0.0036	0.251	0.9885
grouprapachlo not infected	0.7501	0.1509	0
groupcausing not infected	-0.1532	0.1538	0.3192
groupnon-causing not infected	-0.2587	0.2404	0.2819
groupcausing infected	-0.0253	0.1461	0.8624
groupnon-causing infected	0.1814	0.1859	0.3292

Contrast	estimate	SE	p-value
ctrl not infected-dms0 not infected	0.0036	0.251	0.9885
ctrl not infected-rapachlo not infected	-0.7501	0.1509	0
ctrl not infected-causing not infected	0.1532	0.1538	0.5212
ctrl not infected-non-causing not infected	0.2587	0.2404	0.5212
ctrl not infected-causing infected	0.0253	0.1461	0.9639
ctrl not infected-non-causing infected	-0.1814	0.1859	0.5212
dms0 not infected-rapachlo not infected	-0.7537	0.2179	0.0017
dms0 not infected-causing not infected	0.1496	0.2205	0.6301
dms0 not infected-non-causing not infected	0.2551	0.2877	0.5485
dms0 not infected-causing infected	0.0217	0.2152	0.9707
dms0 not infected-non-causing infected	-0.1851	0.244	0.6083
rapachlo not infected-causing not infected	0.9034	0.0916	0
rapachlo not infected-non-causing not infected	1.0088	0.2062	0
rapachlo not infected-causing infected	0.7755	0.078	0
rapachlo not infected-non-causing infected	0.5687	0.1389	2e-04
causing not infected-non-causing not infected	0.1054	0.2083	0.7278
causing infected-non-causing infected	-0.2068	0.1337	0.2579
causing not infected-causing infected	-0.1279	0.066	0.125
non-causing not infected-non-causing infected	-0.4401	0.1932	0.0618

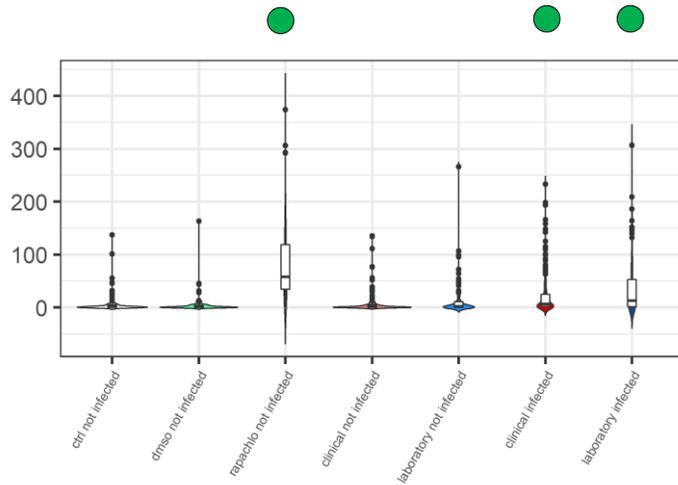
● Autophagosomes observed

⊘ Autophagosomes not observed (blocked)

Grouping: Laboratory adaptation

M1 – 24 h.p.i.

a) Number of autophagosomes

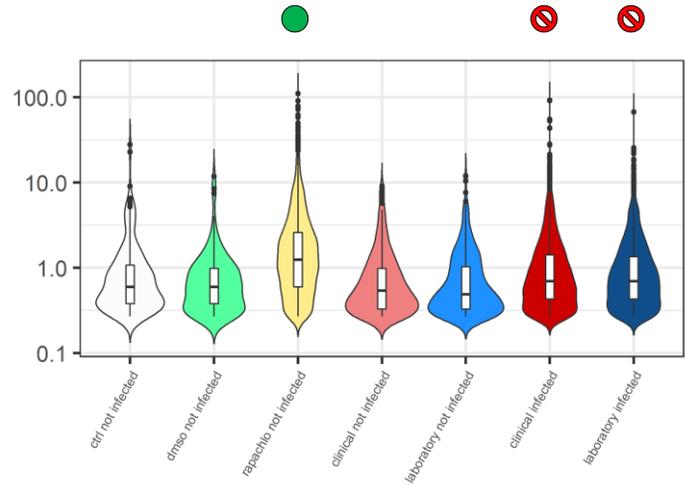


Conditional model	Estimate	SE	p-value
(Intercept)	1.7348	0.3358	0
groupdms0 not infected	-0.0045	0.4386	0.9918
grouprapachlo not infected	2.4297	0.386	0
groupclinical not infected	0.2143	0.3366	0.5243
grouplaboratory not infected	0.6847	0.379	0.0708
groupclinical infected	1.1845	0.3357	4e-04
grouplaboratory infected	1.6229	0.3782	0

Zero-inflation model	Estimate	SE	p-value
(Intercept)	-0.7799	0.3708	0.0354
groupdms0 not infected	0.1674	0.5769	0.7717
grouprapachlo not infected	-2.4772	0.6863	3e-04
groupclinical not infected	0.3778	0.427	0.3763
grouplaboratory not infected	-0.0615	0.5003	0.9021
groupclinical infected	-1.0595	0.4375	0.0154
grouplaboratory infected	-0.9966	0.5011	0.0467

Contrast	estimate	SE	p-value
ctrl not infected-dms0 not infected	0.0045	0.4386	0.9918
ctrl not infected-rapachlo not infected	-2.4297	0.386	0
ctrl not infected-clinical not infected	-0.2143	0.3366	0.5861
ctrl not infected-laboratory not infected	-0.6847	0.379	0.1039
ctrl not infected-clinical infected	-1.1845	0.3357	8e-04
ctrl not infected-laboratory infected	-1.6229	0.3782	1e-04
dms0 not infected-rapachlo not infected	-2.4342	0.4228	0
dms0 not infected-clinical not infected	-0.2188	0.3785	0.5946
dms0 not infected-laboratory not infected	-0.6893	0.4166	0.1335
dms0 not infected-clinical infected	-1.1891	0.3778	0.0029
dms0 not infected-laboratory infected	-1.6274	0.4159	2e-04
rapachlo not infected-clinical not infected	2.2154	0.3143	0
rapachlo not infected-laboratory not infected	1.7449	0.3593	0
rapachlo not infected-clinical infected	1.2452	0.3134	2e-04
rapachlo not infected-laboratory infected	0.8068	0.3585	0.039
clinical not infected-laboratory not infected	-0.4704	0.3039	0.1544
clinical infected-laboratory infected	-0.4383	0.3019	0.1744
clinical not infected-clinical infected	-0.9702	0.0299	0
laboratory not infected-laboratory infected	-0.9381	0.0308	0

b) Area of autophagosomes



Par	Value	Std.Error	p-value
(Intercept)	-0.2652	0.1037	0.0105
groupdms0 not infected	-0.0596	0.1533	0.6991
grouprapachlo not infected	0.503	0.1106	0
groupclinical not infected	-0.199	0.1133	0.0791
grouplaboratory not infected	-0.1809	0.1231	0.1416
groupclinical infected	0.0245	0.1055	0.8161
grouplaboratory infected	0.1363	0.1156	0.2384

Contrast	estimate	SE	p-value
ctrl not infected-dms0 not infected	0.0596	0.1533	0.7814
ctrl not infected-rapachlo not infected	-0.503	0.1106	1e-04
ctrl not infected-clinical not infected	0.199	0.1133	0.167
ctrl not infected-laboratory not infected	0.1809	0.1231	0.2556
ctrl not infected-clinical infected	-0.0245	0.1055	0.8545
ctrl not infected-laboratory infected	-0.1363	0.1156	0.3484
dms0 not infected-rapachlo not infected	-0.5826	0.1344	3e-04
dms0 not infected-clinical not infected	0.1394	0.1362	0.4228
dms0 not infected-laboratory not infected	0.1213	0.1439	0.5116
dms0 not infected-clinical infected	-0.0842	0.1298	0.6173
dms0 not infected-laboratory infected	-0.1959	0.1377	0.2556
rapachlo not infected-clinical not infected	0.702	0.0831	0
rapachlo not infected-laboratory not infected	0.6839	0.0967	0
rapachlo not infected-clinical infected	0.4784	0.072	0
rapachlo not infected-laboratory infected	0.3667	0.0869	3e-04
clinical not infected-laboratory not infected	-0.0181	0.0987	0.8545
clinical infected-laboratory infected	-0.1118	0.0791	0.2556
clinical not infected-clinical infected	-0.2235	0.0508	0
laboratory not infected-laboratory infected	-0.3172	0.0499	0

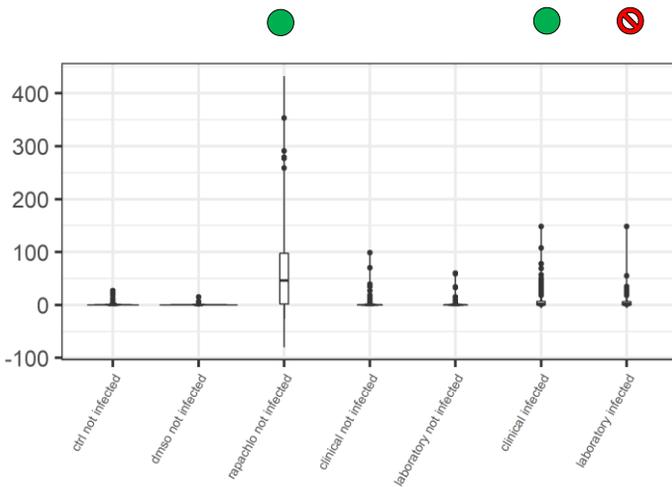
● Autophagosomes observed

⊘ Autophagosomes not observed (blocked)

Grouping: Laboratory adaptation

M2 – 24 h.p.i.

a) Number of autophagosomes

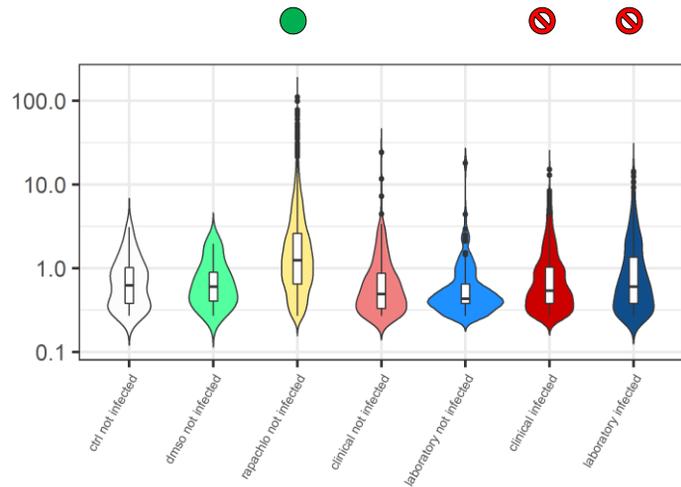


Conditional model	Estimate	SE	p-value
(Intercept)	0.914	0.3485	0.0087
groupdmsmo not infected	-0.4316	0.5008	0.3889
grouprapachlo not infected	2.9482	0.4113	0
groupclinical not infected	0.4813	0.3687	0.1918
grouplaboratory not infected	0.4107	0.4155	0.323
groupclinical infected	1.1228	0.3671	0.0022
grouplaboratory infected	0.8507	0.4128	0.0393

Zero-inflation model	Estimate	SE	p-value
(Intercept)	0.068	0.4064	0.8671
groupdmsmo not infected	0.6889	0.6069	0.2563
grouprapachlo not infected	-2.1348	0.5971	3e-04
groupclinical not infected	-0.1002	0.4465	0.8224
grouplaboratory not infected	-0.0604	0.5069	0.9052
groupclinical infected	-1.0259	0.4446	0.021
grouplaboratory infected	-1.3583	0.5156	0.0084

Contrast	estimate	SE	p-value
ctrl not infected-dmsmo not infected	0.4316	0.5008	0.421
ctrl not infected-rapachlo not infected	-2.9482	0.4113	0
ctrl not infected-clinical not infected	-0.4813	0.3687	0.2434
ctrl not infected-laboratory not infected	-0.4107	0.4155	0.3839
ctrl not infected-clinical infected	-1.1228	0.3671	0.0043
ctrl not infected-laboratory infected	-0.8507	0.4128	0.0579
dmsmo not infected-rapachlo not infected	-3.3798	0.4478	0
dmsmo not infected-clinical not infected	-0.9129	0.4366	0.0579
dmsmo not infected-laboratory not infected	-0.8422	0.4766	0.1052
dmsmo not infected-clinical infected	-1.5543	0.4352	8e-04
dmsmo not infected-laboratory infected	-1.2822	0.4742	0.0121
rapachlo not infected-clinical not infected	2.4669	0.3263	0
rapachlo not infected-laboratory not infected	2.5376	0.3781	0
rapachlo not infected-clinical infected	1.8255	0.3243	0
rapachlo not infected-laboratory infected	2.0976	0.3749	0
clinical not infected-laboratory not infected	0.0706	0.328	0.8296
clinical infected-laboratory infected	0.2721	0.3224	0.421
clinical not infected-clinical infected	-0.6415	0.0478	0
laboratory not infected-laboratory infected	-0.44	0.0682	0

b) Area of autophagosomes



Par	Value	Std.Error	p-value
(Intercept)	-0.5257	0.1401	2e-04
groupdmsmo not infected	-0.0016	0.2538	0.9948
grouprapachlo not infected	0.7482	0.1542	0
groupclinical not infected	-0.1113	0.1612	0.4898
grouplaboratory not infected	-0.273	0.1815	0.1325
groupclinical infected	-0.0453	0.151	0.7641
grouplaboratory infected	0.1032	0.1626	0.5253

Contrast	estimate	SE	p-value
ctrl not infected-dmsmo not infected	0.0016	0.2538	0.9948
ctrl not infected-rapachlo not infected	-0.7482	0.1542	0
ctrl not infected-clinical not infected	0.1113	0.1612	0.713
ctrl not infected-laboratory not infected	0.273	0.1815	0.2894
ctrl not infected-clinical infected	0.0453	0.151	0.854
ctrl not infected-laboratory infected	-0.1032	0.1626	0.713
dmsmo not infected-rapachlo not infected	-0.7499	0.2206	0.0022
dmsmo not infected-clinical not infected	0.1097	0.2262	0.765
dmsmo not infected-laboratory not infected	0.2714	0.241	0.4496
dmsmo not infected-clinical infected	0.0437	0.2191	0.8887
dmsmo not infected-laboratory infected	-0.1049	0.2271	0.765
rapachlo not infected-clinical not infected	0.8595	0.1025	0
rapachlo not infected-laboratory not infected	1.0212	0.1321	0
rapachlo not infected-clinical infected	0.7935	0.0856	0
rapachlo not infected-laboratory infected	0.645	0.1046	0
clinical not infected-laboratory not infected	0.1617	0.1402	0.4496
clinical infected-laboratory infected	-0.1486	0.0999	0.2894
clinical not infected-clinical infected	-0.066	0.0754	0.6035
laboratory not infected-laboratory infected	-0.3763	0.1126	0.0023

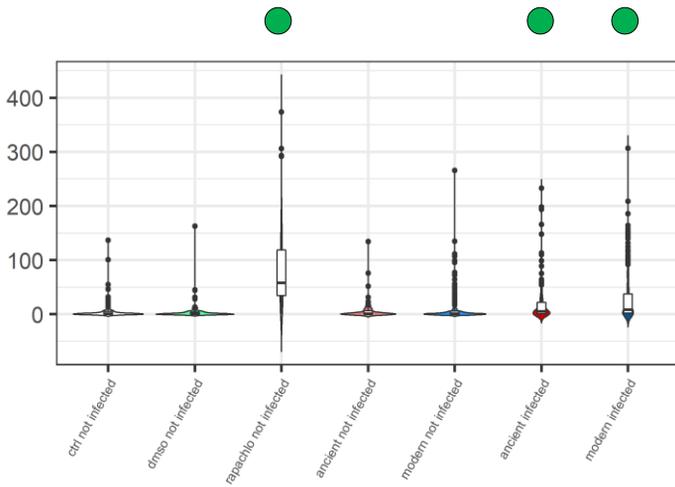
● Autophagosomes observed

⊘ Autophagosomes not observed (blocked)

Grouping: Phylogeny

M1 – 24 h p.i.

a) Number of autophagosomes

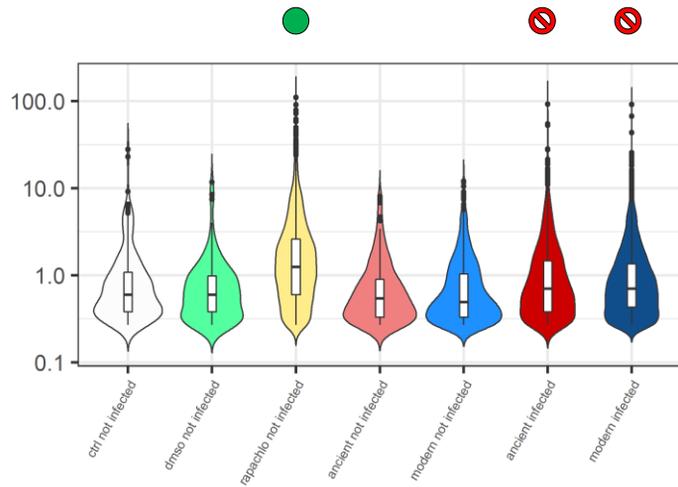


Conditional model	Estimate	SE	p-value
(Intercept)	1.7337	0.3375	0
groupdms0 not infected	-0.0032	0.4421	0.9943
grouprapachlo not infected	2.431	0.3892	0
groupancient not infected	0.0702	0.3831	0.8547
groupmodern not infected	0.5187	0.339	0.1259
groupancient infected	1.1045	0.3816	0.0038
groupmodern infected	1.4461	0.3384	0

Zero-inflation model	Estimate	SE	p-value
(Intercept)	-0.7825	0.3777	0.0383
groupdms0 not infected	0.1651	0.585	0.7778
grouprapachlo not infected	-2.4843	0.6922	3e-04
groupancient not infected	0.3061	0.4891	0.5315
groupmodern not infected	0.233	0.4371	0.594
groupancient infected	-0.8203	0.4998	0.1007
groupmodern infected	-1.1715	0.4466	0.0087

Contrast	estimate	SE	p-value
ctrl not infected-dms0 not infected	0.0032	0.4421	0.9943
ctrl not infected-rapachlo not infected	-2.431	0.3892	0
ctrl not infected-ancient not infected	-0.0702	0.3831	0.9097
ctrl not infected-modern not infected	-0.5187	0.339	0.1845
ctrl not infected-ancient infected	-1.1045	0.3816	0.0067
ctrl not infected-modern infected	-1.4461	0.3384	1e-04
dms0 not infected-rapachlo not infected	-2.4341	0.4263	0
dms0 not infected-ancient not infected	-0.0733	0.4211	0.9097
dms0 not infected-modern not infected	-0.5219	0.3813	0.2171
dms0 not infected-ancient infected	-1.1076	0.4197	0.0134
dms0 not infected-modern infected	-1.4492	0.3808	4e-04
rapachlo not infected-ancient not infected	2.3608	0.3636	0
rapachlo not infected-modern not infected	1.9122	0.3166	0
rapachlo not infected-ancient infected	1.3265	0.3619	5e-04
rapachlo not infected-modern infected	0.9849	0.3159	0.0036
ancient not infected-modern not infected	-0.4486	0.3074	0.1966
ancient infected-modern infected	-0.3416	0.3048	0.312
ancient not infected-ancient infected	-1.0343	0.0429	0
modern not infected-modern infected	-0.9273	0.0248	0

b) Area of autophagosomes



Par	Value	Std.Error	p-value
(Intercept)	-0.2654	0.1031	0.0101
groupdms0 not infected	-0.0604	0.1533	0.6955
grouprapachlo not infected	0.5039	0.1106	0
groupancient not infected	-0.2355	0.1285	0.0669
groupmodern not infected	-0.192	0.1111	0.0839
groupancient infected	0.0086	0.1156	0.9405
groupmodern infected	0.0885	0.1055	0.4016

Contrast	estimate	SE	p-value
ctrl not infected-dms0 not infected	0.0604	0.1533	0.7342
ctrl not infected-rapachlo not infected	-0.5039	0.1106	1e-04
ctrl not infected-ancient not infected	0.2355	0.1285	0.1412
ctrl not infected-modern not infected	0.192	0.1111	0.1594
ctrl not infected-ancient infected	-0.0086	0.1156	0.9405
ctrl not infected-modern infected	-0.0885	0.1055	0.5086
dms0 not infected-rapachlo not infected	-0.5643	0.1344	3e-04
dms0 not infected-ancient not infected	0.1751	0.1491	0.4057
dms0 not infected-modern not infected	0.1316	0.1341	0.4498
dms0 not infected-ancient infected	-0.069	0.1382	0.7342
dms0 not infected-modern infected	-0.1489	0.1295	0.4057
rapachlo not infected-ancient not infected	0.7394	0.1028	0
rapachlo not infected-modern not infected	0.6959	0.0805	0
rapachlo not infected-ancient infected	0.4953	0.0861	0
rapachlo not infected-modern infected	0.4154	0.0726	0
ancient not infected-modern not infected	-0.0435	0.1033	0.7342
ancient infected-modern infected	-0.0799	0.0794	0.4498
ancient not infected-ancient infected	-0.2441	0.0706	0.0013
modern not infected-modern infected	-0.2805	0.0412	0

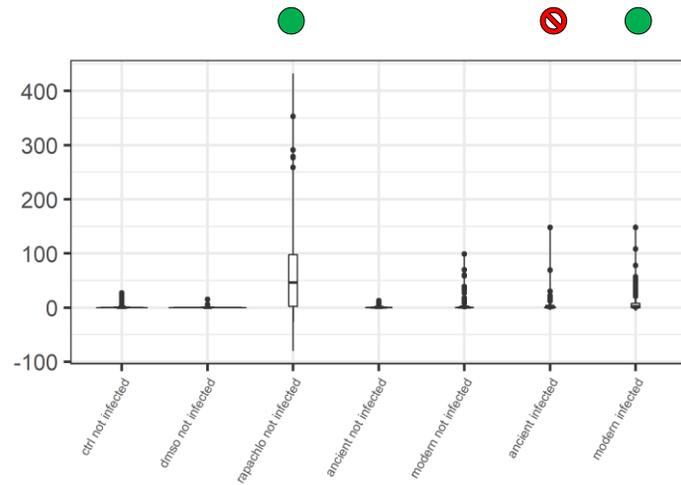
● Autophagosomes observed

⊘ Autophagosomes not observed (blocked)

Grouping: Phylogeny

M2 – 24 h.p.i.

a) Number of autophagosomes

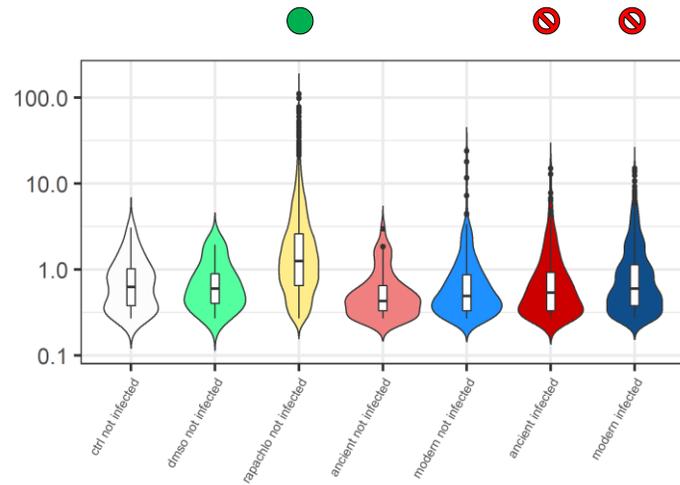


Conditional model	Estimate	SE	p-value
(Intercept)	0.9155	0.3453	0.008
groupdms0 not infected	-0.4274	0.4938	0.3868
grouprapachlo not infected	2.9445	0.4047	0
groupancient not infected	-0.2741	0.4129	0.5069
groupmodern not infected	0.7098	0.3628	0.0504
groupancient infected	0.8379	0.4057	0.0389
groupmodern infected	1.1514	0.3615	0.0014

Zero-inflation model	Estimate	SE	p-value
(Intercept)	0.0742	0.3978	0.852
groupdms0 not infected	0.6855	0.5952	0.2495
grouprapachlo not infected	-2.1116	0.5851	3e-04
groupancient not infected	-0.1643	0.5037	0.7443
groupmodern not infected	-0.1464	0.4402	0.7395
groupancient infected	-0.6477	0.4867	0.1833
groupmodern infected	-1.3785	0.4416	0.0018

Contrast	estimate	SE	p-value
ctrl not infected-dms0 not infected	0.4274	0.4938	0.4326
ctrl not infected-rapachlo not infected	-2.9445	0.4047	0
ctrl not infected-ancient not infected	0.2741	0.4129	0.5352
ctrl not infected-modern not infected	-0.7098	0.3628	0.0642
ctrl not infected-ancient infected	-0.8379	0.4057	0.0532
ctrl not infected-modern infected	-1.1514	0.3615	0.0028
dms0 not infected-rapachlo not infected	-3.3719	0.4423	0
dms0 not infected-ancient not infected	-0.1533	0.4738	0.7464
dms0 not infected-modern not infected	-1.1372	0.4306	0.0123
dms0 not infected-ancient infected	-1.2652	0.4675	0.011
dms0 not infected-modern infected	-1.5787	0.4296	5e-04
rapachlo not infected-ancient not infected	3.2186	0.3768	0
rapachlo not infected-modern not infected	2.2347	0.3204	0
rapachlo not infected-ancient infected	2.1066	0.3687	0
rapachlo not infected-modern infected	1.7931	0.3189	0
ancient not infected-modern not infected	-0.9839	0.3283	0.0048
ancient infected-modern infected	-0.3135	0.3174	0.3842
ancient not infected-ancient infected	-1.112	0.0942	0
modern not infected-modern infected	-0.4416	0.0432	0

b) Area of autophagosomes



Par	Value	Std.Error	p-value
(Intercept)	-0.5245	0.1368	1e-04
groupdms0 not infected	-0.0046	0.2495	0.9852
grouprapachlo not infected	0.7512	0.1492	0
groupancient not infected	-0.305	0.1952	0.1183
groupmodern not infected	-0.1101	0.1546	0.4762
groupancient infected	-0.1494	0.1627	0.3586
groupmodern infected	0.0554	0.1461	0.7044

Contrast	estimate	SE	p-value
ctrl not infected-dms0 not infected	0.0046	0.2495	0.9852
ctrl not infected-rapachlo not infected	-0.7512	0.1492	0
ctrl not infected-ancient not infected	0.305	0.1952	0.2498
ctrl not infected-modern not infected	0.1101	0.1546	0.6463
ctrl not infected-ancient infected	0.1494	0.1627	0.5241
ctrl not infected-modern infected	-0.0554	0.1461	0.7872
dms0 not infected-rapachlo not infected	-0.7558	0.2165	0.0015
dms0 not infected-ancient not infected	0.3003	0.2509	0.3997
dms0 not infected-modern not infected	0.1055	0.2208	0.7514
dms0 not infected-ancient infected	0.1448	0.2266	0.6622
dms0 not infected-modern infected	-0.06	0.2149	0.8233
rapachlo not infected-ancient not infected	1.0561	0.1513	0
rapachlo not infected-modern not infected	0.8613	0.0935	0
rapachlo not infected-ancient infected	0.9006	0.1064	0
rapachlo not infected-modern infected	0.6958	0.0786	0
ancient not infected-modern not infected	-0.1948	0.1568	0.3997
ancient infected-modern infected	-0.2048	0.102	0.106
ancient not infected-ancient infected	-0.1555	0.1442	0.4447
modern not infected-modern infected	-0.1655	0.0691	0.0451

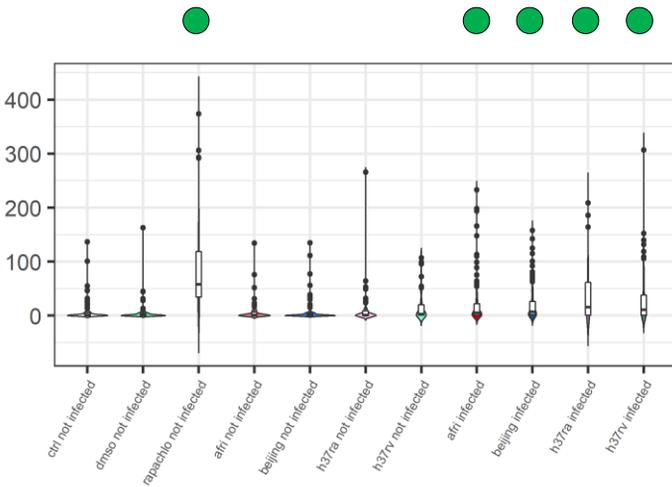
● Autophagosomes observed

⊘ Autophagosomes not observed (blocked)

Grouping: Sample

M1 – 24 h.p.i.

a) Number of autophagosomes

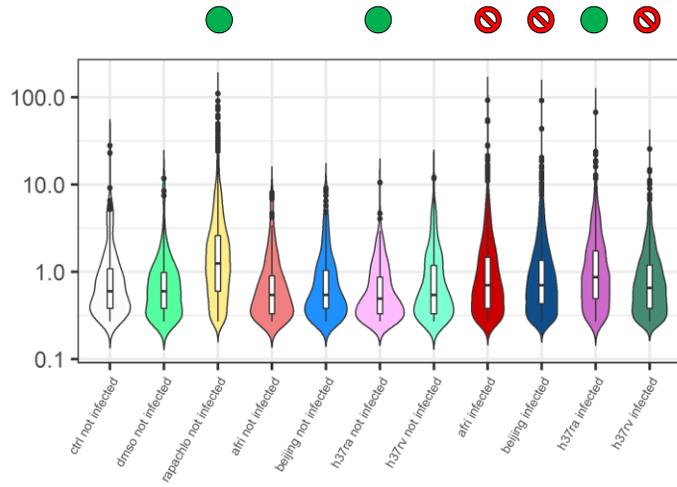


Conditional model	Estimate	SE	p-value
(Intercept)	1.7318	0.3391	0
groupdms0 not infected	-0.0094	0.4321	0.9826
grouprapachlo not infected	2.4283	0.3804	0
groupafri not infected	0.072	0.3747	0.8476
groupbeijing not infected	0.3621	0.3744	0.3336
group37ra not infected	0.4201	0.492	0.3931
group37rv not infected	0.8728	0.4393	0.047
groupafri infected	1.1064	0.3731	0.003
groupbeijing infected	1.2688	0.3729	7e-04
group37ra infected	1.393	0.4908	0.0045
group37rv infected	1.7815	0.438	0

Zero-inflation model	Estimate	SE	p-value
(Intercept)	-0.7554	0.3686	0.0404
groupdms0 not infected	0.1755	0.5553	0.752
grouprapachlo not infected	2.4925	0.6732	2e-04
groupafri not infected	0.2864	0.4651	0.5381
groupbeijing not infected	0.4171	0.4636	0.3683
group37ra not infected	0.323	0.6114	0.5973
group37rv not infected	-0.459	0.6239	0.462
groupafri infected	-0.8359	0.4764	0.0794
groupbeijing infected	-1.3416	0.4919	0.0064
group37ra infected	-0.7319	0.6441	0.2558
group37rv infected	-1.1875	0.5809	0.0409

Contrast	estimate	SE	p-value
ctrl not infected-dms0 not infected	0.0094	0.4321	0.9826
ctrl not infected-rapachlo not infected	-2.4283	0.3804	0
ctrl not infected-afri not infected	-0.072	0.3747	0.889
ctrl not infected-beijing not infected	-0.3621	0.3744	0.4949
ctrl not infected-h37ra not infected	-0.4201	0.492	0.5211
ctrl not infected-h37rv not infected	-0.8728	0.4393	0.0923
ctrl not infected-afri infected	-1.1064	0.3731	0.0078
ctrl not infected-beijing infected	-1.2688	0.3729	0.0021
ctrl not infected-h37ra infected	-1.393	0.4908	0.0111
ctrl not infected-h37rv infected	-1.7815	0.438	2e-04
dms0 not infected-rapachlo not infected	2.4377	0.4167	0
dms0 not infected-afri not infected	-0.0815	0.4118	0.889
dms0 not infected-beijing not infected	-0.3715	0.4115	0.5211
dms0 not infected-h37ra not infected	-0.4296	0.5176	0.5211
dms0 not infected-h37rv not infected	-0.8822	0.4736	0.1125
dms0 not infected-afri infected	-1.1158	0.4104	0.0145
dms0 not infected-beijing infected	-1.2783	0.4101	0.0051
dms0 not infected-h37ra infected	-1.4024	0.5165	0.0145
dms0 not infected-h37rv infected	-1.7909	0.4723	6e-04
rapachlo not infected-afri not infected	2.3563	0.3553	0
rapachlo not infected-beijing not infected	2.0663	0.3548	0
rapachlo not infected-h37ra not infected	2.0082	0.4803	2e-04
rapachlo not infected-h37rv not infected	1.5555	0.4204	8e-04
rapachlo not infected-afri infected	1.3219	0.3536	7e-04
rapachlo not infected-beijing infected	1.1595	0.3532	0.003
rapachlo not infected-h37ra infected	1.0353	0.4791	0.0633
rapachlo not infected-h37rv infected	0.6468	0.419	0.2035
afri not infected-beijing not infected	-0.29	0.3471	0.5211
afri not infected-h37ra not infected	-0.3481	0.4787	0.5729
afri not infected-h37rv not infected	-0.8007	0.4108	0.0964
beijing not infected-h37ra not infected	-0.0581	0.4785	0.9249
beijing not infected-h37rv not infected	-0.5107	0.4104	0.328
h37ra not infected-h37rv not infected	-0.4526	0.5515	0.5211
afri infected-beijing infected	-0.1625	0.3437	0.7203
afri infected-h37ra infected	-0.2866	0.4763	0.6362
afri infected-h37rv infected	-0.6751	0.4079	0.1689
beijing infected-h37ra infected	-0.1242	0.476	0.8757
beijing infected-h37rv infected	-0.5127	0.4075	0.328
h37ra infected-h37rv infected	-0.3885	0.5494	0.5729

b) Area of autophagosomes



Par	Value	Std.Error	p-value
(Intercept)	-0.2574	0.1046	0.0138
groupdms0 not infected	-0.0567	0.1556	0.717
grouprapachlo not infected	0.4966	0.1129	3e-04
groupafri not infected	-0.2415	0.1305	0.0642
groupbeijing not infected	-0.172	0.1312	0.1898
group37ra not infected	-0.3725	0.1464	0.011
group37rv not infected	-0.0446	0.1376	0.7462
groupafri infected	0.0021	0.1179	0.986
groupbeijing infected	0.0308	0.1161	0.7908
group37ra infected	0.3378	0.1302	0.0095
group37rv infected	-0.006	0.1246	0.9618

Contrast	estimate	SE	p-value
ctrl not infected-dms0 not infected	0.0567	0.1556	0.855
ctrl not infected-rapachlo not infected	-0.4966	0.1129	3e-04
ctrl not infected-afri not infected	0.2415	0.1305	0.1315
ctrl not infected-beijing not infected	0.172	0.1312	0.3265
ctrl not infected-h37ra not infected	0.3725	0.1464	0.0276
ctrl not infected-h37rv not infected	0.0446	0.1376	0.855
ctrl not infected-afri infected	-0.0021	0.1179	0.986
ctrl not infected-beijing infected	-0.308	0.1161	0.872
ctrl not infected-h37ra infected	-0.3378	0.1302	0.0255
ctrl not infected-h37rv infected	0.006	0.1246	0.9847
dms0 not infected-rapachlo not infected	-0.5533	0.1366	8e-04
dms0 not infected-afri not infected	0.1847	0.151	0.3764
dms0 not infected-beijing not infected	0.1153	0.1516	0.6688
dms0 not infected-h37ra not infected	0.3157	0.1642	0.1305
dms0 not infected-h37rv not infected	-0.0122	0.1571	0.9843
dms0 not infected-afri infected	-0.0588	0.1404	0.855
dms0 not infected-beijing infected	-0.0875	0.1387	0.7612
dms0 not infected-h37ra infected	-0.3945	0.15	0.0276
dms0 not infected-h37rv infected	-0.0508	0.1458	0.855
rapachlo not infected-afri not infected	0.738	0.1046	0
rapachlo not infected-beijing not infected	0.6686	0.1056	0
rapachlo not infected-h37ra not infected	0.8691	0.1255	0
rapachlo not infected-h37rv not infected	0.5411	0.1133	1e-04
rapachlo not infected-afri infected	0.4945	0.0884	0
rapachlo not infected-beijing infected	0.4658	0.0861	0
rapachlo not infected-h37ra infected	0.1588	0.1059	0.2629
rapachlo not infected-h37rv infected	0.5026	0.0971	0
afri not infected-beijing not infected	-0.0695	0.124	0.7732
afri not infected-h37ra not infected	0.131	0.1409	0.5416
afri not infected-h37rv not infected	-0.1969	0.1303	0.2558
beijing not infected-h37ra not infected	0.2005	0.1417	0.2813
beijing not infected-h37rv not infected	-0.1274	0.1301	0.5214
h37ra not infected-h37rv not infected	-0.3279	0.1325	0.0302
afri infected-beijing infected	-0.0287	0.0923	0.855
afri infected-h37ra infected	-0.3357	0.1106	0.0079
afri infected-h37rv infected	0.008	0.1023	0.9843
beijing infected-h37ra infected	-0.307	0.1083	0.0141
beijing infected-h37rv infected	0.0368	0.0994	0.855
h37ra infected-h37rv infected	0.3438	0.0947	0.0011
afri not infected-afri infected	-0.2435	0.0704	0.002
beijing not infected-beijing infected	-0.2028	0.0729	0.0155
h37ra not infected-h37ra infected	-0.7103	0.0768	0
h37rv not infected-h37rv infected	-0.0386	0.0653	0.7693

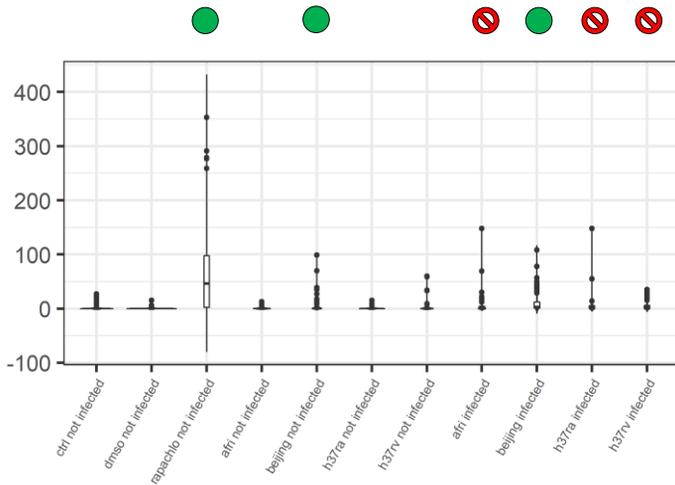
● Autophagosomes observed

⊘ Autophagosomes not observed (blocked)

Grouping: Sample

M2 – 24 h p.i.

a) Number of autophagosomes

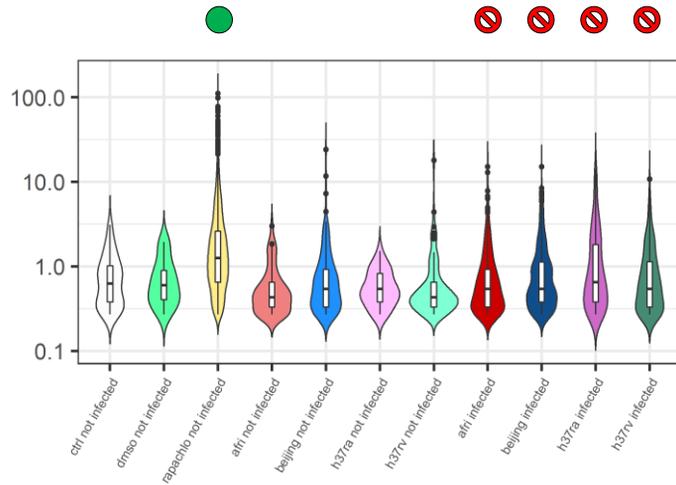


Conditional model	Estimate	SE	p-value
(Intercept)	0.9132	0.3393	0.0071
groupdmsol not infected	-0.4106	0.4739	0.3863
grouprapachlo not infected	2.9377	0.3858	0
groupafri not infected	-0.2701	0.3944	0.4934
groupbeijing not infected	1.0085	0.3868	0.0091
grouph37ra not infected	-0.4356	0.5111	0.394
grouph37rv not infected	0.8393	0.4565	0.066
groupafri infected	0.8416	0.3869	0.0296
groupbeijing infected	1.4501	0.3847	2e-04
grouph37ra infected	0.6558	0.4973	0.1873
grouph37rv infected	0.9766	0.4541	0.0315

Zero-inflation model	Estimate	SE	p-value
(Intercept)	0.092	0.3961	0.8163
groupdmsol not infected	0.674	0.5865	0.2505
grouprapachlo not infected	-2.1108	0.5782	8e-04
groupafri not infected	-0.1785	0.4967	0.7193
groupbeijing not infected	-0.2535	0.4915	0.606
grouph37ra not infected	-0.3638	0.6681	0.5861
grouph37rv not infected	-0.0641	0.5851	0.9127
groupafri infected	-0.6637	0.4802	0.1669
groupbeijing infected	-1.4196	0.4873	0.0036
grouph37ra infected	-1.3229	0.6502	0.0419
grouph37rv infected	-1.3892	0.5963	0.0198

Contrast	estimate	SE	p-value
ctrl not infected-dmsol not infected	0.4106	0.4739	0.4843
ctrl not infected-rapachlo not infected	-2.9377	0.3858	0
ctrl not infected-afri not infected	0.2701	0.3944	0.5895
ctrl not infected-beijing not infected	-1.0085	0.3868	0.0181
ctrl not infected-h37ra not infected	0.4356	0.5111	0.4843
ctrl not infected-h37rv not infected	-0.8393	0.4565	0.1019
ctrl not infected-afri infected	-0.8416	0.3869	0.0514
ctrl not infected-beijing infected	1.4501	0.3847	8e-04
ctrl not infected-h37ra infected	-0.6558	0.4973	0.2521
ctrl not infected-h37rv infected	-0.9766	0.4541	0.0525
dmsol not infected-rapachlo not infected	-3.3484	0.4261	0
dmsol not infected-afri not infected	-0.1405	0.4552	0.7757
dmsol not infected-beijing not infected	-1.4192	0.4487	0.0041
dmsol not infected-h37ra not infected	0.025	0.5638	0.9647
dmsol not infected-h37rv not infected	-1.2499	0.5062	0.0256
dmsol not infected-afri infected	-1.2522	0.4487	0.0121
dmsol not infected-beijing infected	-1.8607	0.4468	1e-04
dmsol not infected-h37ra infected	-1.0664	0.5514	0.0851
dmsol not infected-h37rv infected	-1.3873	0.504	0.013
rapachlo not infected-afri not infected	3.2079	0.3588	0
rapachlo not infected-beijing not infected	1.9292	0.3498	0
rapachlo not infected-h37ra not infected	3.5734	0.4897	0
rapachlo not infected-h37rv not infected	2.0985	0.4209	0
rapachlo not infected-afri infected	2.0962	0.3503	0
rapachlo not infected-beijing infected	1.4877	0.3474	1e-04
rapachlo not infected-h37ra infected	2.282	0.4754	0
rapachlo not infected-h37rv infected	1.9611	0.4181	0
afri not infected-beijing not infected	-1.2786	0.3573	0.001
afri not infected-h37ra not infected	0.1655	0.4986	0.7757
afri not infected-h37rv not infected	-1.1094	0.4239	0.0181
beijing not infected-h37ra not infected	1.4441	0.4925	0.0082
beijing not infected-h37rv not infected	0.1693	0.4156	0.7694
h37ra not infected-h37rv not infected	-1.2749	0.5642	0.0431
afri infected-beijing infected	-0.6085	0.3463	0.1174
afri infected-h37ra infected	0.1858	0.4784	0.7694
afri infected-h37rv infected	-0.135	0.4137	0.7757
beijing infected-h37ra infected	0.7943	0.4766	0.1332
beijing infected-h37rv infected	0.4734	0.4106	0.3247
h37ra infected-h37rv infected	-0.3208	0.5498	0.6504

b) Area of autophagosomes



Par	Value	Std.Error	p-value
(Intercept)	-0.525	0.138	1e-04
groupdmsol not infected	-0.0035	0.251	0.989
grouprapachlo not infected	0.7501	0.151	0
groupafri not infected	-0.3039	0.1967	0.1224
groupbeijing not infected	-0.0075	0.1677	0.9645
grouph37ra not infected	-0.2585	0.2404	0.2823
grouph37rv not infected	-0.2842	0.1956	0.1464
groupafri infected	-0.1504	0.1645	0.3606
groupbeijing infected	0.0289	0.1543	0.8513
grouph37ra infected	0.1817	0.1861	0.3288
grouph37rv infected	0.0488	0.1728	0.7775

Contrast	estimate	SE	p-value
ctrl not infected-dmsol not infected	0.0035	0.251	0.989
ctrl not infected-rapachlo not infected	-0.7501	0.151	0
ctrl not infected-afri not infected	0.3039	0.1967	0.3096
ctrl not infected-beijing not infected	0.0075	0.1677	0.989
ctrl not infected-h37ra not infected	0.2585	0.2404	0.4951
ctrl not infected-h37rv not infected	0.2842	0.1956	0.3325
ctrl not infected-afri infected	0.1504	0.1645	0.5743
ctrl not infected-beijing infected	-0.0289	0.1543	0.989
ctrl not infected-h37ra infected	-0.1817	0.1861	0.5437
ctrl not infected-h37rv infected	-0.0488	0.1728	0.989
dmsol not infected-rapachlo not infected	-0.7535	0.2179	0.0024
dmsol not infected-afri not infected	0.0004	0.2523	0.4951
dmsol not infected-beijing not infected	0.004	0.2304	0.989
dmsol not infected-h37ra not infected	0.255	0.2877	0.5764
dmsol not infected-h37rv not infected	0.2807	0.2514	0.4951
dmsol not infected-afri infected	0.1469	0.228	0.7206
dmsol not infected-beijing infected	-0.0324	0.2208	0.989
dmsol not infected-h37ra infected	-0.1852	0.2441	0.6421
dmsol not infected-h37rv infected	-0.0523	0.2341	0.989
rapachlo not infected-afri not infected	1.054	0.153	0
rapachlo not infected-beijing not infected	0.7575	0.1134	0
rapachlo not infected-h37ra not infected	1.0086	0.2063	0
rapachlo not infected-h37rv not infected	1.0342	0.1517	0
rapachlo not infected-afri infected	0.9004	0.1085	0
rapachlo not infected-beijing infected	0.7211	0.0925	0
rapachlo not infected-h37ra infected	0.5683	0.1391	2e-04
rapachlo not infected-h37rv infected	0.7012	0.1208	0
afri not infected-beijing not infected	-0.2964	0.1696	0.2474
afri not infected-h37ra not infected	-0.0454	0.2417	0.989
afri not infected-h37rv not infected	-0.0197	0.1972	0.989
beijing not infected-h37ra not infected	0.251	0.2188	0.4951
beijing not infected-h37rv not infected	0.2767	0.1684	0.2876
h37ra not infected-h37rv not infected	0.0257	0.2409	0.989
afri infected-beijing infected	-0.1793	0.1132	0.3041
afri infected-h37ra infected	-0.3321	0.1536	0.1015
afri infected-h37rv infected	-0.1992	0.1373	0.3325
beijing infected-h37ra infected	-0.1528	0.1428	0.4951
beijing infected-h37rv infected	-0.0199	0.125	0.989
h37ra infected-h37rv infected	0.1329	0.1626	0.6135
afri not infected-afri infected	-0.1535	0.1444	0.4951
beijing not infected-beijing infected	-0.0364	0.088	0.9124
h37ra not infected-h37ra infected	-0.4402	0.1931	0.0813
h37rv not infected-h37rv infected	-0.333	0.1384	0.0631

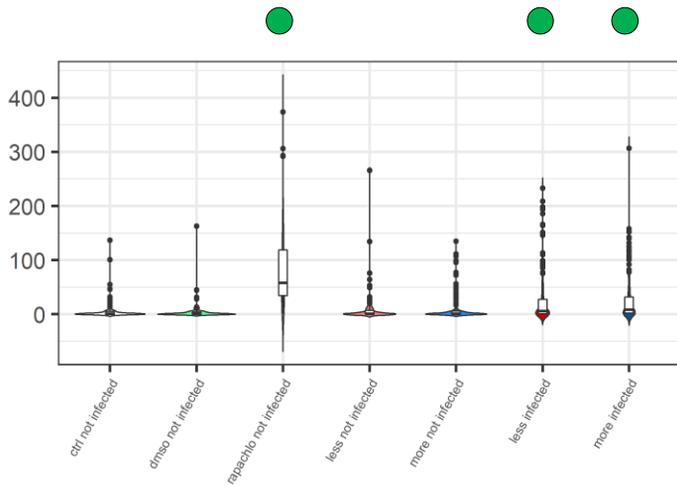
● Autophagosomes observed

⊘ Autophagosomes not observed (blocked)

Grouping: Virulence (absolute)

M1 – 24 h p.i.

a) Number of autophagosomes

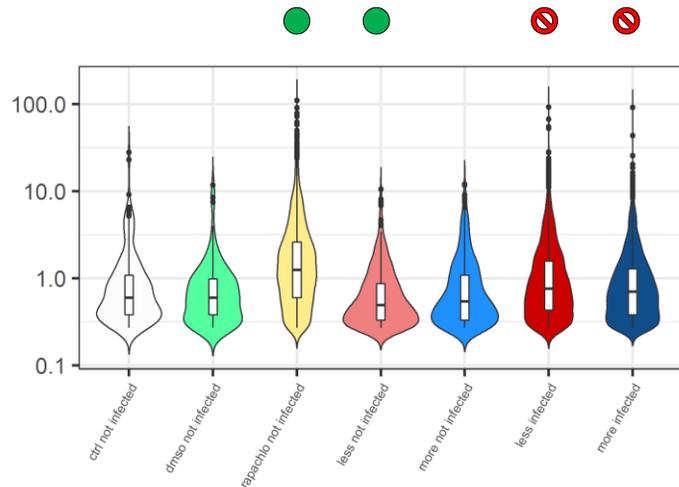


Conditional model	Estimate	SE	p-value
(Intercept)	1.7288	0.3423	0
groupdmsmo not infected	-0.0058	0.4427	0.9896
grouprapachlo not infected	2.4329	0.3898	0
groupless not infected	0.1886	0.3587	0.599
groupmore not infected	0.5518	0.3548	0.1199
groupless infected	1.1944	0.3578	8e-04
groupmore infected	1.4598	0.354	0

Zero-inflation model	Estimate	SE	p-value
(Intercept)	-0.7778	0.3791	0.0402
groupdmsmo not infected	0.1669	0.5813	0.774
grouprapachlo not infected	-2.4891	0.6899	3e-04
groupless not infected	0.303	0.4541	0.5046
groupmore not infected	0.22	0.4574	0.6305
groupless infected	-0.8096	0.4637	0.0808
groupmore infected	-1.2812	0.4676	0.0061

Contrast	estimate	SE	p-value
ctrl not infected-dmsmo not infected	0.0058	0.4427	0.9896
ctrl not infected-rapachlo not infected	-2.4329	0.3898	0
ctrl not infected-less not infected	-0.1886	0.3587	0.6601
ctrl not infected-more not infected	-0.5518	0.3548	0.1758
ctrl not infected-less infected	-1.1944	0.3578	0.0017
ctrl not infected-more infected	-1.4598	0.354	1e-04
dmsmo not infected-rapachlo not infected	-2.4387	0.427	0
dmsmo not infected-less not infected	-0.1944	0.3979	0.6601
dmsmo not infected-more not infected	-0.5575	0.3965	0.2172
dmsmo not infected-less infected	-1.2001	0.3972	0.0045
dmsmo not infected-more infected	-1.4655	0.3958	5e-04
rapachlo not infected-less not infected	2.2443	0.3386	0
rapachlo not infected-more not infected	1.8811	0.3325	0
rapachlo not infected-less infected	1.2385	0.3377	5e-04
rapachlo not infected-more infected	0.9731	0.3316	0.0054
less not infected-more not infected	-0.3632	0.2993	0.2854
less infected-more infected	-0.2654	0.2973	0.442
less not infected-less infected	-1.0058	0.0313	0
more not infected-more infected	-0.908	0.0295	0

b) Area of autophagosomes



Par	Value	Std.Error	p-value
(Intercept)	-0.2615	0.1042	0.0121
groupdmsmo not infected	-0.0602	0.1576	0.7041
grouprapachlo not infected	0.5016	0.1147	1e-04
groupless not infected	-0.3127	0.1195	0.0089
groupmore not infected	-0.1227	0.1182	0.2992
groupless infected	0.1492	0.1115	0.1809
groupmore infected	-0.0145	0.1104	0.8952

Contrast	estimate	SE	p-value
ctrl not infected-dmsmo not infected	0.0602	0.1576	0.776
ctrl not infected-rapachlo not infected	-0.5016	0.1147	2e-04
ctrl not infected-less not infected	0.3127	0.1195	0.0211
ctrl not infected-more not infected	0.1227	0.1182	0.379
ctrl not infected-less infected	-0.1492	0.1115	0.2455
ctrl not infected-more infected	0.0145	0.1104	0.8952
dmsmo not infected-rapachlo not infected	-0.5018	0.1382	5e-04
dmsmo not infected-less not infected	0.2524	0.1416	0.1286
dmsmo not infected-more not infected	0.0625	0.1408	0.776
dmsmo not infected-less infected	-0.2094	0.135	0.1865
dmsmo not infected-more infected	-0.0457	0.1342	0.776
rapachlo not infected-less not infected	0.8143	0.0907	0
rapachlo not infected-more not infected	0.6243	0.0881	0
rapachlo not infected-less infected	0.3524	0.0798	2e-04
rapachlo not infected-more infected	0.5161	0.0772	0
less not infected-more not infected	-0.1899	0.0912	0.0645
less infected-more infected	0.1638	0.0669	0.0303
less not infected-less infected	-0.4619	0.0519	0
more not infected-more infected	-0.1082	0.0487	0.0501

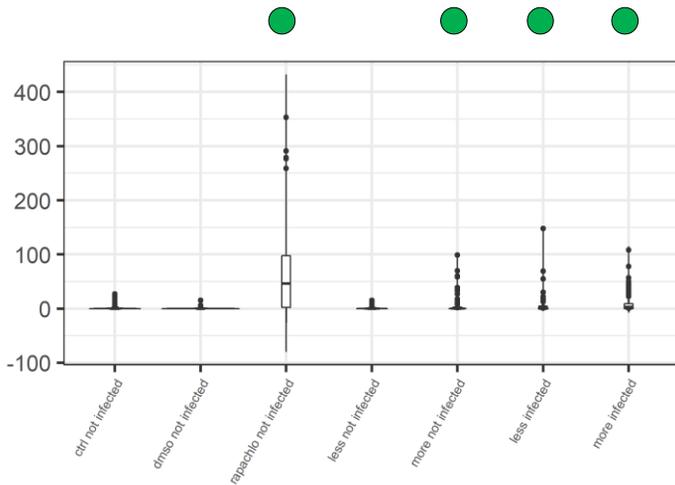
● Autophagosomes observed

⊘ Autophagosomes not observed (blocked)

Grouping: Virulence (absolute)

M2 – 24 h.p.i.

a) Number of autophagosomes

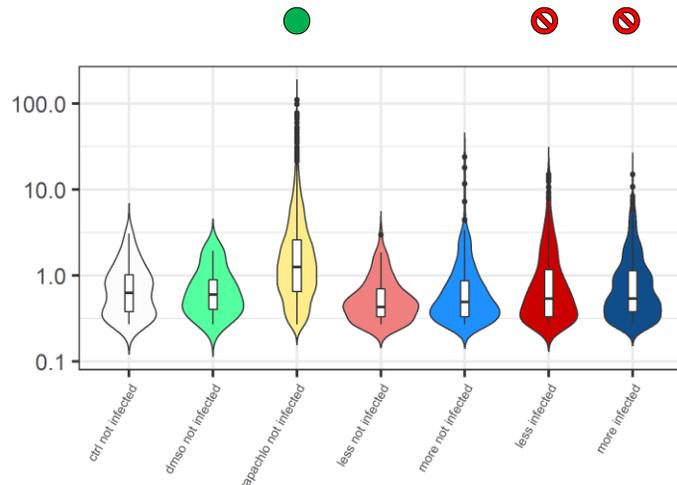


Conditional model	Estimate	SE	p-value
(Intercept)	0.9112	0.3425	0.0078
groupdmsa not infected	-0.4118	0.4779	0.3889
grouprapachlo not infected	2.9391	0.3895	0
groupless not infected	-0.3171	0.3708	0.3924
groupmore not infected	0.9399	0.3647	0.01
groupless infected	0.7887	0.3652	0.0308
groupmore infected	1.2889	0.3634	4e-04

Zero-inflation model	Estimate	SE	p-value
(Intercept)	0.1093	0.4008	0.7851
groupdmsa not infected	0.663	0.5865	0.2583
grouprapachlo not infected	-2.1417	0.579	2e-04
groupless not infected	-0.2317	0.4615	0.6157
groupmore not infected	-0.2147	0.457	0.6385
groupless infected	-0.8417	0.4466	0.0595
groupmore infected	-1.4343	0.4574	0.0017

Contrast	estimate	SE	p-value
ctrl not infected-dmsa not infected	0.4118	0.4779	0.4145
ctrl not infected-rapachlo not infected	-2.9391	0.3895	0
ctrl not infected-less not infected	-0.3171	0.3708	0.4145
ctrl not infected-more not infected	-0.9399	0.3647	0.0137
ctrl not infected-less infected	-0.7887	0.3652	0.0393
ctrl not infected-more infected	-1.2889	0.3634	7e-04
dmsa not infected-rapachlo not infected	-3.3509	0.4292	0
dmsa not infected-less not infected	-0.0946	0.437	0.8286
dmsa not infected-more not infected	-1.3517	0.4287	0.0026
dmsa not infected-less infected	-1.2005	0.4322	0.0082
dmsa not infected-more infected	-1.7007	0.4276	1e-04
rapachlo not infected-less not infected	3.2562	0.3344	0
rapachlo not infected-more not infected	1.9992	0.3229	0
rapachlo not infected-less infected	2.1504	0.3281	0
rapachlo not infected-more infected	1.6502	0.3213	0
less not infected-more not infected	-1.2571	0.3042	1e-04
less not infected-more infected	-0.5002	0.2954	0.1078
less not infected-less infected	-1.1058	0.0778	0
more not infected-more infected	-0.349	0.046	0

b) Area of autophagosomes



Par	Value	Std.Error	p-value
(Intercept)	-0.5253	0.1391	2e-04
groupdmsa not infected	-0.0026	0.2525	0.9918
grouprapachlo not infected	0.7491	0.1526	0
groupless not infected	-0.2982	0.1804	0.0983
groupmore not infected	-0.0932	0.1602	0.5606
groupless infected	-0.0376	0.1574	0.8113
groupmore infected	0.0296	0.1509	0.8447

Contrast	estimate	SE	p-value
ctrl not infected-dmsa not infected	0.0026	0.2525	0.9918
ctrl not infected-rapachlo not infected	-0.7491	0.1526	0
ctrl not infected-less not infected	0.2982	0.1804	0.2084
ctrl not infected-more not infected	0.0932	0.1602	0.8193
ctrl not infected-less infected	0.0376	0.1574	0.9322
ctrl not infected-more infected	-0.0296	0.1509	0.9322
dmsa not infected-rapachlo not infected	-0.7517	0.2193	0.0019
dmsa not infected-less not infected	0.2956	0.24	0.3769
dmsa not infected-more not infected	0.0906	0.2252	0.9322
dmsa not infected-less infected	0.035	0.2233	0.9322
dmsa not infected-more infected	-0.0322	0.2187	0.9322
rapachlo not infected-less not infected	1.0473	0.1309	0
rapachlo not infected-more not infected	0.8424	0.1013	0
rapachlo not infected-less infected	0.7867	0.0969	0
rapachlo not infected-more infected	0.7196	0.0859	0
less not infected-more not infected	-0.205	0.1397	0.2703
less not infected-more infected	-0.0671	0.0942	0.7538
less not infected-less infected	-0.2606	0.1158	0.0664
more not infected-more infected	-0.1228	0.0743	0.2084

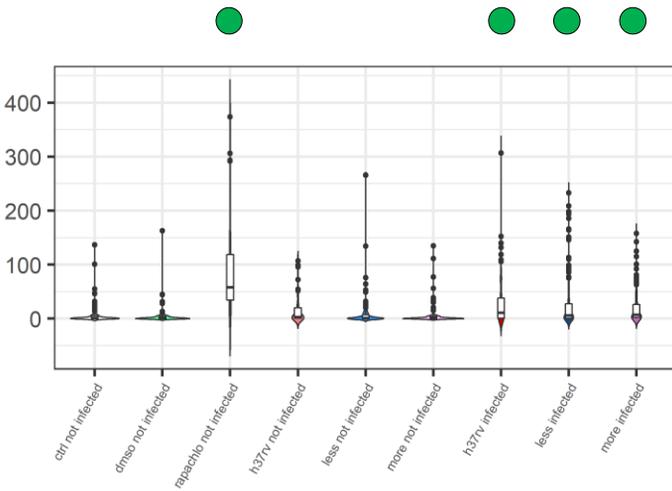
● Autophagosomes observed

⊘ Autophagosomes not observed (blocked)

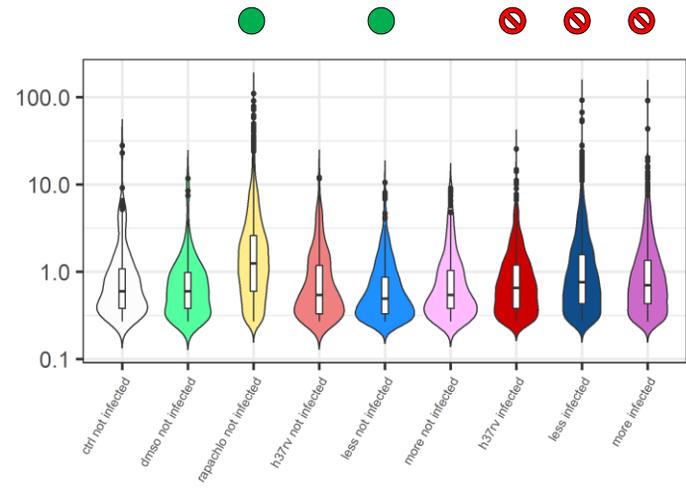
Grouping: Virulence (relative)

M1 – 24 h.p.i.

a) Number of autophagosomes



b) Area of autophagosomes



Conditional model	Estimate	SE	p-value
(Intercept)	1.7265	0.3436	0
groupdmsno not infected	-0.0113	0.4328	0.9791
grouprapachlo not infected	2.431	0.381	0
grouph37rv not infected	0.8953	0.4389	0.0414
groupless not infected	0.1788	0.3507	0.6101
groupmore not infected	0.3672	0.375	0.3275
grouph37rv infected	1.8041	0.4376	0
groupless infected	1.1846	0.3498	7e-04
groupmore infected	1.274	0.3735	6e-04

Zero-inflation model	Estimate	SE	p-value
(Intercept)	-0.7583	0.3682	0.0394
groupdmsno not infected	0.1736	0.5571	0.7554
grouprapachlo not infected	-2.49	0.6738	2e-04
grouph37rv not infected	-0.4497	0.6209	0.4689
groupless not infected	0.2999	0.4349	0.4905
groupmore not infected	0.4204	0.4643	0.3652
grouph37rv infected	-1.1795	0.5786	0.0415
groupless infected	-0.807	0.4448	0.0696
groupmore infected	-1.3388	0.4926	0.0066

Contrast	estimate	SE	p-value
ctrl not infected-dmsno not infected	0.0113	0.4328	0.9791
ctrl not infected-rapachlo not infected	-2.431	0.381	0
ctrl not infected-h37rv not infected	-0.8953	0.4389	0.0735
ctrl not infected-less not infected	-0.1788	0.3507	0.6698
ctrl not infected-more not infected	-0.3672	0.375	0.4097
ctrl not infected-h37rv infected	-1.8041	0.4376	2e-04
ctrl not infected-less infected	-1.1846	0.3498	0.0017
ctrl not infected-more infected	-1.274	0.3735	0.0017
dmsno not infected-rapachlo not infected	-2.4423	0.4173	0
dmsno not infected-h37rv not infected	-0.9066	0.473	0.0926
dmsno not infected-less not infected	-0.1901	0.389	0.6698
dmsno not infected-more not infected	-0.3785	0.412	0.4302
dmsno not infected-h37rv infected	-1.8154	0.4718	4e-04
dmsno not infected-less infected	-1.196	0.3882	0.004
dmsno not infected-more infected	-1.2853	0.4106	0.0036
rapachlo not infected-h37rv not infected	-1.5357	0.4201	7e-04
rapachlo not infected-less not infected	2.2522	0.331	0
rapachlo not infected-more not infected	2.0638	0.3554	0
rapachlo not infected-h37rv infected	0.6269	0.4187	0.1924
rapachlo not infected-less infected	1.2463	0.33	5e-04
rapachlo not infected-more infected	1.157	0.3538	0.0024
h37rv not infected-less not infected	0.7165	0.3998	0.116
h37rv not infected-more not infected	0.5281	0.4103	0.2587
less not infected-more not infected	-0.1884	0.3238	0.6472
h37rv infected-less infected	0.6194	0.3976	0.1793
h37rv infected-more infected	0.5301	0.4074	0.2587
less infected-more infected	-0.0893	0.3211	0.8078
h37rv not infected-h37rv infected	-0.9087	0.0416	0
less not infected-less infected	-1.0058	0.0313	0
more not infected-more infected	-0.9068	0.0419	0

Par	Value	Std.Error	p-value
(Intercept)	-0.2603	0.1046	0.0129
groupdmsno not infected	-0.06	0.159	0.7074
grouprapachlo not infected	0.5009	0.116	1e-04
grouph37rv not infected	-0.1042	0.1389	0.453
groupless not infected	-0.3207	0.1209	0.008
groupmore not infected	-0.1729	0.134	0.1968
grouph37rv infected	-0.0696	0.1256	0.5794
groupless infected	0.1404	0.1131	0.2147
groupmore infected	0.0302	0.1192	0.7997

Contrast	estimate	SE	p-value
ctrl not infected-dmsno not infected	0.06	0.159	0.786
ctrl not infected-rapachlo not infected	-0.5009	0.116	3e-04
ctrl not infected-h37rv not infected	0.1042	0.1389	0.6387
ctrl not infected-less not infected	0.3207	0.1209	0.0218
ctrl not infected-more not infected	0.1729	0.134	0.3293
ctrl not infected-h37rv infected	0.0696	0.1256	0.6919
ctrl not infected-less infected	-0.1404	0.1131	0.339
ctrl not infected-more infected	-0.0302	0.1192	0.8272
dmsno not infected-rapachlo not infected	-0.5609	0.1394	7e-04
dmsno not infected-less not infected	0.0442	0.1587	0.8272
dmsno not infected-less not infected	0.2607	0.143	0.1603
dmsno not infected-more not infected	0.1129	0.1544	0.6387
dmsno not infected-h37rv infected	0.0096	0.1472	0.9484
dmsno not infected-less infected	-0.2004	0.1365	0.2977
dmsno not infected-more infected	-0.0903	0.1417	0.6876
rapachlo not infected-h37rv not infected	0.6051	0.1137	0
rapachlo not infected-less not infected	0.8216	0.0921	0
rapachlo not infected-more not infected	0.6738	0.1081	0
rapachlo not infected-h37rv infected	0.5705	0.097	0
rapachlo not infected-less infected	0.3605	0.0814	2e-04
rapachlo not infected-more infected	0.4706	0.0892	0
h37rv not infected-less not infected	0.2165	0.1117	0.1215
h37rv not infected-more not infected	0.0687	0.1309	0.6919
less not infected-more not infected	-0.1478	0.1136	0.3293
h37rv infected-less infected	-0.21	0.0825	0.0273
h37rv infected-more infected	-0.0999	0.0999	0.4763
less infected-more infected	0.1101	0.0855	0.3293
h37rv not infected-h37rv infected	-0.0346	0.0654	0.6919
less not infected-less infected	-0.4611	0.0519	0
more not infected-more infected	-0.2032	0.073	0.0163

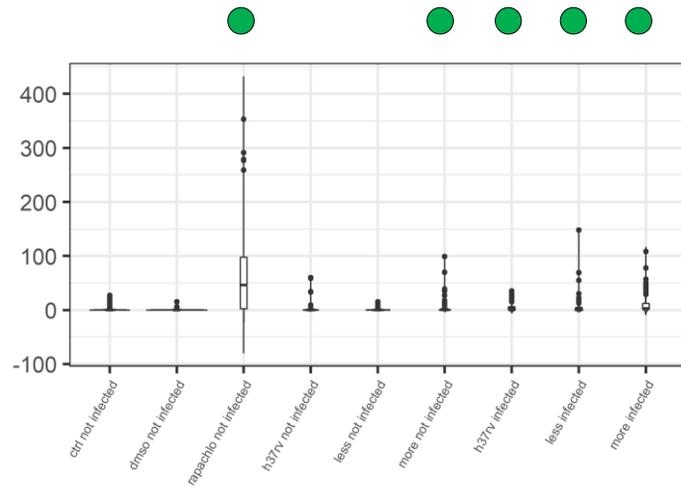
● Autophagosomes observed

⊘ Autophagosomes not observed (blocked)

Grouping: Virulence (relative)

M2 – 24 h.p.i.

a) Number of autophagosomes

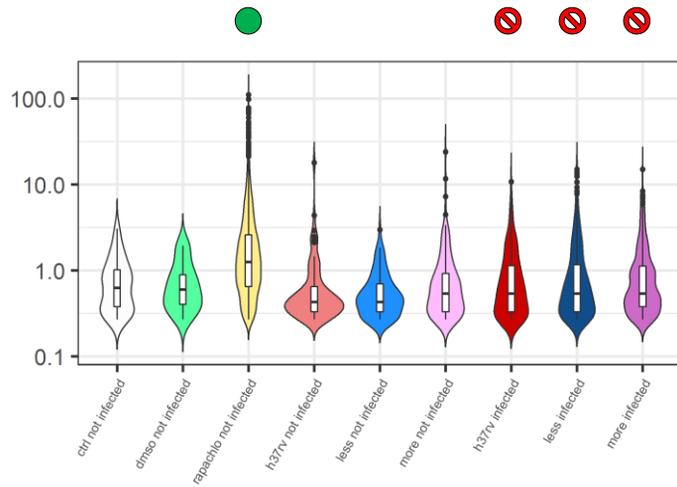


Conditional model	Estimate	SE	p-value
(Intercept)	0.9214	0.3369	0.0062
groupdmsmo not infected	-0.4132	0.474	0.3833
grouprapachlo not infected	2.9321	0.386	0
grouph37rv not infected	0.8209	0.4548	0.0711
groupless not infected	-0.3188	0.3676	0.3857
groupmore not infected	1.0011	0.3869	0.0097
grouph37rv infected	0.9582	0.4523	0.0342
groupless infected	0.7869	0.3619	0.0297
groupmore infected	1.4426	0.3847	2e-04

Zero-inflation model	Estimate	SE	p-value
(Intercept)	0.1127	0.4009	0.7786
groupdmsmo not infected	0.6644	0.586	0.2569
grouprapachlo not infected	-2.1435	0.5792	2e-04
grouph37rv not infected	-0.1159	0.5828	0.8423
groupless not infected	-0.2352	0.4612	0.61
groupmore not infected	-0.2675	0.4911	0.586
grouph37rv infected	-1.4435	0.5945	0.0152
groupless infected	-0.8454	0.4464	0.0582
groupmore infected	-1.4417	0.4866	0.003

Contrast	estimate	SE	p-value
ctrl not infected-dmsmo not infected	0.4132	0.474	0.4289
ctrl not infected-rapachlo not infected	-2.9321	0.386	0
ctrl not infected-h37rv not infected	-0.8209	0.4548	0.0931
ctrl not infected-less not infected	0.3188	0.3676	0.4289
ctrl not infected-more not infected	-1.0011	0.3869	0.0163
ctrl not infected-h37rv infected	-0.9582	0.4523	0.0492
ctrl not infected-less infected	-0.7869	0.3619	0.0449
ctrl not infected-more infected	-1.4426	0.3847	4e-04
dmsmo not infected-rapachlo not infected	-3.3453	0.4261	0
dmsmo not infected-h37rv not infected	-1.2341	0.5049	0.0232
dmsmo not infected-less not infected	-0.0944	0.4336	0.8277
dmsmo not infected-more not infected	-1.4144	0.4486	0.0036
dmsmo not infected-h37rv infected	-1.3714	0.5027	0.0114
dmsmo not infected-less infected	-1.2001	0.4288	0.0101
dmsmo not infected-more infected	-1.8559	0.4468	1e-04
rapachlo not infected-h37rv not infected	2.1112	0.4199	0
rapachlo not infected-less not infected	3.2509	0.3314	0
rapachlo not infected-more not infected	1.9309	0.3501	0
rapachlo not infected-h37rv infected	1.9739	0.4171	0
rapachlo not infected-less infected	2.1452	0.325	0
rapachlo not infected-more infected	1.4894	0.3477	1e-04
h37rv not infected-less not infected	1.1397	0.4087	0.0101
h37rv not infected-more not infected	-0.1802	0.4148	0.6922
less not infected-more not infected	-1.32	0.3313	2e-04
h37rv infected-less infected	0.1713	0.4006	0.6922
h37rv infected-more infected	-0.4845	0.4099	0.285
less infected-more infected	-0.6557	0.3223	0.0575
h37rv not infected-h37rv infected	-0.1373	0.0824	0.1203
less not infected-less infected	-1.1057	0.0778	0
more not infected-more infected	-0.4415	0.0559	0

b) Area of autophagosomes



Par	Value	Std.Error	p-value
(Intercept)	-0.5258	0.1404	2e-04
groupdmsmo not infected	-0.0014	0.2542	0.9956
grouprapachlo not infected	0.748	0.1546	0
grouph37rv not infected	-0.2846	0.1997	0.1543
groupless not infected	-0.2985	0.1819	0.1009
groupmore not infected	-0.0061	0.1712	0.9716
grouph37rv infected	0.0521	0.1769	0.7686
groupless infected	-0.0394	0.1592	0.8048
groupmore infected	0.0283	0.1579	0.8578

Contrast	estimate	SE	p-value
ctrl not infected-dmsmo not infected	0.0014	0.2542	0.9956
ctrl not infected-rapachlo not infected	-0.748	0.1546	0
ctrl not infected-h37rv not infected	0.2846	0.1997	0.3307
ctrl not infected-less not infected	0.2985	0.1819	0.2459
ctrl not infected-more not infected	0.0061	0.1712	0.9956
ctrl not infected-h37rv infected	-0.0521	0.1769	0.9956
ctrl not infected-less infected	0.0394	0.1592	0.9956
ctrl not infected-more infected	-0.0283	0.1579	0.9956
dmsmo not infected-rapachlo not infected	-0.7494	0.221	0.0026
dmsmo not infected-less not infected	0.2832	0.2552	0.5009
dmsmo not infected-more not infected	0.2971	0.2415	0.4371
dmsmo not infected-h37rv infected	0.0047	0.2335	0.9956
dmsmo not infected-less infected	-0.0535	0.2377	0.9956
dmsmo not infected-more infected	0.038	0.2249	0.9956
rapachlo not infected-h37rv not infected	-0.0297	0.2239	0.9956
rapachlo not infected-less not infected	1.0325	0.1561	0
rapachlo not infected-h37rv infected	1.0465	0.1326	0
rapachlo not infected-more not infected	0.7541	0.1174	0
rapachlo not infected-h37rv infected	0.6959	0.1256	0
rapachlo not infected-less infected	0.7873	0.0992	0
rapachlo not infected-more infected	0.7197	0.0971	0
h37rv not infected-less not infected	0.014	0.1832	0.9956
h37rv not infected-more not infected	-0.2785	0.1725	0.2459
less not infected-more not infected	-0.2924	0.1516	0.1465
h37rv infected-less infected	0.0914	0.1312	0.8579
h37rv infected-more infected	0.0238	0.1296	0.9956
less infected-more infected	-0.0677	0.1042	0.8604
h37rv not infected-h37rv infected	-0.3366	0.1391	0.0519
less not infected-less infected	-0.2592	0.116	0.0764
more not infected-more infected	-0.0344	0.0883	0.9956

● Autophagosomes observed

⊘ Autophagosomes not observed (blocked)

Apoptosis

Apoptosis/necrosis kit	
Wells/sample	2
Acquisition positions/well	2
Channels	CytoCalcein, dsRed/7-AAD, apopxin
Z-stacks	1.48 μm x 4

		NOT infected	Infected	TOT		
		N of cells	N of cells	NOT inf	Inf	
M1	Afri	205	136	689	451	1140
	Bei	164	163			
	H37Ra	94	82			
	H37Rv	88	70			
	CTRL	138	na			
M2	Afri	180	133	725	475	1200
	Bei	188	179			
	H37Ra	107	81			
	H37Rv	86	82			
	CTRL	164	na			
TOT		1414	926			
		2340				

Acquisition:

obj. 63x, oil NA 1.40

z-stacks: 1.48 μm x 4 stacks

Res. 16-bit format: 1024x1024

Laser 405 (for CytoCalcein Violet) = 6%; gain: 625, offset: 0% pinhole: 95.60

Laser 488 (for Apopxin) = 10% (Argon power: 20%); gain: 670, offset: 0% pinhole: 330

Laser 561 (for dsRed and 7-AAD) = 8%; gain: 700, offset: 0% pinhole: 95.60; Emission window: 570-662

Confocal: AP 24 FD 6 O INT 0 FIM 100%

CytoCalcein (Ex/Em 405/450)

Apopxin (Ex/Em 490/525)

dsRed (Ex/Em 584-557)

7-AAD (Ex/Em 546/657)

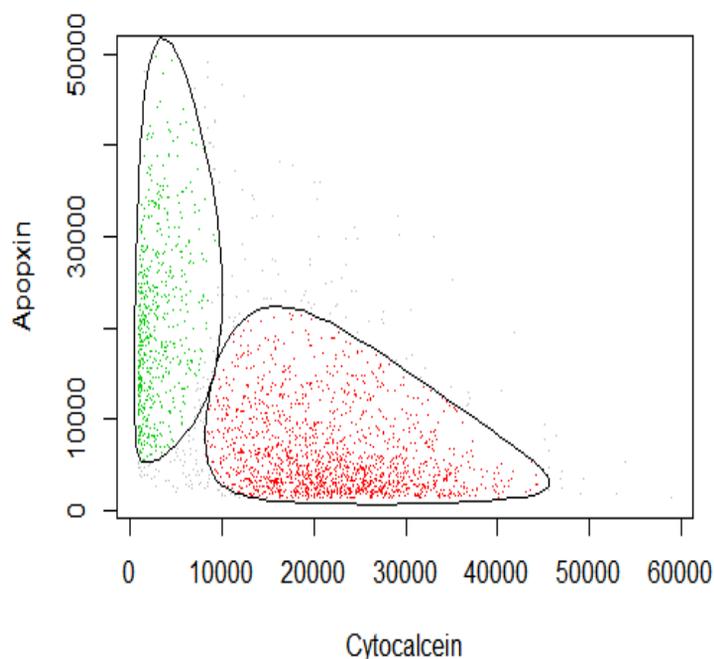
Analysis:

24 h p.i.; percentages of apoptotic/necrotic/live cells

Automated gating of apopxin-positive (apoptosis) and cytoCalcein-positive (live) cells using flowClust¹

Rule for identifying outliers: 83% quantile

Number of outliers: 232 (9.39%)



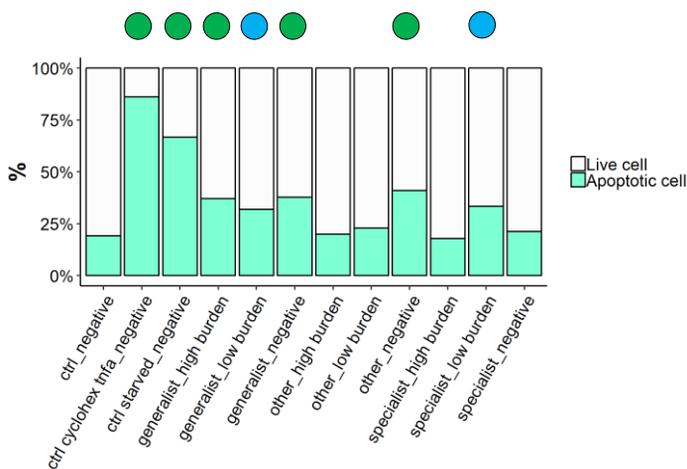
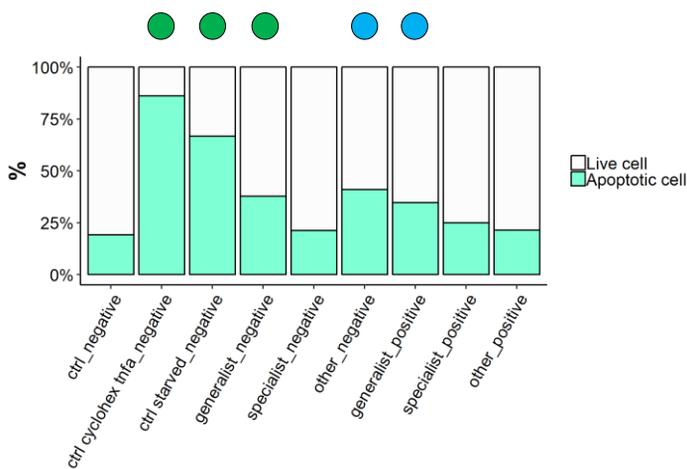
¹ Lo K, Hahne F, Brinkman RR, Gottardo R. flowClust: a Bioconductor package for automated gating of flow cytometry data. BMC Bioinformatics. 2009 May 14;10:145. doi: 10.1186/1471-2105-10-145. PMID: 19442304; PMCID: PMC2701419.

Grouping: Geo-host adaptation

M1 – 24 h.p.i.

a) Percentage of live/apoptotic cells

b) Percentage of live/apoptotic cells per mycobacterial burden



Par	Estimate	se	OR	pvalue
(Intercept)	-1.2682	0.3791	0.2813	8e-04
group_Mycctrl_cyclohex_tnf_alpha_negative	2.8044	0.6405	16.5175	0
group_Mycctrl_starved_negative	1.6833	0.5879	5.3834	0.0042
group_Mycgeneralist_negative	0.9441	0.3532	2.5704	0.0075
group_Mycspecialist_negative	0.0385	0.3808	1.0392	0.9195
group_Mycother_negative	0.9317	0.4243	2.5387	0.0281
group_Mycgeneralist_positive	0.6259	0.3592	1.8698	0.0814
group_Mycspecialist_positive	0.1463	0.3962	1.1576	0.7119
group_Mycother_positive	-0.0624	0.4652	0.9395	0.8933

Par	Estimate	se	OR	pvalue
(Intercept)	-1.2669	0.3822	0.2817	9e-04
group_Mycctrl_cyclohex_tnf_alpha_negative	2.8078	0.6485	16.5731	0
group_Mycctrl_starved_negative	1.6874	0.5967	5.4052	0.0047
group_Myc3generalist_high_burden	0.7716	0.3911	2.1632	0.0485
group_Myc3generalist_low_burden	0.4496	0.4057	1.5676	0.2679
group_Myc3generalist_negative	0.946	0.3596	2.5755	0.0085
group_Myc3other_high_burden	-0.1119	0.5629	0.8941	0.8424
group_Myc3other_low_burden	-0.0179	0.5552	0.9823	0.9743
group_Myc3other_negative	0.9275	0.4321	2.5282	0.0318
group_Myc3specialist_high_burden	-0.3219	0.4755	0.7248	0.4984
group_Myc3specialist_low_burden	0.5749	0.4474	1.777	0.1988
group_Myc3specialist_negative	0.029	0.3876	1.0294	0.9403

Contrast	estimate	SE	p.value
ctrl_negative-ctrl_cyclohex_tnf_alpha_negative	2.8044	0.6405	1e-04
ctrl_negative-ctrl_starved_negative	1.6833	0.5879	0.0117
ctrl_negative-generalist_negative	0.9441	0.3532	0.0174
ctrl_negative-specialist_negative	-0.0385	0.3808	0.9513
ctrl_negative-other_negative	-0.9317	0.4243	0.0527
ctrl_negative-generalist_positive	-0.6259	0.3592	0.1357
ctrl_negative-specialist_positive	-0.1463	0.3962	0.791
ctrl_negative-other_positive	0.0624	0.4652	0.9513
ctrl_cyclohex_tnf_alpha_negative-ctrl_starved_negative	1.1211	0.74	0.1883
ctrl_cyclohex_tnf_alpha_negative-generalist_negative	1.8604	0.5942	0.0086
ctrl_cyclohex_tnf_alpha_negative-specialist_negative	2.766	0.6083	1e-04
ctrl_cyclohex_tnf_alpha_negative-other_negative	1.8727	0.6364	0.0108
ctrl_cyclohex_tnf_alpha_negative-generalist_positive	2.1786	0.5921	0.0014
ctrl_cyclohex_tnf_alpha_negative-specialist_positive	2.6581	0.616	1e-04
ctrl_cyclohex_tnf_alpha_negative-other_positive	2.8668	0.6622	1e-04
ctrl_starved_negative-generalist_negative	0.7393	0.5371	0.2201
ctrl_starved_negative-specialist_negative	1.6449	0.5527	0.0108
ctrl_starved_negative-other_negative	0.7516	0.5835	0.2471
ctrl_starved_negative-generalist_positive	1.0575	0.5348	0.0847
ctrl_starved_negative-specialist_positive	1.537	0.5612	0.0154
ctrl_starved_negative-other_positive	1.7457	0.6115	0.0117
generalist_negative-specialist_negative	0.9056	0.293	0.0086
generalist_tnf_alpha_negative-specialist_negative	0.0124	0.3476	0.9716
specialist_negative-other_negative	-0.8932	0.3754	0.0347
generalist_positive-specialist_positive	0.4795	0.3182	0.1883
generalist_positive-other_positive	0.6883	0.4008	0.1357
specialist_positive-other_positive	0.2087	0.4345	0.7571
generalist_negative-generalist_positive	0.3182	0.2155	0.1906
specialist_negative-specialist_positive	-0.1079	0.2834	0.791
other_negative-other_positive	0.9941	0.3758	0.0175

OBSERVATIONS

In M1 macrophages, bystander non-infected cells of generalist and other strains showed increased apoptosis (40%). Despite not statistically significant, also macrophages infected with generalist strains showed a similar increase in the percentage of apoptosis. Stratification of the data by mycobacterial burden showed higher apoptotic levels in bystander non-infected cells or in cells infected with lower mycobacterial load. In M2 macrophages, only bystander non-infected cells of other strains showed a slight, statistically not significant increase in apoptosis (30%).

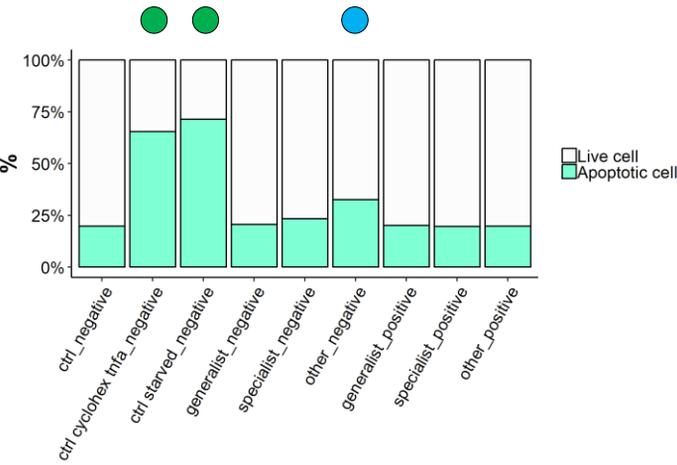
● Apoptosis > not infected control

● Increased apoptosis observed, not statistically significant

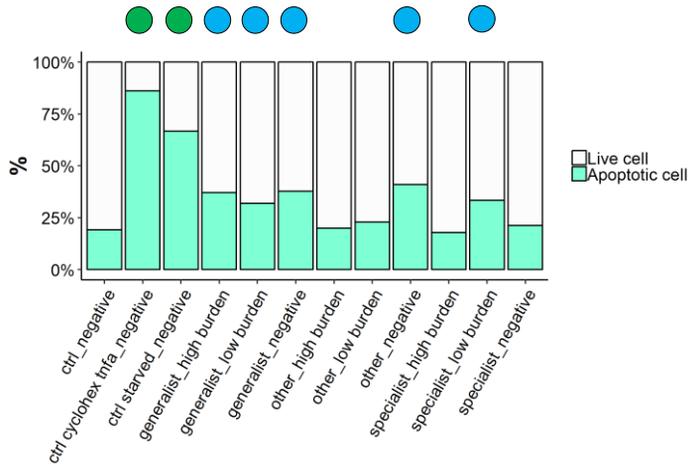
Grouping: Geo-host adaptation

M2 – 24 h p.i.

a) Percentage of live/apoptotic cells



b) Percentage of live/apoptotic cells per mycobacterial burden



Par	Estimate	se	OR	pvalue
(Intercept)	-1.4453	0.4587	0.2357	0.0016
group_Myctrl_cyclohex_tnfa_negative	1.6424	0.6211	5.1677	0.0082
group_Myctrl_starved_negative	1.9314	0.6735	6.8991	0.0041
group_Mygeneralist_negative	0.1847	0.376	1.2029	0.6233
group_Myspecialist_negative	0.3807	0.403	1.4634	0.3448
group_Myother_negative	0.7893	0.4487	2.2018	0.0786
group_Mygeneralist_positive	0.0858	0.38	1.0896	0.8213
group_Myspecialist_positive	-0.0294	0.4383	0.971	0.9465
group_Myother_positive	0.077	0.4908	1.08	0.8754

Par	Estimate	se	OR	pvalue
(Intercept)	-1.4519	0.4671	0.2341	0.0019
group_Myc3ctrl_cyclohex_tnfa_negative	1.6437	0.6321	5.1741	0.0093
group_Myc3ctrl_starved_negative	1.9332	0.6838	6.9113	0.0047
group_Myc3generalist_high_burden	0.2176	0.4263	1.2431	0.6097
group_Myc3generalist_low_burden	-0.0153	0.4183	0.9849	0.9709
group_Myc3generalist_negative	0.1931	0.384	1.213	0.6152
group_Myc3other_high_burden	-0.0368	0.6144	0.9639	0.9523
group_Myc3other_low_burden	0.17	0.5606	1.1853	0.7617
group_Myc3other_negative	0.7962	0.4586	2.2171	0.0825
group_Myc3specialist_high_burden	-0.5125	0.5468	0.599	0.3487
group_Myc3specialist_low_burden	0.3892	0.4961	1.4757	0.4328
group_Myc3specialist_negative	0.389	0.4118	1.4755	0.3448

Contrast	estimate	SE	p.value
ctrl_negative-ctrl_cyclohex_tnfa_negative	-1.6424	0.6211	0.0314
ctrl_negative-ctrl_starved_negative	-1.9314	0.6735	0.0314
ctrl_negative-generalist_negative	-0.1847	0.376	0.8499
ctrl_negative-specialist_negative	-0.3807	0.403	0.5172
ctrl_negative-other_negative	-0.7893	0.4487	0.1684
ctrl_negative-generalist_positive	-0.0858	0.38	0.9177
ctrl_negative-specialist_positive	0.0294	0.4383	0.9792
ctrl_negative-other_positive	-0.077	0.4908	0.9379
ctrl_cyclohex_tnfa_negative-ctrl_starved_negative	-0.289	0.7839	0.8905
ctrl_cyclohex_tnfa_negative-generalist_negative	1.4577	0.5727	0.0364
ctrl_cyclohex_tnfa_negative-specialist_negative	1.2617	0.5907	0.0817
ctrl_cyclohex_tnfa_negative-other_negative	0.8531	0.6223	0.3007
ctrl_cyclohex_tnfa_negative-generalist_positive	1.5566	0.5753	0.0314
ctrl_cyclohex_tnfa_negative-specialist_positive	1.6718	0.6127	0.0314
ctrl_cyclohex_tnfa_negative-other_positive	1.5654	0.6533	0.0452
ctrl_starved_negative-generalist_negative	1.7467	0.6291	0.0314
ctrl_starved_negative-specialist_negative	1.5507	0.6455	0.0452
ctrl_starved_negative-other_negative	1.1421	0.6746	0.1809
ctrl_starved_negative-generalist_positive	1.8456	0.6314	0.0314
ctrl_starved_negative-specialist_positive	1.9698	0.6657	0.0314
ctrl_starved_negative-other_positive	1.8544	0.7032	0.0314
generalist_negative-specialist_negative	-0.196	0.3238	0.7784
generalist_negative-other_negative	-0.6046	0.3792	0.2078
specialist_negative-other_negative	-0.4086	0.4059	0.4961
generalist_positive-specialist_positive	0.1152	0.3707	0.9071
generalist_positive-other_positive	0.0089	0.4316	0.9836
specialist_positive-other_positive	-0.1064	0.4836	0.9177
generalist_negative-generalist_positive	0.0989	0.2299	0.8702
specialist_negative-specialist_positive	0.4101	0.322	0.338
other_negative-other_positive	0.7123	0.3748	0.1325

OBSERVATIONS

In M2 macrophages, only bystander non-infected cells of other strains showed a slight, statistically not significant increase in apoptosis (30%).

● Apoptosis > not infected control

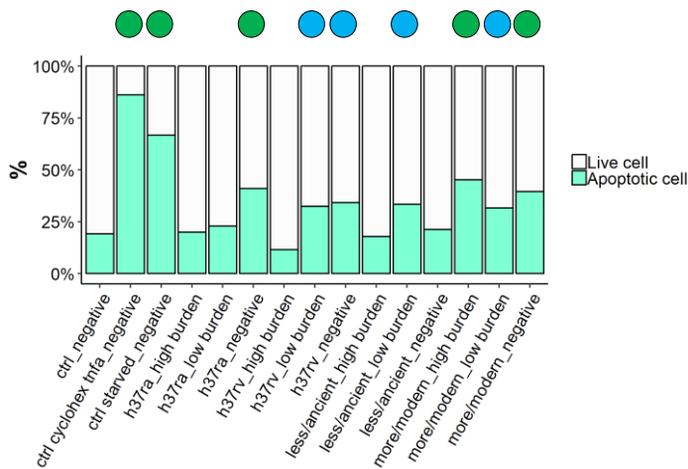
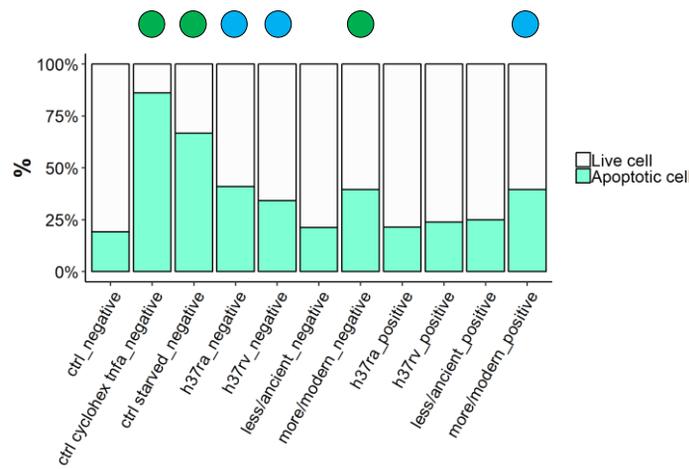
● Increased apoptosis observed, not statistically significant

Grouping: Basic

M1 – 24 h.p.i.

a) Percentage of live/apoptotic cells

b) Percentage of live/apoptotic cells per mycobacterial burden



Par	Estimate	se	OR	pvalue
(Intercept)	-1.2726	0.3691	0.2801	6e-04
group_Mycctrl_cyclohex_tnfa_negative	2.8169	0.6296	16.7247	0
group_Mycctrl_starved_negative	1.6948	0.576	5.4457	0.0033
group_Myc3h7ra_negative	0.9373	0.4135	2.553	0.0234
group_Myc3h7rv_negative	0.7326	0.4232	2.0805	0.0834
group_Mycless/ancient_negative	0.0445	0.3715	1.0455	0.9047
group_Myemore/modern_negative	1.0439	0.367	2.8402	0.0045
group_Myc3h7ra_positive	-0.0568	0.4554	0.9448	0.9008
group_Myc3h7rv_positive	0.1894	0.4636	1.2086	0.6828
group_Mycless/ancient_positive	0.1552	0.3872	1.1679	0.6886
group_Myemore/modern_positive	0.8104	0.3706	2.2489	0.0288

Par	Estimate	se	OR	pvalue
(Intercept)	-1.2712	0.3719	0.2805	6e-04
group_Mycctrl_cyclohex_tnfa_negative	2.8327	0.6446	16.9909	0
group_Mycctrl_starved_negative	1.7119	0.5924	5.5396	0.0039
group_Myc3h7ra_high_burden	-0.1092	0.56	0.8965	0.8454
group_Myc3h7ra_low_burden	-0.0101	0.5521	0.9899	0.9854
group_Myc3h7ra_negative	0.9303	0.4284	2.5352	0.0299
group_Myc3h7rv_high_burden	-0.5856	0.7226	0.5568	0.4178
group_Myc3h7rv_low_burden	0.5727	0.5218	1.7731	0.2723
group_Myc3h7rv_negative	0.7139	0.438	2.0419	0.1032
group_Myc3less/ancient_high_burden	-0.315	0.4725	0.7298	0.505
group_Myc3less/ancient_low_burden	0.5803	0.4445	1.7867	0.1917
group_Myc3less/ancient_negative	0.032	0.3843	1.0326	0.9336
group_Myc3more/modern_high_burden	1.0956	0.409	2.991	0.0074
group_Myc3more/modern_low_burden	0.3759	0.4518	1.4563	0.4054
group_Myc3more/modern_negative	1.051	0.3801	2.8606	0.0057

Contrast	estimate	SE	p.value
ctrl_negative-ctrl_cyclohex_tnfa_negative	-2.8169	0.6296	1e-04
ctrl_negative-ctrl_starved_negative	-1.6948	0.576	0.0116
ctrl_negative-h37ra_negative	-0.9373	0.4135	0.053
ctrl_negative-h37rv_negative	-0.7326	0.4232	0.1494
ctrl_negative-less/ancient_negative	-0.0445	0.3715	0.9263
ctrl_negative-more/modern_negative	-1.0439	0.367	0.0137
ctrl_negative-h37ra_positive	0.0568	0.4554	0.9263
ctrl_negative-h37rv_positive	-0.1894	0.4636	0.7666
ctrl_negative-less/ancient_positive	-0.1552	0.3872	0.7666
ctrl_negative-more/modern_positive	-0.8104	0.3706	0.0618
ctrl_cyclohex_tnfa_negative-ctrl_starved_negative	1.1221	0.7278	0.1961
ctrl_cyclohex_tnfa_negative-h37ra_negative	1.8796	0.6255	0.0114
ctrl_cyclohex_tnfa_negative-h37rv_negative	2.0843	0.634	0.0062
ctrl_cyclohex_tnfa_negative-less/ancient_negative	2.7724	0.5987	1e-04
ctrl_cyclohex_tnfa_negative-more/modern_negative	1.773	0.599	0.0116
ctrl_cyclohex_tnfa_negative-h37ra_positive	2.8737	0.6516	1e-04
ctrl_cyclohex_tnfa_negative-h37rv_positive	2.6274	0.6601	6e-04
ctrl_cyclohex_tnfa_negative-less/ancient_positive	2.6617	0.6064	1e-04
ctrl_cyclohex_tnfa_negative-more/modern_positive	2.0064	0.5936	0.0052
ctrl_starved_negative-h37ra_negative	0.7575	0.5714	0.2651
ctrl_starved_negative-h37rv_negative	0.9622	0.5807	0.164
ctrl_starved_negative-less/ancient_negative	1.6504	0.5419	0.0111
ctrl_starved_negative-more/modern_negative	0.6509	0.5423	0.3191
ctrl_starved_negative-h37ra_positive	1.7516	0.5999	0.0116
ctrl_starved_negative-h37rv_positive	1.5054	0.6091	0.034
ctrl_starved_negative-less/ancient_positive	1.5396	0.5505	0.0148
ctrl_starved_negative-more/modern_positive	0.8844	0.5364	0.164
h37ra_negative-h37rv_negative	0.2047	0.4185	0.7463
h37ra_negative-less/ancient_negative	0.8928	0.3661	0.0352
h37ra_negative-more/modern_negative	-0.1066	0.3618	0.8259
h37rv_negative-less/ancient_negative	0.6881	0.3771	0.1272
h37rv_negative-more/modern_negative	-0.3113	0.3724	0.5254
less/ancient_negative-more/modern_negative	0.9994	0.3129	0.0075
h37ra_positive-h37rv_positive	-0.2462	0.4972	0.7463
h37ra_positive-less/ancient_positive	-0.212	0.4263	0.7463
h37ra_positive-more/modern_positive	-0.8672	0.4106	0.071
h37rv_positive-less/ancient_positive	0.0343	0.4356	0.9373
h37rv_positive-more/modern_positive	-0.621	0.4208	0.215
less/ancient_positive-more/modern_positive	-0.6553	0.3337	0.0969
h37ra_negative-h37ra_positive	0.9941	0.3754	0.0217
h37rv_negative-h37rv_positive	0.5432	0.3864	0.2369
less/ancient_negative-less/ancient_positive	-0.1107	0.2827	0.7666
more/modern_negative-more/modern_positive	0.2334	0.2607	0.4981

OBSERVATIONS

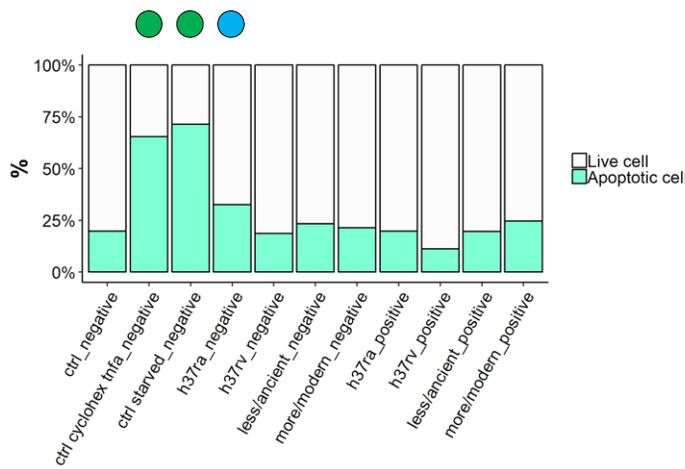
In M1 macrophages, bystander non-infected cells of more/modern strains showed increased apoptosis (40%). Despite not statistically significant, also bystander non-infected cells of H37Ra and H37Rv and macrophages infected with more/modern strains showed a similar increase in the percentage of apoptosis. Stratification of the data by mycobacterial burden showed higher apoptotic levels in bystander non-infected cells or in cells infected with lower mycobacterial load.

- Apoptosis > not infected control
- Increased apoptosis observed, not statistically significant

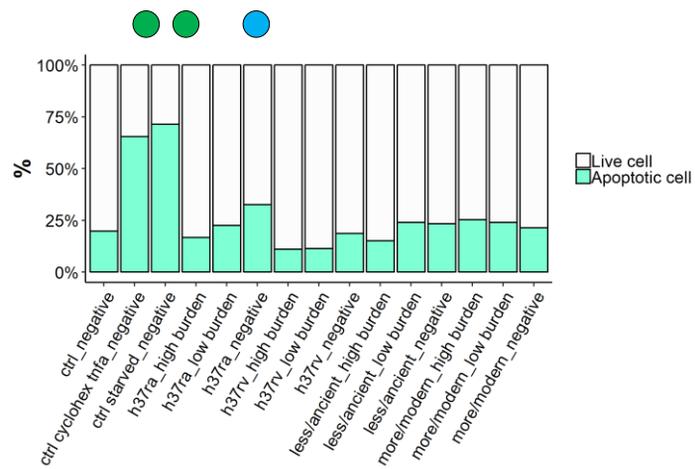
Grouping: Basic

M2 – 24 h p.i.

a) Percentage of live/apoptotic cells



b) Percentage of live/apoptotic cells per mycobacterial burden



Par	Estimate	se	OR	pvalue
(Intercept)	-1.4301	0.4477	0.2393	0.0014
group_Mycctrl cyclohex tnfα_negative	1.626	0.5984	5.0835	0.0066
group_Mycctrl starved_negative	1.9138	0.6521	6.7791	0.0033
group_Myc37ra_negative	0.7735	0.428	2.1674	0.0707
group_Myc37rv_negative	-0.0049	0.4601	0.9951	0.9915
group_Mycless/ancient_negative	0.3668	0.3852	1.4432	0.341
group_Mycomore/modern_negative	0.2543	0.3839	1.2896	0.5077
group_Myc37ra_positive	0.0671	0.4716	1.0694	0.8869
group_Myc37rv_positive	-0.5298	0.5107	0.5887	0.2995
group_Mycless/ancient_positive	-0.0332	0.4215	0.9674	0.9373
group_Mycomore/modern_positive	0.31	0.3841	1.3634	0.4197

Par	Estimate	se	OR	pvalue
(Intercept)	-1.4382	0.456	0.2373	0.0016
group_Myc3ctrl cyclohex tnfα_negative	1.6302	0.6108	5.1051	0.0076
group_Myc3ctrl starved_negative	1.9187	0.6637	6.8121	0.0038
group_Myc37ra_high burden	-0.0443	0.5994	0.9567	0.9411
group_Myc37ra_low burden	0.1597	0.5445	1.1731	0.7693
group_Myc37ra_negative	0.782	0.4392	2.1859	0.075
group_Myc37rv_high burden	-0.4505	0.7279	0.6373	0.536
group_Myc37rv_low burden	-0.5554	0.577	0.5738	0.3358
group_Myc37rv_negative	0.004	0.4704	1.004	0.9932
group_Myc3less/ancient_high burden	-0.5063	0.5331	0.6027	0.3422
group_Myc3less/ancient_low burden	0.3776	0.4817	1.4588	0.4332
group_Myc3less/ancient_negative	0.3765	0.3951	1.4571	0.3407
group_Myc3more/modern_high burden	0.4025	0.4347	1.4956	0.3545
group_Myc3more/modern_low burden	0.2268	0.4393	1.2546	0.6056
group_Myc3more/modern_negative	0.2645	0.3935	1.3028	0.5014

Contrast	estimate	SE	pvalue
ctrl_negative-ctrl cyclohex tnfα_negative	1.626	0.5984	0.0369
ctrl_negative-ctrl starved_negative	1.9138	0.6521	0.0358
ctrl_negative-h37ra_negative	-0.7735	0.428	0.1689
ctrl_negative-h37rv_negative	0.0049	0.4601	0.9915
ctrl_negative-less/ancient_negative	-0.3668	0.3852	0.4887
ctrl_negative-more/modern_negative	-0.2543	0.3839	0.642
ctrl_negative-h37ra_positive	-0.0671	0.4716	0.9302
ctrl_negative-h37rv_positive	0.5298	0.5107	0.46
ctrl_negative-less/ancient_positive	0.0332	0.4215	0.9596
ctrl_negative-more/modern_positive	-0.31	0.3841	0.5468
ctrl cyclohex tnfα_negative-ctrl starved_negative	-0.2879	0.7613	0.8197
ctrl cyclohex tnfα_negative-h37ra_negative	0.8524	0.6003	0.3042
ctrl cyclohex tnfα_negative-h37rv_negative	1.6309	0.6232	0.0381
ctrl cyclohex tnfα_negative-less/ancient_negative	1.2592	0.5712	0.0739
ctrl cyclohex tnfα_negative-more/modern_negative	1.3716	0.5708	0.0499
ctrl cyclohex tnfα_negative-h37ra_positive	1.5589	0.6322	0.0452
ctrl cyclohex tnfα_negative-h37rv_positive	2.1558	0.6634	0.0248
ctrl cyclohex tnfα_negative-less/ancient_positive	1.6592	0.5936	0.0369
ctrl cyclohex tnfα_negative-more/modern_positive	1.316	0.57	0.0601
ctrl starved_negative-h37ra_negative	1.1403	0.6539	0.1745
ctrl starved_negative-h37rv_negative	1.9187	0.6749	0.0369
ctrl starved_negative-less/ancient_negative	1.547	0.6272	0.0452
ctrl starved_negative-more/modern_negative	1.6595	0.6268	0.0381
ctrl starved_negative-h37ra_positive	1.8468	0.6832	0.0369
ctrl starved_negative-h37rv_positive	2.4436	0.7122	0.0248
ctrl starved_negative-less/ancient_positive	1.947	0.6477	0.0358
ctrl starved_negative-more/modern_positive	1.6039	0.6261	0.0407
h37ra_negative-h37rv_negative	0.7784	0.4633	0.1903
h37ra_negative-less/ancient_negative	0.4067	0.3892	0.46
h37ra_negative-more/modern_negative	0.5192	0.3879	0.3379
h37rv_negative-less/ancient_negative	-0.3717	0.4242	0.5117
h37rv_negative-more/modern_negative	-0.2592	0.423	0.6634
less/ancient_negative-more/modern_negative	0.1125	0.3401	0.8383
h37ra_positive-h37rv_positive	0.5969	0.5506	0.46
h37ra_positive-less/ancient_positive	0.1002	0.4689	0.8994
h37ra_positive-more/modern_positive	-0.2429	0.4357	0.6894
h37rv_positive-less/ancient_positive	-0.4966	0.5084	0.4873
h37rv_positive-more/modern_positive	-0.8398	0.4778	0.1745
less/ancient_positive-more/modern_positive	-0.3432	0.3808	0.5097
h37ra_negative-h37ra_positive	0.7065	0.374	0.1489
h37rv_negative-h37rv_positive	0.5249	0.4546	0.4269
less/ancient_negative-less/ancient_positive	0.4	0.3207	0.3803
more/modern_negative-more/modern_positive	-0.0556	0.2698	0.8994

OBSERVATIONS

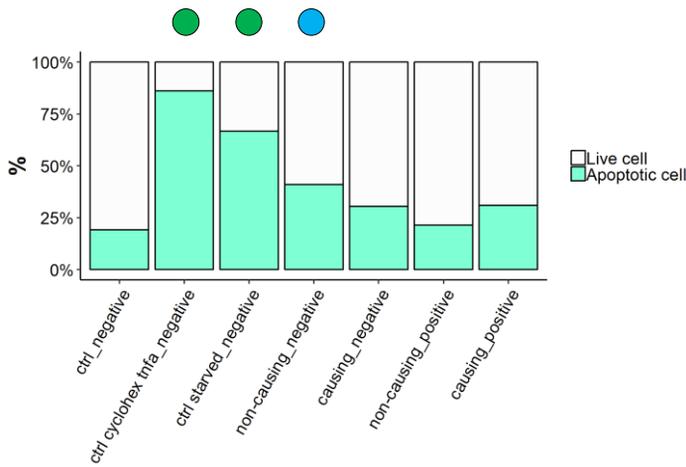
In contrast to what observed for M1, none of the category induced apoptosis in M2a macrophages at the time point considered.

- Apoptosis > not infected control
- Increased apoptosis observed, not statistically significant

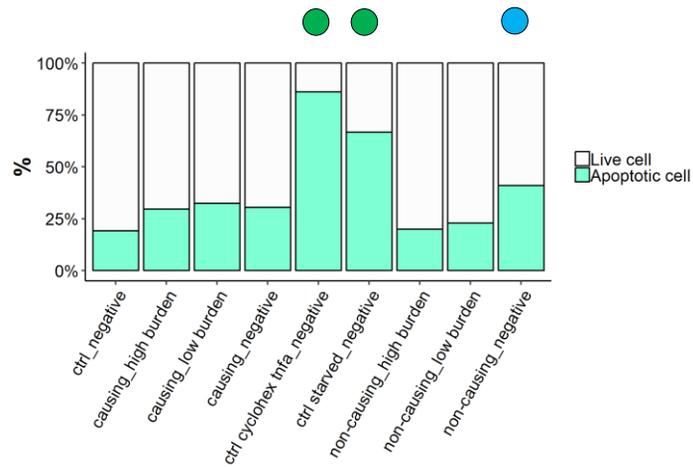
Grouping: Disease

M1 – 24 h p.i.

a) Percentage of live/apoptotic cells



b) Percentage of live/apoptotic cells per mycobacterial burden



Par	Estimate	se	OR	pvalue
(Intercept)	-1.2627	0.3961	0.2829	0.0014
group_Mycctrl cyclohex tnfα_negative	2.832	0.6877	16.9798	0
group_Mycctrl starved_negative	1.7147	0.6395	5.555	0.0073
group_Myccnon-causing_negative	0.9165	0.4703	2.5005	0.0513
group_Mycccausing_negative	0.5746	0.37	1.7764	0.1204
group_Myccnon-causing_positive	-0.071	0.5076	0.9315	0.8888
group_Mycccausing_positive	0.4299	0.375	1.5372	0.2516

Contrast	estimate	SE	p-value
ctrl_negative-ctrl cyclohex tnfα_negative	-2.832	0.6877	4e-04
ctrl_negative-ctrl starved_negative	-1.7147	0.6395	0.0199
ctrl_negative-non-causing_negative	-0.9165	0.4703	0.0887
ctrl_negative-causing_negative	-0.5746	0.37	0.1906
ctrl_negative-non-causing_positive	0.071	0.5076	0.8888
ctrl_negative-causing_positive	-0.4299	0.375	0.2987
ctrl cyclohex tnfα_negative-ctrl starved_negative	1.1173	0.7935	0.2325
ctrl cyclohex tnfα_negative-non-causing_negative	1.9155	0.6838	0.0193
ctrl cyclohex tnfα_negative-causing_negative	2.2575	0.6205	0.0013
ctrl cyclohex tnfα_negative-non-causing_positive	2.903	0.7083	4e-04
ctrl cyclohex tnfα_negative-causing_positive	2.4021	0.6199	7e-04
ctrl starved_negative-non-causing_negative	0.7982	0.6353	0.2836
ctrl starved_negative-causing_negative	1.1401	0.5666	0.084
ctrl starved_negative-non-causing_positive	1.7856	0.6615	0.0199
ctrl starved_negative-causing_positive	1.2847	0.5659	0.0489
non-causing_negative-causing_negative	0.3419	0.3643	0.3889
non-causing_positive-causing_positive	-0.5009	0.4151	0.2883
non-causing_negative-non-causing_positive	0.9875	0.377	0.0209
causing_negative-causing_positive	0.1446	0.1722	0.4232

Par	Estimate	se	OR	pvalue
(Intercept)	-1.2631	0.3952	0.2828	0.0014
group_Myc3causing_high burden	0.3793	0.3949	1.4613	0.3368
group_Myc3causing_low burden	0.4877	0.3996	1.6286	0.2222
group_Myc3causing_negative	0.5731	0.3712	1.7738	0.1226
group_Myc3ctrl cyclohex tnfα_negative	2.8381	0.6893	17.0825	0
group_Myc3ctrl starved_negative	1.7208	0.6412	5.5892	0.0073
group_Myc3non-causing_high burden	-0.1357	0.5944	0.8731	0.8194
group_Myc3non-causing_low burden	-0.0066	0.5873	0.9935	0.9911
group_Myc3non-causing_negative	0.9146	0.4719	2.4957	0.0526

OBSERVATIONS

Despite not statistically significant, in M1 macrophages bystander non-infected cells of non-causing disease strains showed increased apoptosis (40%). Also macrophages infected with non-causing disease strains showed an increase in the percentage of apoptosis. Stratification of the data by mycobacterial burden showed higher apoptotic levels in bystander non-infected cells.

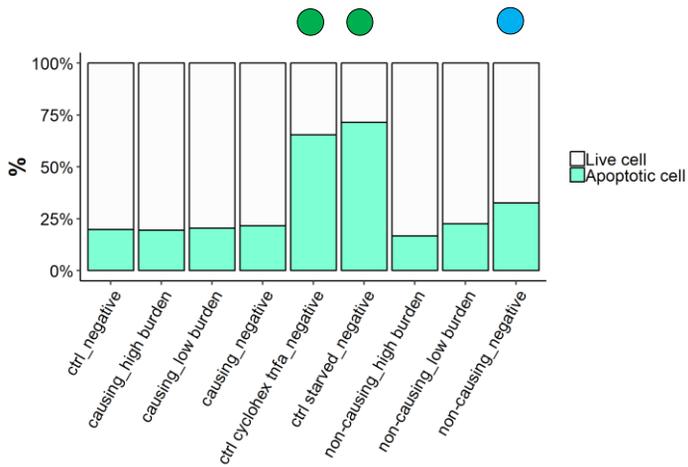
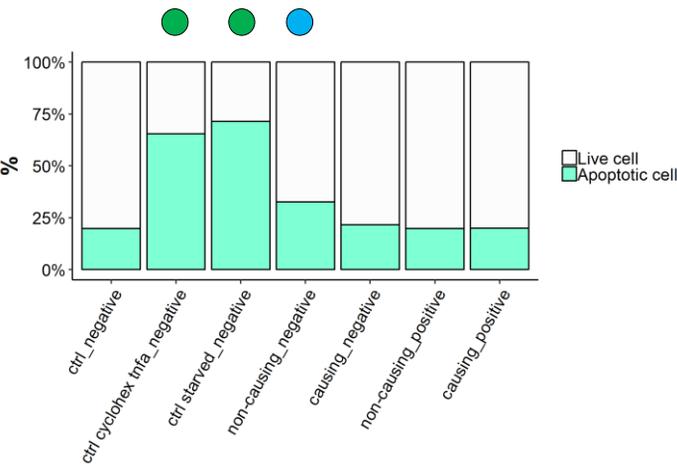
- Apoptosis > not infected control
- Increased apoptosis observed, not statistically significant

Grouping: Disease

M2 – 24 h p.i.

a) Percentage of live/apoptotic cells

b) Percentage of live/apoptotic cells per mycobacterial burden



Par	Estimate	se	OR	pvalue
(Intercept)	-1.4453	0.4574	0.2357	0.0016
group_Mycctrl_cyclohex_tnfa_negative	1.6451	0.621	5.1815	0.0081
group_Mycctrl_starved_negative	1.9341	0.6734	6.9175	0.0041
group_Mycon-causing_negative	0.7893	0.4487	2.2019	0.0785
group_Mycausing_negative	0.262	0.353	1.2995	0.4579
group_Mycon-causing_positive	0.077	0.4907	1.0801	0.8752
group_Mycausing_positive	0.0543	0.3599	1.0558	0.88

Contrast	estimate	SE	p.value
ctrl_negative-ctrl_cyclohex_tnfa_negative	-1.6451	0.621	0.0262
ctrl_negative-ctrl_starved_negative	-1.9341	0.6734	0.0262
ctrl_negative-non-causing_negative	-0.7893	0.4487	0.1492
ctrl_negative-causing_negative	-0.262	0.353	0.5801
ctrl_negative-non-causing_positive	-0.077	0.4907	0.9289
ctrl_negative-causing_positive	-0.0543	0.3599	0.9289
ctrl_cyclohex_tnfa_negative-ctrl_starved_negative	-0.289	0.7838	0.8459
ctrl_cyclohex_tnfa_negative-non-causing_negative	0.8557	0.6222	0.247
ctrl_cyclohex_tnfa_negative-causing_negative	1.3831	0.5577	0.0356
ctrl_cyclohex_tnfa_negative-non-causing_positive	1.568	0.6532	0.0389
ctrl_cyclohex_tnfa_negative-causing_positive	1.5908	0.5613	0.0262
ctrl_starved_negative-non-causing_negative	1.1447	0.6745	0.1549
ctrl_starved_negative-causing_negative	1.6721	0.6154	0.0262
ctrl_starved_negative-non-causing_positive	1.857	0.7032	0.0262
ctrl_starved_negative-causing_positive	1.8797	0.6187	0.0262
non-causing_negative-causing_negative	0.5274	0.3563	0.2198
non-causing_positive-causing_positive	0.0227	0.414	0.9562
non-causing_negative-non-causing_positive	0.7123	0.3748	0.1211
causing_negative-causing_positive	0.2077	0.1861	0.359

Par	Estimate	se	OR	pvalue
(Intercept)	-1.4466	0.4572	0.2354	0.0016
group_Myc3causing_high_burden	-0.002	0.3928	0.9981	0.996
group_Myc3causing_low_burden	0.1032	0.3827	1.1087	0.7875
group_Myc3causing_negative	0.2629	0.3543	1.3007	0.458
group_Myc3ctrl_cyclohex_tnfa_negative	1.6485	0.6229	5.1994	0.0081
group_Myc3ctrl_starved_negative	1.9376	0.6751	6.9421	0.0041
group_Myc3non-causing_high_burden	-0.0406	0.6078	0.9602	0.9467
group_Myc3non-causing_low_burden	0.1668	0.5536	1.1815	0.7632
group_Myc3non-causing_negative	0.7906	0.4502	2.2047	0.0791

OBSERVATIONS

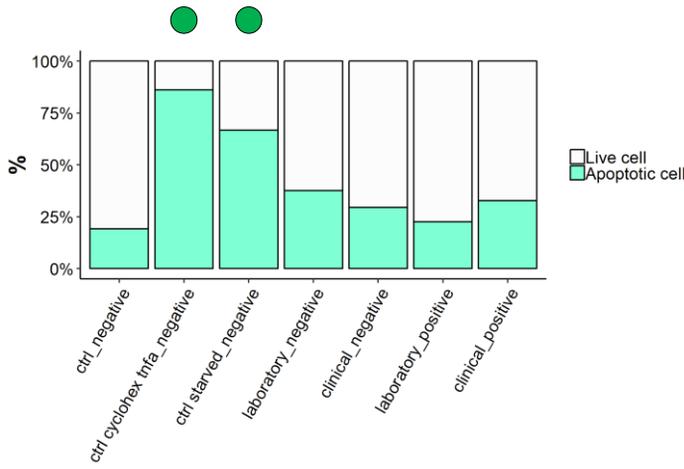
Despite not statistically significant, in M2a macrophages bystander non-infected cells of non-causing disease strains showed increased apoptosis (30%). Stratification of the data by mycobacterial burden showed higher apoptotic levels in bystander non-infected cells.

- Apoptosis > not infected control
- Increased apoptosis observed, not statistically significant

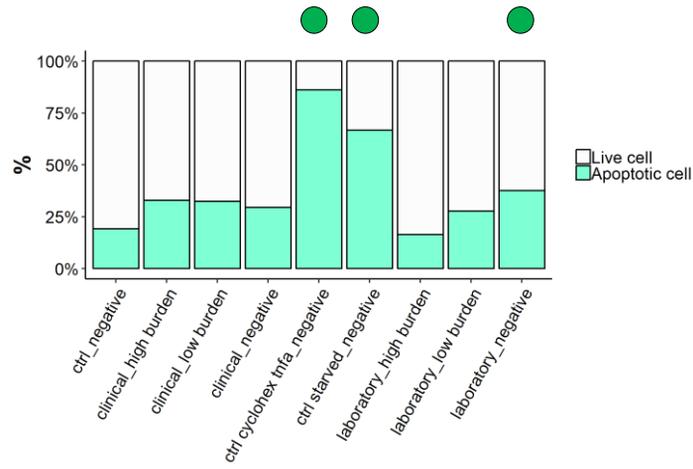
Grouping: Lab adaptation

M1 – 24 h p.i.

a) Percentage of live/apoptotic cells



b) Percentage of live/apoptotic cells per mycobacterial burden



Par	Estimate	se	OR	pvalue
(Intercept)	-1.2641	0.392	0.2825	0.0013
group_Mycctrl cyclohex tnfα_negative	2.8407	0.6851	17.1286	0
group_Mycctrl starved_negative	1.7232	0.6367	5.6026	0.0068
group_Myclaboratory_negative	0.8194	0.4091	2.2692	0.0452
group_Myclinical_negative	0.5345	0.3765	1.7065	0.1557
group_Myclaboratory_positive	0.0477	0.431	1.0488	0.9119
group_Myclinical_positive	0.4889	0.3813	1.6305	0.1998

Par	Estimate	se	OR	pvalue
(Intercept)	-1.2656	0.3889	0.2821	0.0011
group_Myc3clinical_high burden	0.5064	0.4029	1.6592	0.2088
group_Myc3clinical_low burden	0.4703	0.416	1.6004	0.2583
group_Myc3clinical_negative	0.5338	0.3795	1.7054	0.1595
group_Myc3ctrl cyclohex tnfα_negative	2.8591	0.689	17.445	0
group_Myc3ctrl starved_negative	1.7418	0.6408	5.7077	0.0066
group_Myc3laboratory_high burden	-0.3025	0.5165	0.739	0.5581
group_Myc3laboratory_low burden	0.303	0.4688	1.354	0.518
group_Myc3laboratory_negative	0.8105	0.4125	2.2491	0.0494

Contrast	estimate	SE	p.value
ctrl_negative-ctrl cyclohex tnfα_negative	-2.8407	0.6851	3e-04
ctrl_negative-ctrl starved_negative	-1.7232	0.6367	0.0161
ctrl_negative-laboratory_negative	-0.8194	0.4091	0.078
ctrl_negative-clinical_negative	-0.5345	0.3765	0.2138
ctrl_negative-laboratory_positive	-0.0477	0.431	0.9119
ctrl_negative-clinical_positive	-0.4889	0.3813	0.2373
ctrl cyclohex tnfα_negative-ctrl starved_negative	1.1175	0.7906	0.2138
ctrl cyclohex tnfα_negative-laboratory_negative	2.0213	0.6431	0.0064
ctrl cyclohex tnfα_negative-clinical_negative	2.3063	0.6232	0.001
ctrl cyclohex tnfα_negative-laboratory_positive	2.7931	0.6557	3e-04
ctrl cyclohex tnfα_negative-clinical_positive	2.3518	0.622	0.001
ctrl starved_negative-laboratory_negative	0.9038	0.5912	0.2001
ctrl starved_negative-clinical_negative	1.1888	0.5695	0.07
ctrl starved_negative-laboratory_positive	1.6756	0.6048	0.0152
ctrl starved_negative-clinical_positive	1.2343	0.5682	0.063
laboratory_negative-clinical_negative	0.285	0.2932	0.37
laboratory_positive-clinical_positive	-0.4412	0.3281	0.2264
laboratory_negative-laboratory_positive	0.7718	0.2703	0.0136
clinical_negative-clinical_positive	0.0455	0.1922	0.8578

OBSERVATIONS

Despite not statistically significant, in M1 macrophages bystander non-infected cells of laboratory strains and macrophages infected with clinical isolates showed increased apoptosis (35-40%). Stratification of the data by mycobacterial burden showed higher apoptotic levels in bystander non-infected cells or in cells infected with lower mycobacterial load for laboratory strains.

● Apoptosis > not infected control

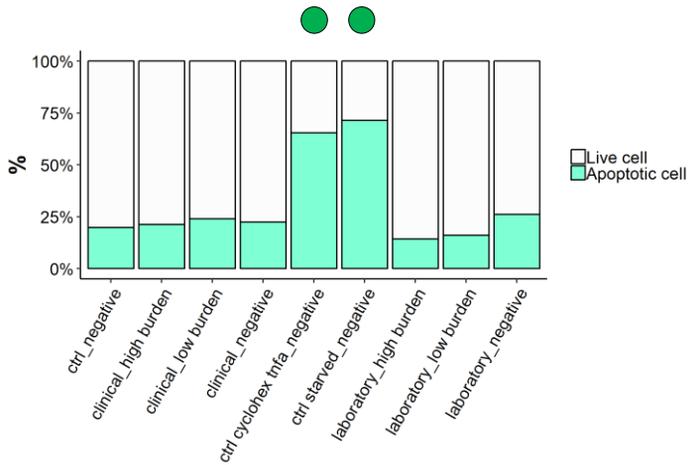
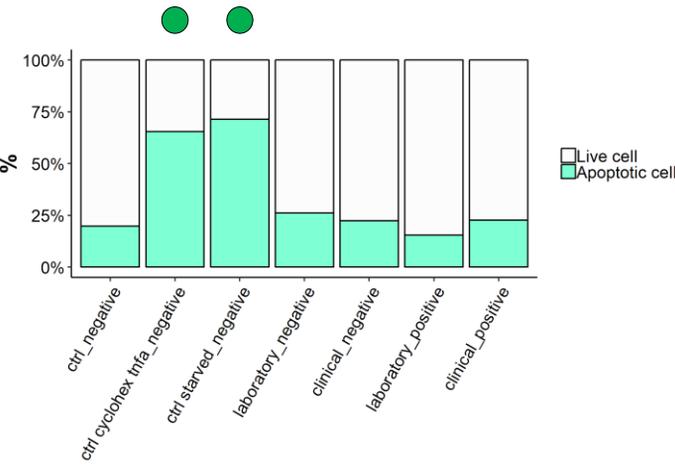
● Increased apoptosis observed, not statistically significant

Grouping: Lab adaptation

M2 – 24 h p.i.

a) Percentage of live/apoptotic cells

b) Percentage of live/apoptotic cells per mycobacterial burden



Par	Estimate	se	OR	pvalue
(Intercept)	-1.4503	0.4578	0.2345	0.0015
group_Mycctrl_cyclohex_infa_negative	1.6569	0.6283	5.243	0.0084
group_Mycctrl_starved_negative	1.9462	0.6802	7.0022	0.0042
group_Myclaboratory_negative	0.4384	0.4	1.5502	0.2731
group_Myclinical_negative	0.3283	0.3666	1.3886	0.3705
group_Myclaboratory_positive	-0.2024	0.4269	0.8168	0.6355
group_Myclinical_positive	0.1872	0.3741	1.2059	0.6168

Par	Estimate	se	OR	pvalue
(Intercept)	-1.4512	0.4583	0.2343	0.0015
group_Myc3clinical_high_burden	0.0859	0.4078	1.0897	0.8331
group_Myc3clinical_low_burden	0.2856	0.4027	1.3306	0.4782
group_Myc3clinical_negative	0.3286	0.3677	1.389	0.3715
group_Myc3ctrl_cyclohex_infa_negative	1.6585	0.6297	5.2512	0.0084
group_Myc3ctrl_starved_negative	1.9478	0.6816	7.0136	0.0043
group_Myc3laboratory_high_burden	-0.2418	0.5206	0.7852	0.6423
group_Myc3laboratory_low_burden	-0.1774	0.463	0.8374	0.7016
group_Myc3laboratory_negative	0.4391	0.4011	1.5513	0.2736

Contrast	estimate	SE	p.value
ctrl_negative-ctrl_cyclohex_infa_negative	-1.6569	0.6283	0.0278
ctrl_negative-ctrl_starved_negative	-1.9462	0.6802	0.0246
ctrl_negative-laboratory_negative	-0.4384	0.4	0.3991
ctrl_negative-clinical_negative	-0.3283	0.3666	0.5029
ctrl_negative-laboratory_positive	0.2024	0.4269	0.7103
ctrl_negative-clinical_positive	-0.1872	0.3741	0.7103
ctrl_cyclohex_infa_negative-ctrl_starved_negative	-0.2893	0.791	0.7146
ctrl_cyclohex_infa_negative-laboratory_negative	1.2185	0.5903	0.0673
ctrl_cyclohex_infa_negative-clinical_negative	1.3286	0.5691	0.0413
ctrl_cyclohex_infa_negative-laboratory_positive	1.8593	0.6098	0.0218
ctrl_cyclohex_infa_negative-clinical_positive	1.4697	0.5725	0.0278
ctrl_starved_negative-laboratory_negative	1.5078	0.6453	0.0413
ctrl_starved_negative-clinical_negative	1.6179	0.626	0.0278
ctrl_starved_negative-laboratory_positive	2.1486	0.6632	0.0218
ctrl_starved_negative-clinical_positive	1.759	0.6291	0.0246
laboratory_negative-clinical_negative	0.1101	0.2988	0.7146
laboratory_positive-clinical_positive	-0.3896	0.3422	0.3991
laboratory_negative-laboratory_positive	0.6407	0.2872	0.0488
clinical_negative-clinical_positive	0.1411	0.2053	0.6233

OBSERVATIONS

In contrast, clinical isolates showed similar apoptotic levels irrespective of the mycobacterial load. In contrast, none of the category induced apoptosis in M2a macrophages at the time point considered.

● Apoptosis > not infected control

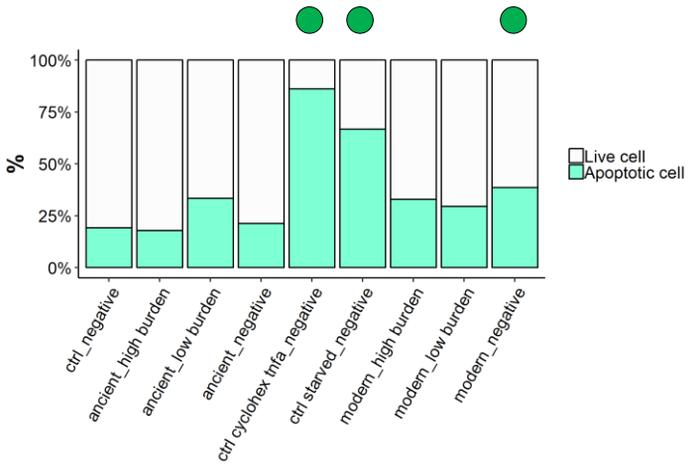
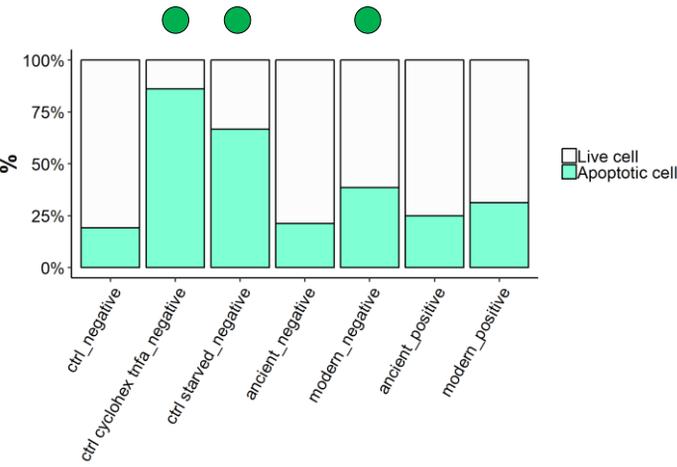
● Increased apoptosis observed, not statistically significant

Grouping: Phylogeny

M1 – 24 h p.i.

a) Percentage of live/apoptotic cells

b) Percentage of live/apoptotic cells per mycobacterial burden



Par	Estimate	se	OR	pvalue
(Intercept)	-1.2657	0.3852	0.282	0.001
group_Mycctrl_cyclohex_infa_negative	2.7973	0.6474	16.4006	0
group_Mycctrl_starved_negative	1.6768	0.5955	5.3484	0.0049
group_Mycaecient_negative	0.0349	0.3866	1.0355	0.928
group_Myccmodern_negative	0.9448	0.3457	2.5722	0.0063
group_Mycaecient_positive	0.1412	0.4018	1.1516	0.7253
group_Myccmodern_positive	0.4561	0.3521	1.5779	0.1952

Contrast	estimate	SE	p-value
ctrl_negative-ctrl_cyclohex_infa_negative	-2.7973	0.6474	1e-04
ctrl_negative-ctrl_starved_negative	-1.6768	0.5955	0.0116
ctrl_negative-ancient_negative	-0.0349	0.3866	0.928
ctrl_negative-modern_negative	-0.9448	0.3457	0.013
ctrl_negative-ancient_positive	-0.1412	0.4018	0.7656
ctrl_negative-modern_positive	-0.4561	0.3521	0.2472
ctrl_cyclohex_infa_negative-ctrl_starved_negative	1.1205	0.7477	0.1958
ctrl_cyclohex_infa_negative-ancient_negative	2.7624	0.6145	1e-04
ctrl_cyclohex_infa_negative-modern_negative	1.8526	0.5917	0.0055
ctrl_cyclohex_infa_negative-ancient_positive	2.6561	0.6222	1e-04
ctrl_cyclohex_infa_negative-modern_positive	2.3412	0.5905	3e-04
ctrl_starved_negative-ancient_negative	1.6419	0.5595	0.0091
ctrl_starved_negative-modern_negative	0.732	0.5344	0.2317
ctrl_starved_negative-ancient_positive	1.5356	0.568	0.013
ctrl_starved_negative-modern_positive	1.2207	0.5331	0.0349
ancient_negative-modern_negative	-0.9098	0.2814	0.0047
ancient_positive-modern_positive	-0.3149	0.3081	0.3642
ancient_negative-ancient_positive	-0.1063	0.2838	0.7656
modern_negative-modern_positive	0.4886	0.1865	0.0152

Par	Estimate	se	OR	pvalue
(Intercept)	-1.2645	0.3886	0.2824	0.0011
group_Myc3ancient_high_burden	-0.3279	0.4789	0.7204	0.4935
group_Myc3ancient_low_burden	0.5719	0.4509	1.7717	0.2047
group_Myc3ancient_negative	0.0262	0.3914	1.0266	0.9466
group_Myc3ctrl_cyclohex_infa_negative	2.7964	0.6531	16.386	0
group_Myc3ctrl_starved_negative	1.6764	0.6018	5.3462	0.0053
group_Myc3modern_high_burden	0.5664	0.3775	1.7619	0.1335
group_Myc3modern_low_burden	0.3257	0.3876	1.385	0.4007
group_Myc3modern_negative	0.9465	0.3501	2.5767	0.0069

OBSERVATIONS

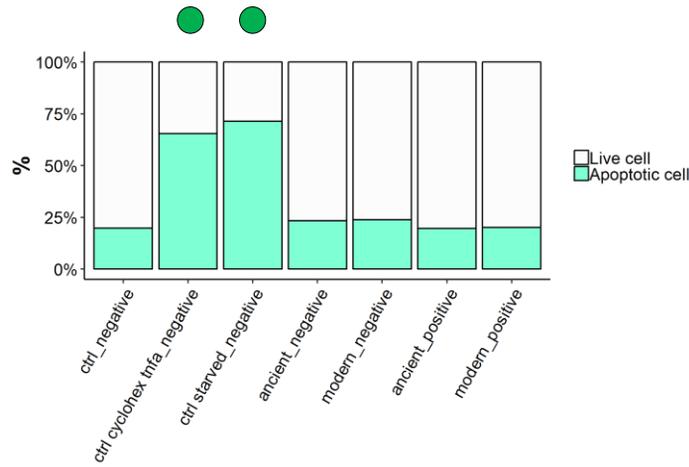
In M1 macrophages, bystander non-infected cells of modern strains showed increased apoptosis (40%). Despite not statistically significant and to a lesser extent, also macrophages infected with modern strains showed a similar increase in the percentage of apoptosis. Stratification of the data by mycobacterial burden showed similar apoptotic levels irrespective of the mycobacterial load in modern strains. Despite not statistically significant, also bystander non-infected cells of ancient strains with lower mycobacterial load showed a similar increase in the percentage of apoptosis.

- Apoptosis > not infected control
- Increased apoptosis observed, not statistically significant

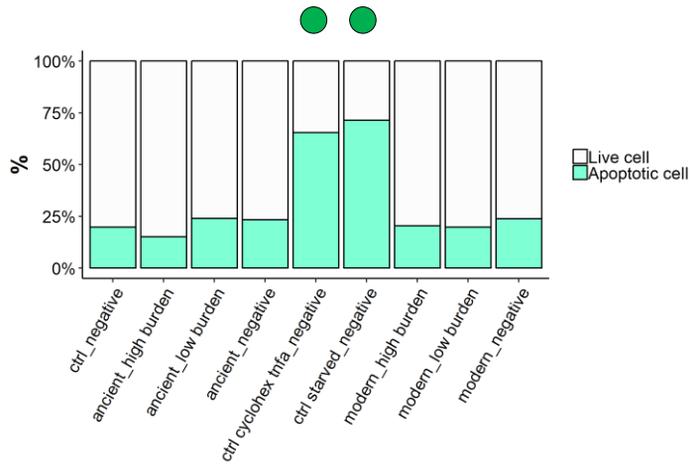
Grouping: Phylogeny

M2 – 24 h p.i.

a) Percentage of live/apoptotic cells



b) Percentage of live/apoptotic cells per mycobacterial burden



Par	Estimate	se	OR	pvalue
(Intercept)	-1.4534	0.4649	0.2338	0.0018
group_Mycctrl_cyclohex_tnfa_negative	1.6515	0.6339	5.215	0.0092
group_Mycctrl_starved_negative	1.9411	0.6855	6.9665	0.0046
group_Mycancient_negative	0.3881	0.4129	1.4741	0.3473
group_Mycmodern_negative	0.3686	0.3681	1.4458	0.3166
group_Mycancient_positive	-0.0272	0.4476	0.9732	0.9516
group_Mycmodern_positive	0.0892	0.3746	1.0933	0.8118

Contrast	estimate	SE	p.value
ctrl_negative-ctrl_cyclohex_tnfa_negative	-1.6515	0.6339	0.029
ctrl_negative-ctrl_starved_negative	-1.9411	0.6855	0.0269
ctrl_negative-ancient_negative	-0.3881	0.4129	0.4714
ctrl_negative-modern_negative	-0.3686	0.3681	0.4628
ctrl_negative-ancient_positive	0.0272	0.4476	0.9516
ctrl_negative-modern_positive	-0.0892	0.3746	0.9073
ctrl_cyclohex_tnfa_negative-ancient_negative	-0.2896	0.7965	0.8877
ctrl_cyclohex_tnfa_negative-modern_negative	1.2635	0.6016	0.0678
ctrl_cyclohex_tnfa_negative-modern_positive	1.2829	0.5717	0.0524
ctrl_cyclohex_tnfa_negative-ancient_positive	1.6787	0.6233	0.0269
ctrl_cyclohex_tnfa_negative-modern_positive	1.5623	0.576	0.0269
ctrl_starved_negative-ancient_negative	1.5531	0.6557	0.0424
ctrl_starved_negative-modern_negative	1.5725	0.6284	0.0335
ctrl_starved_negative-ancient_positive	1.9683	0.6758	0.0269
ctrl_starved_negative-modern_positive	1.8519	0.6323	0.0269
ancient_negative-modern_negative	0.0194	0.31	0.9516
ancient_positive-modern_positive	-0.1164	0.3616	0.8877
ancient_negative-ancient_positive	0.4152	0.3227	0.3138
modern_negative-modern_positive	0.2795	0.1948	0.2616

Par	Estimate	se	OR	pvalue
(Intercept)	-1.4594	0.4738	0.2324	0.0021
group_Myc3ancient_high_burden	-0.5157	0.5546	0.5971	0.3524
group_Myc3ancient_low_burden	0.3953	0.5042	1.4849	0.433
group_Myc3ancient_negative	0.3957	0.4211	1.4855	0.3474
group_Myc3ctrl_cyclohex_tnfa_negative	1.6502	0.6441	5.2079	0.0104
group_Myc3ctrl_starved_negative	1.9402	0.6952	6.9604	0.0053
group_Myc3modern_high_burden	0.1666	0.4136	1.1813	0.6871
group_Myc3modern_low_burden	0.034	0.4046	1.0346	0.933
group_Myc3modern_negative	0.3751	0.3755	1.4552	0.3178

OBSERVATIONS

In contrast, none of the category induced apoptosis in M2a macrophages at the time point considered.

● Apoptosis > not infected control

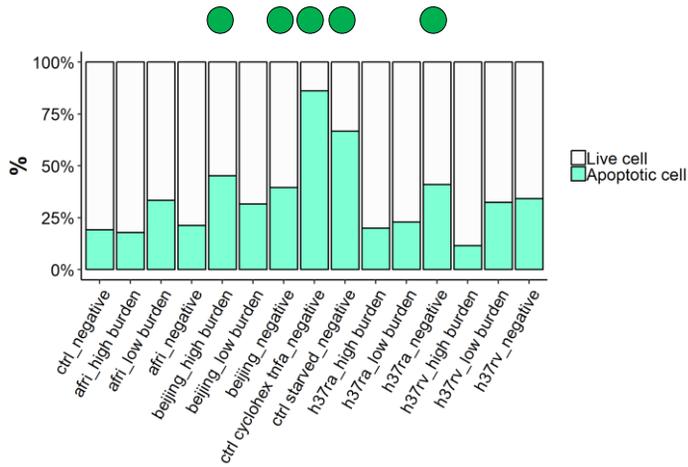
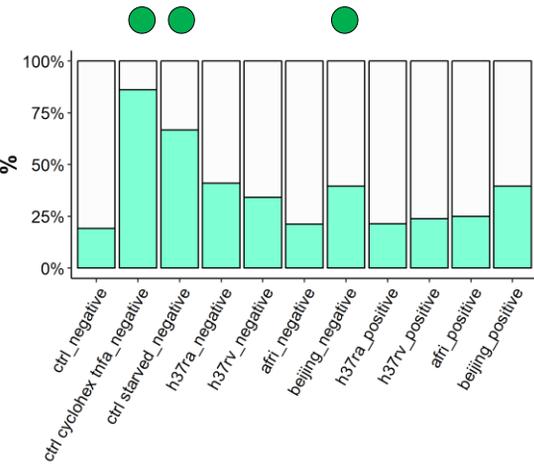
● Increased apoptosis observed, not statistically significant

Grouping: Sample

M1 – 24 h.p.i.

a) Percentage of live/apoptotic cells

b) Percentage of live/apoptotic cells per mycobacterial burden



Par	Estimate	se	OR	pvalue
(Intercept)	-1.2726	0.3691	0.2801	6e-04
group_Mycctrl_cyclohex_tnfa_negative	2.8169	0.6296	16.7247	0
group_Mycctrl_starved_negative	1.6948	0.576	5.4457	0.0033
group_Mych37ra_negative	0.9373	0.4135	2.553	0.0234
group_Mych37rv_negative	0.7326	0.4232	2.0805	0.0834
group_Mycafri_negative	0.0445	0.3715	1.0455	0.9047
group_Mybeijing_negative	1.0439	0.367	2.8402	0.0045
group_Mych37ra_positive	-0.0568	0.4554	0.9448	0.9008
group_Mych37rv_positive	0.1894	0.4636	1.2086	0.6828
group_Mycafri_positive	0.1552	0.3872	1.1679	0.6886
group_Mybeijing_positive	0.8104	0.3706	2.2489	0.0288

Par	Estimate	se	OR	pvalue
(Intercept)	-1.2712	0.3719	0.2805	6e-04
group_Myc3afri_high_burden	-0.315	0.4725	0.7298	0.505
group_Myc3afri_low_burden	0.5803	0.4445	1.7867	0.1917
group_Myc3afri_negative	0.032	0.3843	1.0326	0.9336
group_Myc3beijing_high_burden	1.0956	0.409	2.991	0.0074
group_Myc3beijing_low_burden	0.3759	0.4518	1.4563	0.4054
group_Myc3beijing_negative	1.051	0.3801	2.8606	0.0057
group_Myc3ctrl_cyclohex_tnfa_negative	2.8327	0.6446	16.9909	0
group_Myc3ctrl_starved_negative	1.7119	0.5924	5.5396	0.0039
group_Myc3h37ra_high_burden	-0.1092	0.56	0.8965	0.8454
group_Myc3h37ra_low_burden	-0.0101	0.5521	0.9899	0.9854
group_Myc3h37ra_negative	0.9303	0.4284	2.5352	0.0299
group_Myc3h37rv_high_burden	-0.5856	0.7226	0.5568	0.4178
group_Myc3h37rv_low_burden	0.5727	0.5218	1.7731	0.2723
group_Myc3h37rv_negative	0.7139	0.438	2.0419	0.1032

Contrast	estimate	SE	p.value
ctrl_negative-ctrl_cyclohex_tnfa_negative	-2.8169	0.6296	1e-04
ctrl_negative-ctrl_starved_negative	-1.6948	0.576	0.0116
ctrl_negative-h37ra_negative	-0.9373	0.4135	0.053
ctrl_negative-h37rv_negative	-0.7326	0.4232	0.1494
ctrl_negative-afri_negative	-0.0445	0.3715	0.9263
ctrl_negative-beijing_negative	-1.0439	0.367	0.0137
ctrl_negative-h37ra_positive	0.0568	0.4554	0.9263
ctrl_negative-h37rv_positive	-0.1894	0.4636	0.7666
ctrl_negative-afri_positive	-0.1552	0.3872	0.7666
ctrl_negative-beijing_positive	-0.8104	0.3706	0.0618
ctrl_cyclohex_tnfa_negative-ctrl_starved_negative	1.1221	0.7278	0.1961
ctrl_cyclohex_tnfa_negative-h37ra_negative	1.8796	0.6255	0.0114
ctrl_cyclohex_tnfa_negative-h37rv_negative	2.0843	0.634	0.0062
ctrl_cyclohex_tnfa_negative-afri_negative	2.7724	0.5987	1e-04
ctrl_cyclohex_tnfa_negative-beijing_negative	1.773	0.599	0.0116
ctrl_cyclohex_tnfa_negative-h37ra_positive	2.8737	0.6516	1e-04
ctrl_cyclohex_tnfa_negative-h37rv_positive	2.6274	0.6601	6e-04
ctrl_cyclohex_tnfa_negative-afri_positive	2.6617	0.6064	1e-04
ctrl_cyclohex_tnfa_negative-beijing_positive	2.0064	0.5936	0.0052
ctrl_starved_negative-h37ra_negative	0.7575	0.5714	0.2651
ctrl_starved_negative-h37rv_negative	0.9622	0.5807	0.164
ctrl_starved_negative-afri_negative	1.6504	0.5419	0.011
ctrl_starved_negative-beijing_negative	0.6509	0.5423	0.3191
ctrl_starved_negative-h37ra_positive	1.7516	0.5999	0.0116
ctrl_starved_negative-h37rv_positive	1.5054	0.6091	0.034
ctrl_starved_negative-afri_positive	1.5396	0.5505	0.0148
ctrl_starved_negative-beijing_positive	0.8844	0.5364	0.164
h37ra_negative-h37rv_negative	0.2047	0.4185	0.7463
h37ra_negative-afri_negative	0.8928	0.3661	0.0352
h37ra_negative-beijing_negative	-0.1066	0.3618	0.8259
h37rv_negative-afri_negative	0.6881	0.3771	0.1272
h37rv_negative-beijing_negative	-0.3113	0.3724	0.5254
afri_negative-beijing_negative	0.9994	0.3129	0.0075
h37ra_positive-h37rv_positive	-0.2462	0.4972	0.7463
h37ra_positive-afri_positive	-0.212	0.4263	0.7463
h37ra_positive-beijing_positive	-0.8672	0.4106	0.071
h37rv_positive-afri_positive	0.0343	0.4356	0.9373
h37rv_positive-beijing_positive	-0.621	0.4208	0.215
afri_positive-beijing_positive	-0.6553	0.3337	0.0969
h37ra_negative-h37ra_positive	0.9941	0.3754	0.0217
h37rv_negative-h37rv_positive	0.5432	0.3864	0.2369
afri_negative-afri_positive	-0.1107	0.2827	0.7666
beijing_negative-beijing_positive	0.2334	0.2607	0.4981

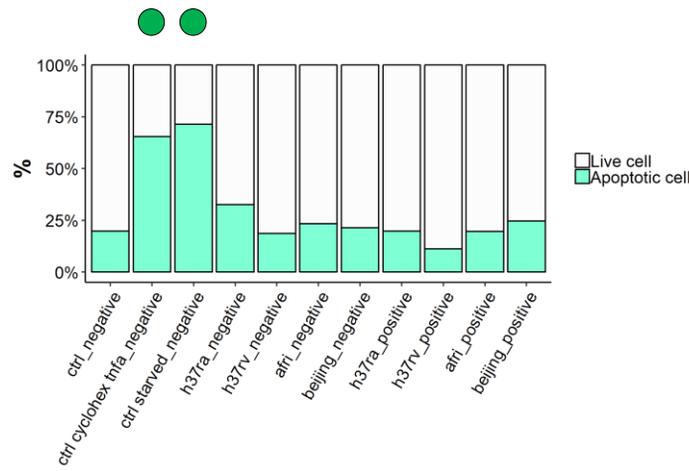
● Apoptosis > not infected control

● Increased apoptosis observed, not statistically significant

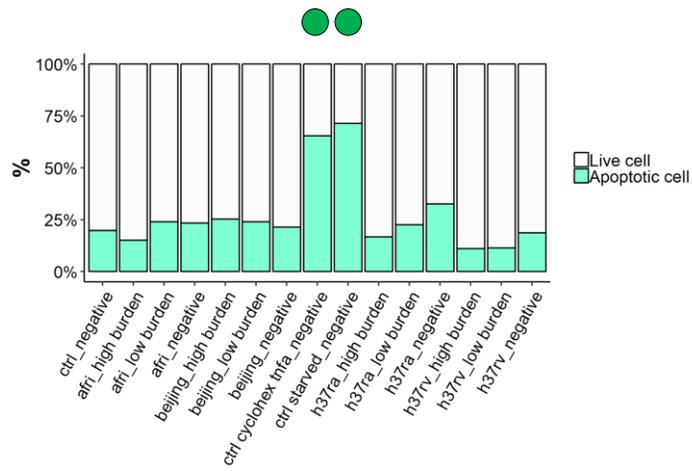
Grouping: Sample

M2 – 24 h.p.i.

a) Percentage of live/apoptotic cells



b) Percentage of live/apoptotic cells per mycobacterial burden



Par	Estimate	se	OR	pvalue
(Intercept)	-1.4301	0.4477	0.2393	0.0014
group_Mycctrl cyclohex tnfα negative	1.626	0.5984	5.0835	0.0066
group_Mycctrl starved negative	1.9138	0.6521	6.7791	0.0033
group_Myc37ra negative	0.7735	0.428	2.1674	0.0707
group_Myc37rv negative	-0.0049	0.4601	0.9951	0.9915
group_Mycafri negative	0.3668	0.3852	1.4432	0.341
group_Mycbeijing negative	0.2543	0.3839	1.2896	0.5077
group_Myc37ra positive	0.0671	0.4716	1.0694	0.8869
group_Myc37rv positive	-0.5298	0.5107	0.5887	0.2995
group_Mycafri positive	-0.0332	0.4215	0.9674	0.9373
group_Mycbeijing positive	0.31	0.3841	1.3634	0.4197

Par	Estimate	se	OR	pvalue
(Intercept)	-1.4382	0.456	0.2373	0.0016
group_Myc3afri high burden	-0.5063	0.5331	0.6027	0.3422
group_Myc3afri low burden	0.3776	0.4817	1.4588	0.4332
group_Myc3afri negative	0.3765	0.3951	1.4571	0.3407
group_Myc3beijing high burden	0.4025	0.4347	1.4956	0.3545
group_Myc3beijing low burden	0.2268	0.4393	1.2546	0.6056
group_Myc3beijing negative	0.2645	0.3935	1.3028	0.5014
group_Myc3ctrl cyclohex tnfα negative	1.6302	0.6108	5.1051	0.0076
group_Myc3ctrl starved negative	1.9187	0.6637	6.8121	0.0038
group_Myc3h37ra high burden	-0.0443	0.5994	0.9567	0.9411
group_Myc3h37ra low burden	0.1597	0.5445	1.1731	0.7693
group_Myc3h37ra negative	0.782	0.4392	2.1859	0.075
group_Myc3h37rv high burden	-0.4505	0.7279	0.6373	0.536
group_Myc3h37rv low burden	-0.5554	0.577	0.5738	0.3358
group_Myc3h37rv negative	0.004	0.4704	1.004	0.9932

Contrast	estimate	SE	p.value
ctrl_negative-ctrl cyclohex tnfα negative	-1.626	0.5984	0.0369
ctrl_negative-ctrl starved negative	-1.9138	0.6521	0.0358
ctrl_negative-h37ra_negative	-0.7735	0.428	0.1689
ctrl_negative-h37rv_negative	0.0049	0.4601	0.9915
ctrl_negative-afri_negative	-0.3668	0.3852	0.4887
ctrl_negative-beijing_negative	-0.2543	0.3839	0.642
ctrl_negative-h37ra_positive	-0.0671	0.4716	0.9302
ctrl_negative-h37rv_positive	0.5298	0.5107	0.46
ctrl_negative-afri_positive	0.0332	0.4215	0.9596
ctrl_negative-beijing_positive	-0.31	0.3841	0.5468
ctrl cyclohex tnfα negative-ctrl starved negative	-0.2879	0.7613	0.8197
ctrl cyclohex tnfα negative-h37ra negative	0.8524	0.6003	0.3042
ctrl cyclohex tnfα negative-h37rv negative	1.6309	0.6232	0.0381
ctrl cyclohex tnfα negative-afri negative	1.2592	0.5712	0.0739
ctrl cyclohex tnfα negative-beijing negative	1.3716	0.5708	0.0499
ctrl cyclohex tnfα negative-h37ra_positive	1.5589	0.6322	0.0452
ctrl cyclohex tnfα negative-h37rv_positive	2.1558	0.6634	0.0248
ctrl cyclohex tnfα negative-afri_positive	1.6592	0.5936	0.0369
ctrl cyclohex tnfα negative-beijing_positive	1.3116	0.57	0.0601
ctrl starved negative-h37ra negative	1.1403	0.6539	0.1745
ctrl starved negative-h37rv_negative	1.9187	0.6749	0.0369
ctrl starved negative-afri_negative	1.547	0.6272	0.0452
ctrl starved negative-beijing_negative	1.6595	0.6268	0.0381
ctrl starved negative-h37ra_positive	1.8468	0.6832	0.0369
ctrl starved negative-h37rv_positive	2.4436	0.7122	0.0248
ctrl starved negative-afri_positive	1.947	0.6477	0.0358
ctrl starved negative-beijing_positive	1.6039	0.6261	0.0407
h37ra_negative-h37rv_negative	0.7784	0.4633	0.1903
h37ra_negative-afri_negative	0.4067	0.3892	0.46
h37ra_negative-beijing_negative	0.5192	0.3879	0.3379
h37rv_negative-afri_negative	-0.3717	0.4242	0.5117
h37rv_negative-beijing_negative	-0.2592	0.423	0.6634
afri_negative-beijing_negative	0.1125	0.3401	0.8383
h37ra_positive-h37rv_positive	0.5969	0.5506	0.46
h37ra_positive-afri_positive	0.1002	0.4689	0.8994
h37ra_positive-beijing_positive	-0.2429	0.4357	0.6894
h37rv_positive-afri_positive	-0.4966	0.5084	0.4873
h37rv_positive-beijing_positive	-0.8398	0.4778	0.1745
afri_positive-beijing_positive	-0.3432	0.3808	0.5097
h37ra_negative-h37ra_positive	0.7065	0.374	0.1489
h37rv_negative-h37rv_positive	0.5249	0.4546	0.4269
afri_negative-afri_positive	0.4	0.3207	0.3803
beijing_negative-beijing_positive	-0.0556	0.2698	0.8994

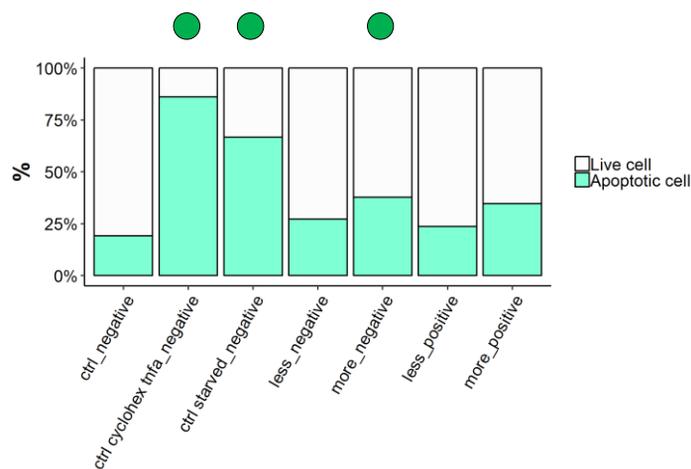
● Apoptosis > not infected control

● Increased apoptosis observed, not statistically significant

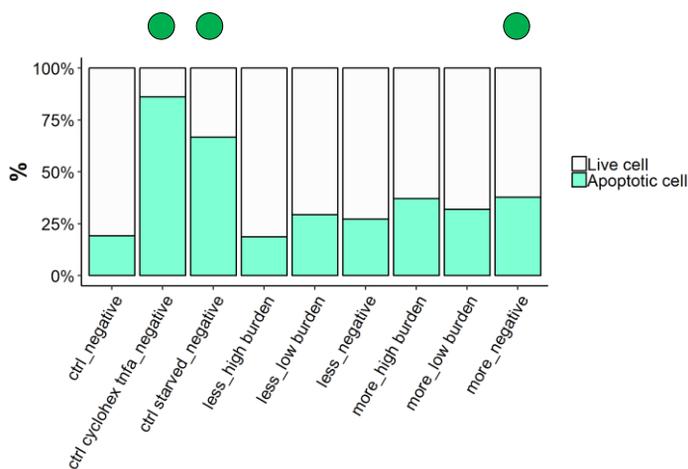
Grouping: Virulence (absolute)

M1 – 24 h.p.i.

a) Percentage of live/apoptotic cells



b) Percentage of live/apoptotic cells per mycobacterial burden



Par	Estimate	se	OR	pvalue
(Intercept)	-1.2651	0.3868	0.2822	0.0011
group_Mycctrl_cyclohex_tnfa_negative	2.8053	0.6554	16.5318	0
group_Mycctrl_starved_negative	1.6854	0.6042	5.3948	0.0053
group_Myeless_negative	0.3578	0.3645	1.4302	0.3262
group_Myemore_negative	0.942	0.3651	2.5651	0.0099
group_Myeless_positive	0.0559	0.3786	1.0575	0.8827
group_Myemore_positive	0.6232	0.3709	1.8648	0.0929

Par	Estimate	se	OR	pvalue
(Intercept)	-1.2649	0.3877	0.2823	0.0011
group_Myc3ctrl_cyclohex_tnfa_negative	2.8168	0.6641	16.7232	0
group_Myc3ctrl_starved_negative	1.6977	0.6138	5.4611	0.0057
group_Myc3less_high_burden	-0.267	0.4323	0.7657	0.5368
group_Myc3less_low_burden	0.3554	0.418	1.4267	0.3952
group_Myc3less_negative	0.3494	0.3714	1.4182	0.3468
group_Myc3more_high_burden	0.7701	0.4028	2.16	0.0559
group_Myc3more_low_burden	0.448	0.4171	1.5652	0.2828
group_Myc3more_negative	0.9438	0.3721	2.5697	0.0112

Contrast	estimate	SE	p.value
ctrl_negative-ctrl_cyclohex_tnfa_negative	-2.8053	0.6554	2e-04
ctrl_negative-ctrl_starved_negative	-1.6854	0.6042	0.0143
ctrl_negative-less_negative	-0.3578	0.3645	0.3443
ctrl_negative-more_negative	-0.942	0.3651	0.0235
ctrl_negative-less_positive	-0.0559	0.3786	0.8827
ctrl_negative-more_positive	-0.6232	0.3709	0.1358
ctrl_cyclohex_tnfa_negative-ctrl_starved_negative	1.1198	0.7567	0.178
ctrl_cyclohex_tnfa_negative-less_negative	2.4475	0.6039	3e-04
ctrl_cyclohex_tnfa_negative-more_negative	1.8633	0.6088	0.0081
ctrl_cyclohex_tnfa_negative-less_positive	2.7494	0.6102	1e-04
ctrl_cyclohex_tnfa_negative-more_positive	2.1821	0.6048	0.0015
ctrl_starved_negative-less_negative	1.3276	0.5479	0.0325
ctrl_starved_negative-more_negative	0.7435	0.5512	0.2013
ctrl_starved_negative-less_positive	1.6296	0.5549	0.0105
ctrl_starved_negative-more_positive	1.0623	0.549	0.0839
less_negative-more_negative	-0.5842	0.2638	0.0509
less_positive-more_positive	-0.5673	0.289	0.0839
less_negative-less_positive	0.3019	0.2253	0.2013
more_negative-more_positive	0.3188	0.2163	0.178

OBSERVATIONS

In M1 macrophages, bystander non-infected cells of more virulent strains showed increased apoptosis (40%). Despite not statistically significant, also macrophages infected with more virulent strains showed a similar increase in the percentage of apoptosis. Stratification of the data by mycobacterial burden showed similar apoptotic levels irrespective of the mycobacterial load in more virulent strains, whereas for H37Rv higher level of apoptosis were induced in bystander non-infected cells or in cells infected with lower mycobacterial load.

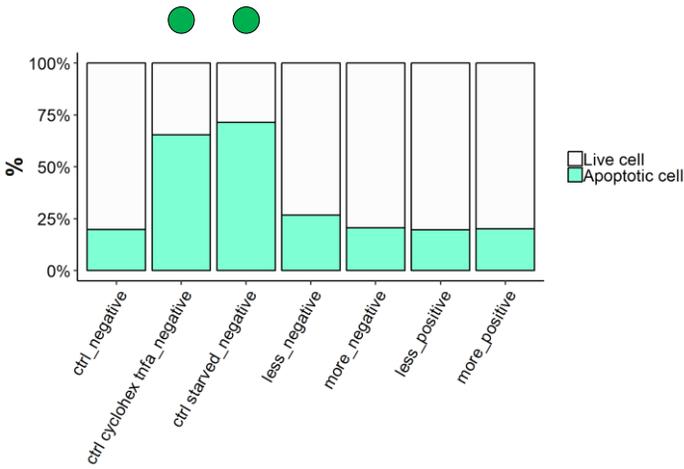
● Apoptosis > not infected control

● Increased apoptosis observed, not statistically significant

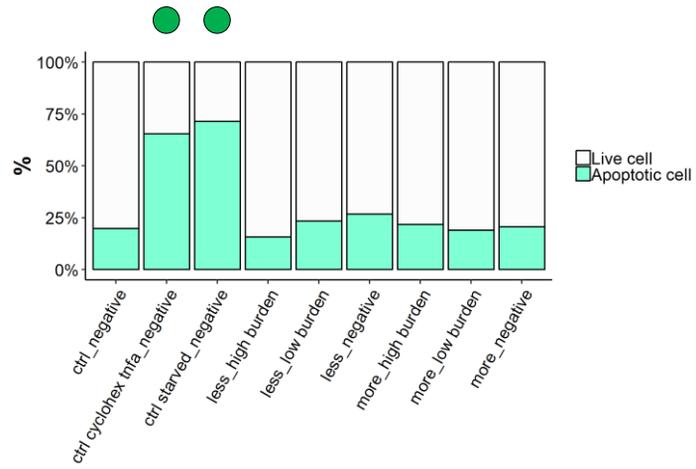
Grouping: Virulence (absolute)

M2 – 24 h.p.i.

a) Percentage of live/apoptotic cells



b) Percentage of live/apoptotic cells per mycobacterial burden



Par	Estimate	se	OR	pvalue
(Intercept)	-1.4496	0.4623	0.2347	0.0017
group_Mycctrl_cyclohex_tnfa_negative	1.6467	0.628	5.1897	0.0087
group_Mycctrl_starved_negative	1.936	0.6799	6.9308	0.0044
group_Myccless_negative	0.54	0.3768	1.716	0.1518
group_Mycmore_negative	0.1892	0.3811	1.2083	0.6196
group_Myccless_positive	0.0013	0.3991	1.0013	0.9974
group_Mycmore_positive	0.0888	0.385	1.0929	0.8176

Contrast	estimate	SE	p-value
ctrl_negative-ctrl_cyclohex_tnfa_negative	-1.6467	0.628	0.0237
ctrl_negative-ctrl_starved_negative	-1.936	0.6799	0.0224
ctrl_negative-less_negative	-0.54	0.3768	0.2404
ctrl_negative-more_negative	-0.1892	0.3811	0.8392
ctrl_negative-less_positive	-0.0013	0.3991	0.9974
ctrl_negative-more_positive	-0.0888	0.385	0.863
ctrl_cyclohex_tnfa_negative-ctrl_starved_negative	-0.2893	0.7907	0.8484
ctrl_cyclohex_tnfa_negative-less_negative	1.1067	0.5752	0.0939
ctrl_cyclohex_tnfa_negative-more_negative	1.4575	0.5783	0.0279
ctrl_cyclohex_tnfa_negative-less_positive	1.6454	0.5885	0.0224
ctrl_cyclohex_tnfa_negative-more_positive	1.5579	0.5809	0.0232
ctrl_starved_negative-less_negative	1.396	0.6315	0.0523
ctrl_starved_negative-more_negative	1.7468	0.6344	0.0224
ctrl_starved_negative-less_positive	1.9347	0.6436	0.0224
ctrl_starved_negative-more_positive	1.8472	0.6367	0.0224
less_negative-more_negative	0.3508	0.2869	0.3237
less_positive-more_positive	-0.0875	0.3203	0.863
less_negative-less_positive	0.5387	0.2444	0.0523
more_negative-more_positive	0.1004	0.23	0.8392

Par	Estimate	se	OR	pvalue
(Intercept)	-1.4542	0.4666	0.2336	0.0018
group_Myc3ctrl_cyclohex_tnfa_negative	1.6503	0.6353	5.2083	0.0094
group_Myc3ctrl_starved_negative	1.9399	0.6868	6.958	0.0047
group_Myc3less_high_burden	-0.3293	0.4683	0.7195	0.482
group_Myc3less_low_burden	0.2742	0.4368	1.3154	0.5302
group_Myc3less_negative	0.546	0.3823	1.7263	0.1532
group_Myc3more_high_burden	0.2189	0.4285	1.2447	0.6094
group_Myc3more_low_burden	-0.0136	0.4205	0.9864	0.9741
group_Myc3more_negative	0.1952	0.3864	1.2156	0.6134

OBSERVATIONS

In contrast, none of the category induced apoptosis in M2a macrophages at the time point considered.

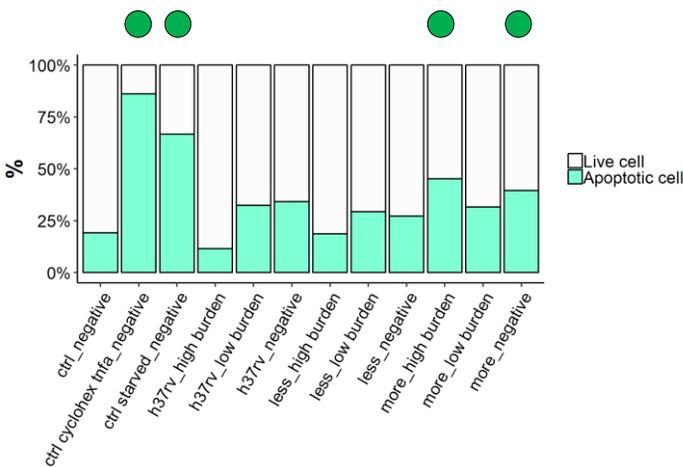
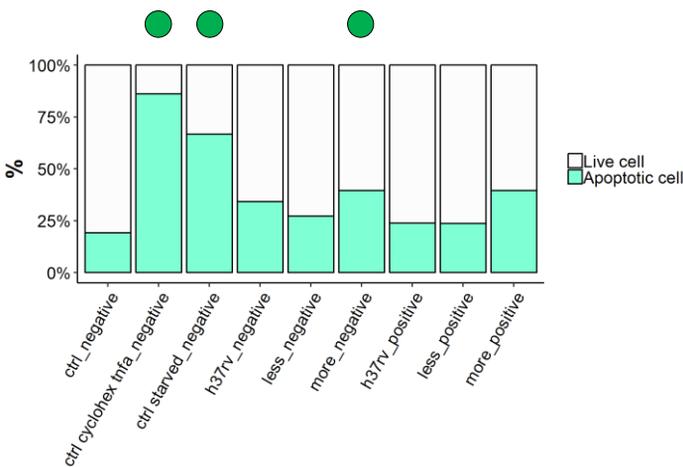
- Apoptosis > not infected control
- Increased apoptosis observed, not statistically significant

Grouping: Virulence (relative)

M1 – 24 h p.i.

a) Percentage of live/apoptotic cells

b) Percentage of live/apoptotic cells per mycobacterial burden



Par	Estimate	se	OR	pvalue
(Intercept)	-1.269	0.377	0.2811	8e-04
group_Mycctrl_cyclohex_tnfa_negative	2.8168	0.6446	16.7231	0
group_Mycctrl_starved_negative	1.696	0.5924	5.4523	0.0042
group_Myc37rv_negative	0.7271	0.438	2.0691	0.0969
group_Myc3less_negative	0.3627	0.3559	1.4372	0.3082
group_Mycmore_negative	1.0425	0.38	2.8362	0.0061
group_Myc37rv_positive	0.1862	0.4775	1.2047	0.6965
group_Myc3less_positive	0.0636	0.3703	1.0657	0.8636
group_Myc3more_positive	0.8085	0.3832	2.2445	0.0349

Par	Estimate	se	OR	pvalue
(Intercept)	-1.2688	0.3779	0.2812	8e-04
group_Myc3ctrl_cyclohex_tnfa_negative	2.8409	0.6602	17.1315	0
group_Myc3ctrl_starved_negative	1.7215	0.6095	5.5928	0.0047
group_Myc3h37rv_high_burden	-0.5929	0.7325	0.5527	0.4182
group_Myc3h37rv_low_burden	0.5753	0.5357	1.7776	0.2829
group_Myc3h37rv_negative	0.7079	0.4535	2.0298	0.1185
group_Myc3less_high_burden	-0.2621	0.4296	0.7694	0.5418
group_Myc3less_low_burden	0.3617	0.4152	1.4357	0.3838
group_Myc3less_negative	0.3518	0.3685	1.4217	0.3396
group_Myc3more_high_burden	1.0983	0.4214	2.999	0.0092
group_Myc3more_low_burden	0.3705	0.4635	1.4485	0.424
group_Myc3more_negative	1.0504	0.3935	2.8588	0.0076

Contrast	estimate	SE	p.value
ctrl_negative-ctrl_cyclohex_tnfa_negative	-2.8168	0.6446	2e-04
ctrl_negative-ctrl_starved_negative	-1.696	0.5924	0.014
ctrl_negative-h37rv_negative	-0.7271	0.438	0.1791
ctrl_negative-less_negative	-0.3627	0.3559	0.377
ctrl_negative-more_negative	-1.0425	0.38	0.0182
ctrl_negative-h37rv_positive	-0.1862	0.4775	0.7463
ctrl_negative-less_positive	-0.0636	0.3703	0.8636
ctrl_negative-more_positive	-0.8085	0.3832	0.0697
ctrl_cyclohex_tnfa_negative-ctrl_starved_negative	1.1208	0.7446	0.2088
ctrl_cyclohex_tnfa_negative-h37rv_negative	2.0897	0.649	0.0064
ctrl_cyclohex_tnfa_negative-less_negative	2.4541	0.5946	4e-04
ctrl_cyclohex_tnfa_negative-more_negative	1.7743	0.6123	0.014
ctrl_cyclohex_tnfa_negative-h37rv_positive	2.6306	0.6748	7e-04
ctrl_cyclohex_tnfa_negative-less_positive	2.7532	0.601	1e-04
ctrl_cyclohex_tnfa_negative-more_positive	2.0083	0.607	0.0056
ctrl_starved_negative-h37rv_negative	0.9689	0.5972	0.1791
ctrl_starved_negative-less_negative	1.3334	0.5376	0.0358
ctrl_starved_negative-more_negative	0.6536	0.5571	0.314
ctrl_starved_negative-h37rv_positive	1.5098	0.6252	0.0368
ctrl_starved_negative-less_positive	1.6324	0.5447	0.0117
ctrl_starved_negative-more_positive	0.8875	0.5513	0.1791
h37rv_negative-less_negative	0.3644	0.3621	0.377
h37rv_negative-more_negative	-0.3154	0.3855	0.4593
less_negative-more_negative	-0.6798	0.2892	0.0402
h37rv_positive-less_positive	0.1226	0.4214	0.7977
h37rv_positive-more_positive	-0.6223	0.4327	0.2256
less_positive-more_positive	-0.7449	0.309	0.0368
h37rv_negative-h37rv_positive	0.5409	0.3871	0.2318
less_negative-less_positive	0.299	0.2248	0.2502
more_negative-more_positive	0.234	0.2621	0.4293

OBSERVATIONS

In M1 macrophages, bystander non-infected cells of more virulent strains showed increased apoptosis (40%). Despite not statistically significant, also macrophages infected with more virulent strains showed a similar increase in the percentage of apoptosis. To a lesser extent, also H37Rv induced increased apoptosis of bystander non-infected cells (35%), but this was not found to be statistically significant. Stratification of the data by mycobacterial burden showed similar apoptotic levels irrespective of the mycobacterial load in more virulent strains.

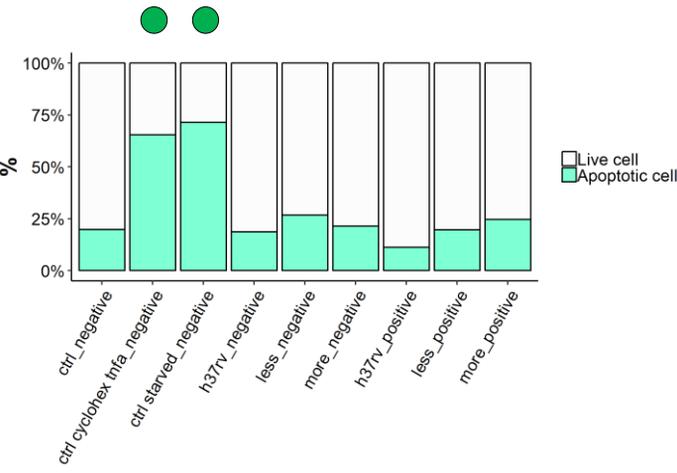
● Apoptosis > not infected control

● Increased apoptosis observed, not statistically significant

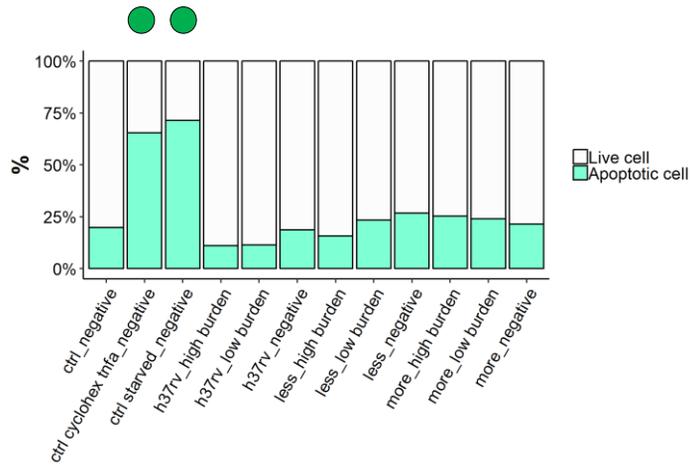
Grouping: Virulence (relative)

M2 – 24 h p.i.

a) Percentage of live/apoptotic cells



b) Percentage of live/apoptotic cells per mycobacterial burden



Par	Estimate	se	OR	pvalue
(Intercept)	-1.435	0.4514	0.2381	0.0015
group_Mycctrl_cyclohex_tnfa_negative	1.6307	0.6055	5.1077	0.0071
group_Mycctrl_starved_negative	1.919	0.6588	6.8139	0.0036
group_Mych37rv_negative	5e-04	0.4661	1.0005	0.9992
group_Mycleless_negative	0.5266	0.3599	1.6931	0.1434
group_Myemore_negative	0.2595	0.3895	1.2962	0.5053
group_Mych37rv_positive	-0.5252	0.5162	0.5914	0.3089
group_Mycleless_positive	-0.0036	0.3829	0.9964	0.9924
group_Myemore_positive	0.3132	0.3897	1.3678	0.4216

Par	Estimate	se	OR	pvalue
(Intercept)	-1.4407	0.4557	0.2368	0.0016
group_Myc3ctrl_cyclohex_tnfa_negative	1.6369	0.6143	5.1395	0.0077
group_Myc3ctrl_starved_negative	1.9256	0.667	6.8592	0.0039
group_Myc3h37rv_low_burden	-0.4493	0.73	0.6381	0.5382
group_Myc3h37rv_high_burden	-0.5531	0.5795	0.5752	0.3398
group_Myc3h37rv_negative	0.0068	0.4734	1.0068	0.9885
group_Myc3less_high_burden	-0.3285	0.4548	0.72	0.4702
group_Myc3less_low_burden	0.2653	0.4227	1.3038	0.5303
group_Myc3less_negative	0.5336	0.3665	1.7051	0.1454
group_Myc3more_high_burden	0.4044	0.4372	1.4983	0.355
group_Myc3more_low_burden	0.2283	0.4418	1.2565	0.6053
group_Myc3more_negative	0.267	0.3962	1.306	0.5005

Contrast	estimate	SE	p-value
ctrl_negative-ctrl_cyclohex_tnfa_negative	-1.6307	0.6055	0.0303
ctrl_negative-ctrl_starved_negative	-1.919	0.6588	0.0243
ctrl_negative-h37rv_negative	-5e-04	0.4661	0.9992
ctrl_negative-less_negative	-0.5266	0.3599	0.2531
ctrl_negative-more_negative	-0.2595	0.3895	0.6064
ctrl_negative-h37rv_positive	0.5252	0.5162	0.4413
ctrl_negative-less_positive	0.0036	0.3829	0.9992
ctrl_cyclohex_tnfa_negative-ctrl_starved_negative	-0.3132	0.3897	0.527
ctrl_cyclohex_tnfa_negative-ctrl_starved_negative	-0.2882	0.7684	0.7862
ctrl_cyclohex_tnfa_negative-h37rv_negative	1.6303	0.6298	0.0321
ctrl_cyclohex_tnfa_negative-less_negative	1.1042	0.5566	0.0946
ctrl_cyclohex_tnfa_negative-more_negative	1.3713	0.5769	0.0476
ctrl_cyclohex_tnfa_negative-h37rv_positive	2.1559	0.6697	0.0193
ctrl_cyclohex_tnfa_negative-less_positive	1.6344	0.5702	0.0243
ctrl_cyclohex_tnfa_negative-more_positive	1.3176	0.5761	0.054
ctrl_starved_negative-h37rv_negative	1.9185	0.6812	0.0243
ctrl_starved_negative-less_negative	1.3924	0.6142	0.054
ctrl_starved_negative-more_negative	1.6595	0.6326	0.0321
ctrl_starved_negative-h37rv_positive	2.4442	0.7182	0.0193
ctrl_starved_negative-less_positive	1.9226	0.6265	0.0215
ctrl_starved_negative-more_positive	1.6058	0.6319	0.0331
h37rv_negative-less_negative	-0.5261	0.4009	0.3158
h37rv_negative-more_negative	-0.259	0.4277	0.6287
less_negative-more_negative	0.2671	0.3086	0.5045
h37rv_positive-less_positive	-0.5216	0.4767	0.4108
h37rv_positive-more_positive	-0.8384	0.482	0.1537
less_positive-more_positive	-0.3168	0.3353	0.4702
h37rv_negative-h37rv_positive	0.5257	0.4546	0.3908
less_negative-less_positive	0.5302	0.2436	0.0633
more_negative-more_positive	-0.0537	0.27	0.9025

OBSERVATIONS

In contrast, none of the category induced apoptosis in M2a macrophages at the time point considered.

- Apoptosis > not infected control
- Increased apoptosis observed, not statistically significant

Immunofluorescence

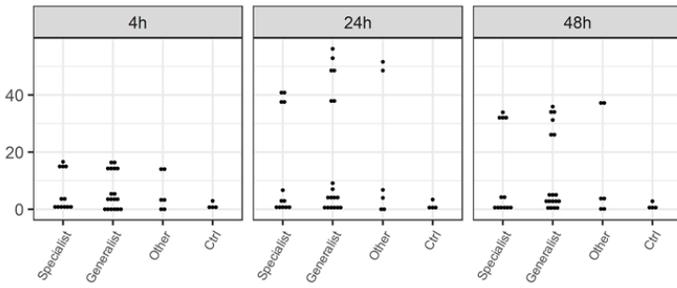
The levels of IFN γ and TNF α cytokines were evaluated in fixed samples at the same positions acquired in live imaging for measuring the acidification of the phagolysosomes by the use of LysoSensor Green. We considered THP-1 M1 and M2a macrophages infected with H37Rv, H37Ra, L6, and L2 at 24 h p.i. A total of 818 single cells (405 macrophages for LysoSensor + IFN γ ; 413 macrophages for LysoSensor + TNF α) were considered for the analysis. Data were normalized on THP-1 macrophages not infected control.

The levels of MIP-1 β and MCP-1 cytokines were evaluated in fixed samples. We considered THP-1 M1 and M2a macrophages infected with H37Rv, H37Ra, L6, and L2 at 24 h p.i. A total of 1268 single cells (652 macrophages for MIP-1 β ; 616 macrophages for MCP-1) were considered for the analysis. Data were normalized on THP-1 macrophages not infected control.

Grouping: Geo-host adaptation

M1 – IFN γ

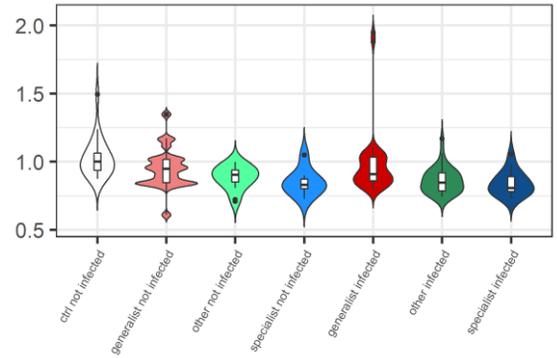
a) BioPlex (bulk)



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.218	0.4333	0.617
Time24h	0.2724	0.1579	0.0905
Time48h	0.0968	0.1579	0.5426
Geo_host_adaptationGeneralist	0.0992	0.1441	0.4944
Time24h:Geo_host_adaptationGeneralist	0.161	0.2038	0.4331
Time48h:Geo_host_adaptationGeneralist	0.1051	0.2038	0.6083

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.0992	0.1441	0.4944
Specialist - Generalist	24h	-0.2602	0.1441	0.0769
Specialist - Generalist	48h	-0.2043	0.1441	0.1624

b) Immunofluorescence (single-cell) – 24 h p.i.



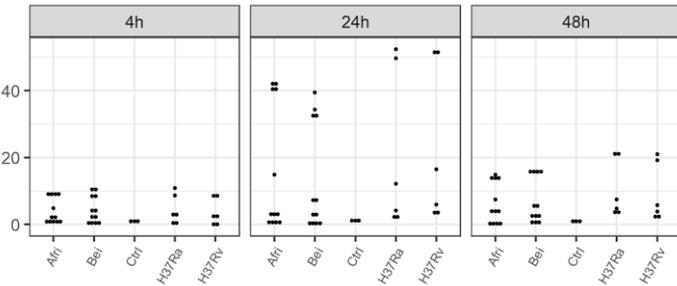
Linear mixed-effects model (orderNorm transformation)

Par	Value	Std.Error	p-value
(Intercept)	0.5686	0.4369	0.195
groupgeneralist not infected	-0.6214	0.2356	0.0092
groupother not infected	-0.9222	0.2623	6e-04
groupspecialist not infected	-0.7686	0.3654	0.037
groupgeneralist infected	-0.5032	0.2162	0.0212
groupother infected	-1.0982	0.2227	0
groupspecialist infected	-0.8599	0.2966	0.0043

Contrast	estimate	SE	p-value
ctrl not infected-generalist not infected	0.6214	0.2356	0.0276
ctrl not infected-other not infected	0.9222	0.2623	0.0043
ctrl not infected-specialist not infected	0.7686	0.3654	0.0793
ctrl not infected-generalist infected	0.5032	0.2162	0.053
ctrl not infected-other infected	1.0982	0.2227	0
ctrl not infected-specialist infected	0.8599	0.2966	0.016
generalist not infected-other not infected	0.3008	0.255	0.3998
generalist not infected-specialist not infected	0.1471	0.3631	0.7349
other not infected-specialist not infected	-0.1537	0.3684	0.7349
generalist infected-other infected	0.595	0.1903	0.0105
generalist infected-specialist infected	0.3567	0.2719	0.3591
other infected-specialist infected	-0.2383	0.2593	0.5392
generalist not infected-generalist infected	-0.1183	0.2061	0.7086
other not infected-other infected	0.1759	0.2378	0.6279
specialist not infected-specialist infected	0.0913	0.3557	0.7977

M2 – IFN γ

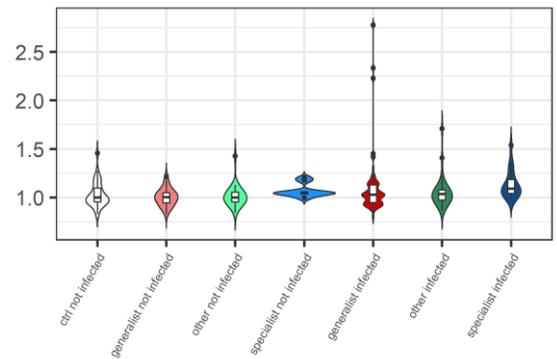
a) BioPlex (bulk)



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.2695	0.4404	0.5441
Time24h	0.626	0.1658	5e-04
Time48h	0.2761	0.1658	0.1037
TypeBei	0.0625	0.1658	0.7083
Time24h:TypeBei	-0.3011	0.2344	0.2066
Time48h:TypeBei	0.0456	0.2344	0.8467

contrast	Time	estimate	SE	p-value
Afri - Bei	4h	-0.0625	0.1658	0.7083
Afri - Bei	24h	0.2386	0.1658	0.158
Afri - Bei	48h	-0.1081	0.1658	0.5181

b) Immunofluorescence (single-cell) – 24 h p.i.



Linear mixed-effects model (orderNorm transformation)

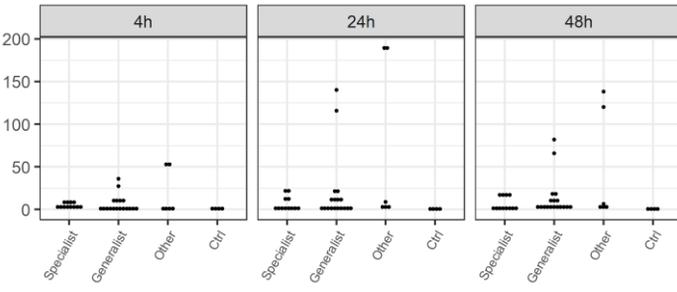
Par	Value	Std.Error	p-value
(Intercept)	-0.0978	0.3076	0.7508
groupgeneralist not infected	-0.0621	0.23	0.7875
groupother not infected	-0.1138	0.2514	0.6513
groupspecialist not infected	0.2591	0.3738	0.4889
groupgeneralist infected	0.2711	0.2111	0.2005
groupother infected	0.1776	0.2419	0.4637
groupspecialist infected	0.5534	0.2919	0.0593

Contrast	estimate	SE	p-value
ctrl not infected-generalist not infected	0.0621	0.23	0.832
ctrl not infected-other not infected	0.1138	0.2514	0.7646
ctrl not infected-specialist not infected	-0.2591	0.3738	0.6667
ctrl not infected-generalist infected	-0.2711	0.2111	0.6667
ctrl not infected-other infected	-0.1776	0.2419	0.6667
ctrl not infected-specialist infected	-0.5534	0.2919	0.5979
generalist not infected-other not infected	0.0517	0.2436	0.832
generalist not infected-specialist not infected	-0.3212	0.376	0.6667
other not infected-specialist not infected	-0.3729	0.382	0.6667
generalist infected-other infected	0.0936	0.2141	0.7646
generalist infected-specialist infected	-0.2822	0.2781	0.6667
other infected-specialist infected	-0.3758	0.2968	0.6667
generalist not infected-generalist infected	-0.3332	0.1892	0.5979
other not infected-other infected	-0.2914	0.243	0.6667
specialist not infected-specialist infected	-0.2942	0.3792	0.6667

Grouping: Geo-host adaptation

M1 – TNF α

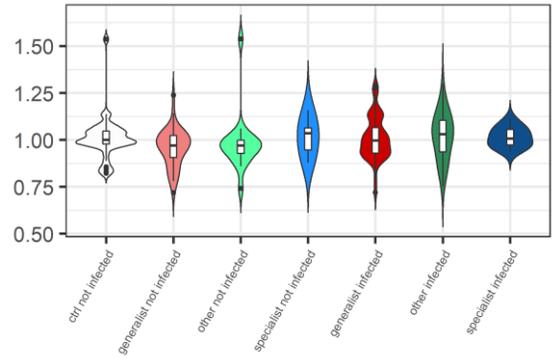
a) BioPlex (bulk)



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.2225	0.4461	0.62
Time24h	0.1489	0.2033	0.4672
Time48h	0.076	0.2033	0.71
Geo_host_adaptationGeneralist	0.1279	0.1856	0.4938
Time24h:Geo_host_adaptationGeneralist	0.1954	0.2625	0.46
Time48h:Geo_host_adaptationGeneralist	0.1498	0.2625	0.5706

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.1279	0.1856	0.4938
Specialist - Generalist	24h	-0.3233	0.1856	0.0875
Specialist - Generalist	48h	-0.2778	0.1856	0.1407

b) Immunofluorescence (single-cell) – 24 h p.i.



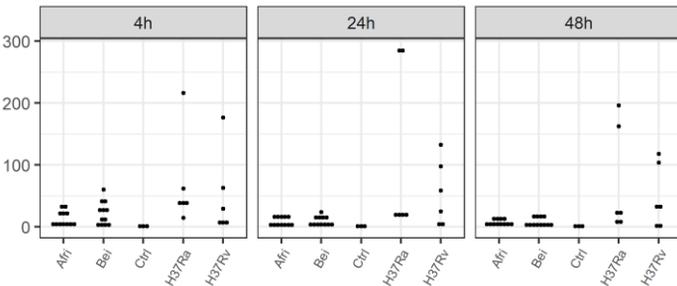
Linear mixed-effects model (orderNorm transformation)

Par	Value	Std.Error	p-value
(Intercept)	0.0802	0.201	0.6904
groupgeneralist not infected	-0.5285	0.2284	0.0218
groupother not infected	-0.3688	0.2884	0.2028
groupspecialist not infected	0.239	0.4778	0.6175
groupgeneralist infected	-0.0776	0.2232	0.7284
groupother infected	0.1729	0.2403	0.4729
groupspecialist infected	0.2155	0.3029	0.4776

Contrast	estimate	SE	p-value
ctrl not infected-generalist not infected	0.5285	0.2284	0.2631
ctrl not infected-other not infected	0.3688	0.2884	0.5795
ctrl not infected-specialist not infected	-0.239	0.4778	0.7718
ctrl not infected-generalist infected	0.0776	0.2232	0.8405
ctrl not infected-other infected	-0.1729	0.2403	0.7165
ctrl not infected-specialist infected	-0.2155	0.3029	0.7165
generalist not infected-other not infected	-0.1598	0.2907	0.7718
generalist not infected-specialist not infected	-0.7676	0.4873	0.4387
other not infected-specialist not infected	-0.6078	0.5066	0.5795
generalist infected-other infected	-0.2505	0.2393	0.6355
generalist infected-specialist infected	-0.2932	0.3132	0.6571
other infected-specialist infected	-0.0427	0.3056	0.9526
generalist not infected-generalist infected	-0.4509	0.2123	0.2631
other not infected-other infected	-0.5416	0.2968	0.3486
specialist not infected-specialist infected	0.0235	0.5003	0.9626

M2 – TNF α

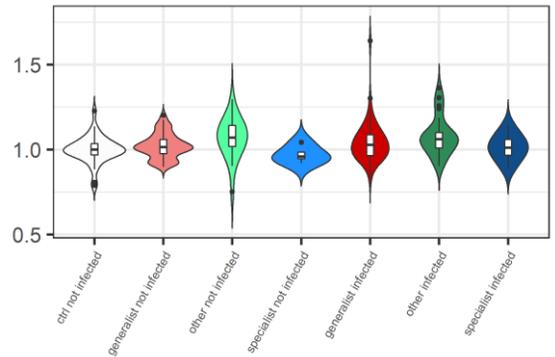
a) BioPlex (bulk)



Parameter	Estimate	Std.Error	p-value
(Intercept)	0.1898	0.4231	0.6562
Time24h	-0.4552	0.287	0.1208
Time48h	-0.6237	0.287	0.0359
TypeBei	0.3431	0.287	0.2391
Time24h:TypeBei	-0.2166	0.4059	0.5966
Time48h:TypeBei	-0.1413	0.4059	0.7296

contrast	Time	estimate	SE	p-value
Afri - Bei	4h	-0.3431	0.287	0.2391
Afri - Bei	24h	-0.1265	0.287	0.6619
Afri - Bei	48h	-0.2018	0.287	0.4862

b) Immunofluorescence (single-cell) – 24 h p.i.



Linear mixed-effects model (orderNorm transformation)

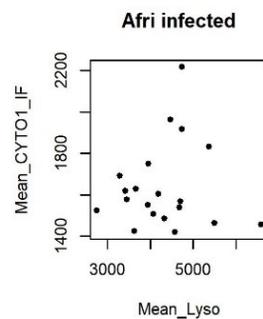
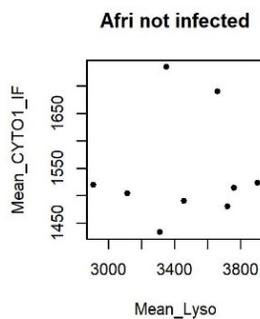
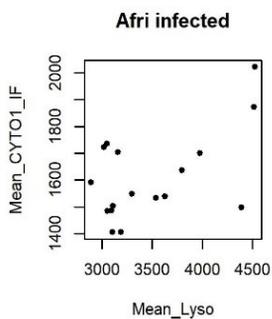
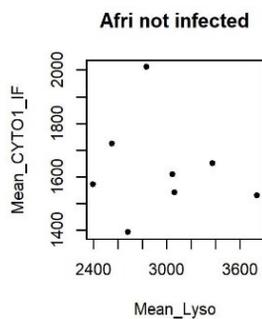
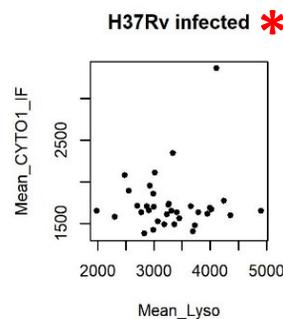
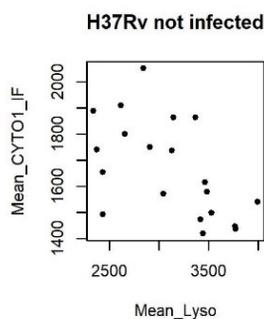
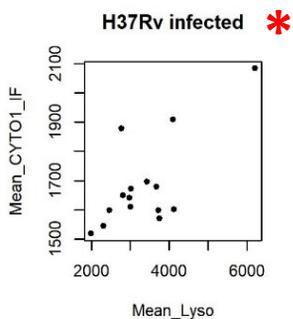
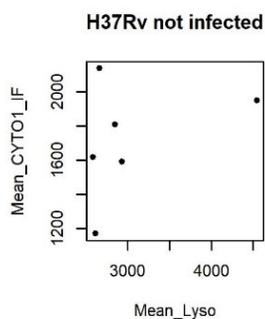
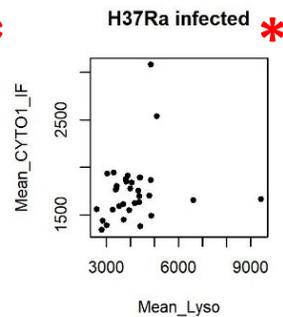
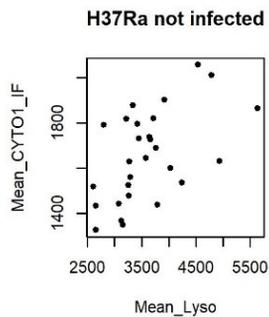
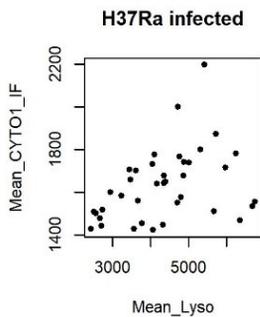
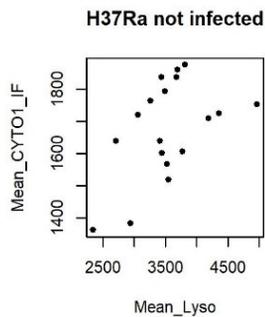
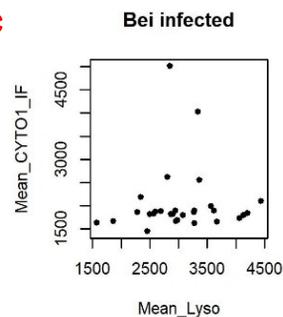
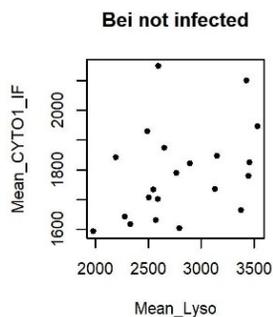
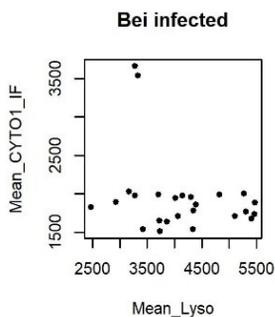
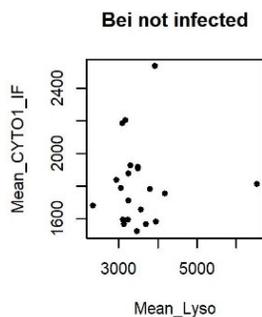
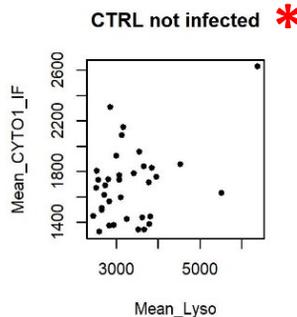
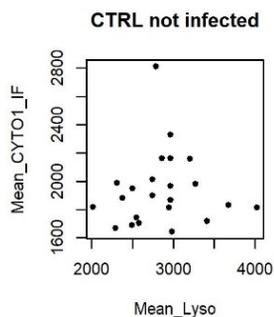
Par	Value	Std.Error	p-value
(Intercept)	-0.3527	0.2867	0.22
groupgeneralist not infected	0.1622	0.3621	0.6547
groupother not infected	0.6882	0.4296	0.1107
groupspecialist not infected	-0.3673	0.6503	0.5728
groupgeneralist infected	0.4523	0.3577	0.2075
groupother infected	0.7937	0.4138	0.0565
groupspecialist infected	0.1405	0.516	0.7856

Contrast	estimate	SE	p-value
ctrl not infected-generalist not infected	-0.1622	0.3621	0.7391
ctrl not infected-other not infected	-0.6882	0.4296	0.4424
ctrl not infected-specialist not infected	0.3673	0.6503	0.716
ctrl not infected-generalist infected	-0.4523	0.3577	0.4559
ctrl not infected-other infected	-0.7937	0.4138	0.4424
ctrl not infected-specialist infected	-0.1405	0.516	0.7856
generalist not infected-other not infected	-0.5261	0.389	0.4559
generalist not infected-specialist not infected	0.5295	0.6242	0.5959
other not infected-specialist not infected	1.0555	0.6656	0.4424
generalist infected-other infected	-0.3414	0.3671	0.589
generalist infected-specialist infected	0.3117	0.4794	0.7039
other infected-specialist infected	0.6532	0.5226	0.4559
generalist not infected-generalist infected	-0.2901	0.1848	0.4424
other not infected-other infected	-0.1055	0.2639	0.7391
specialist not infected-specialist infected	-0.5078	0.5309	0.589

Correlation between phagolysosomal acidification (LysoSensor) and IFN γ (immunofluorescence)

M1 – 24 h p.i.

M2 – 24 h p.i.

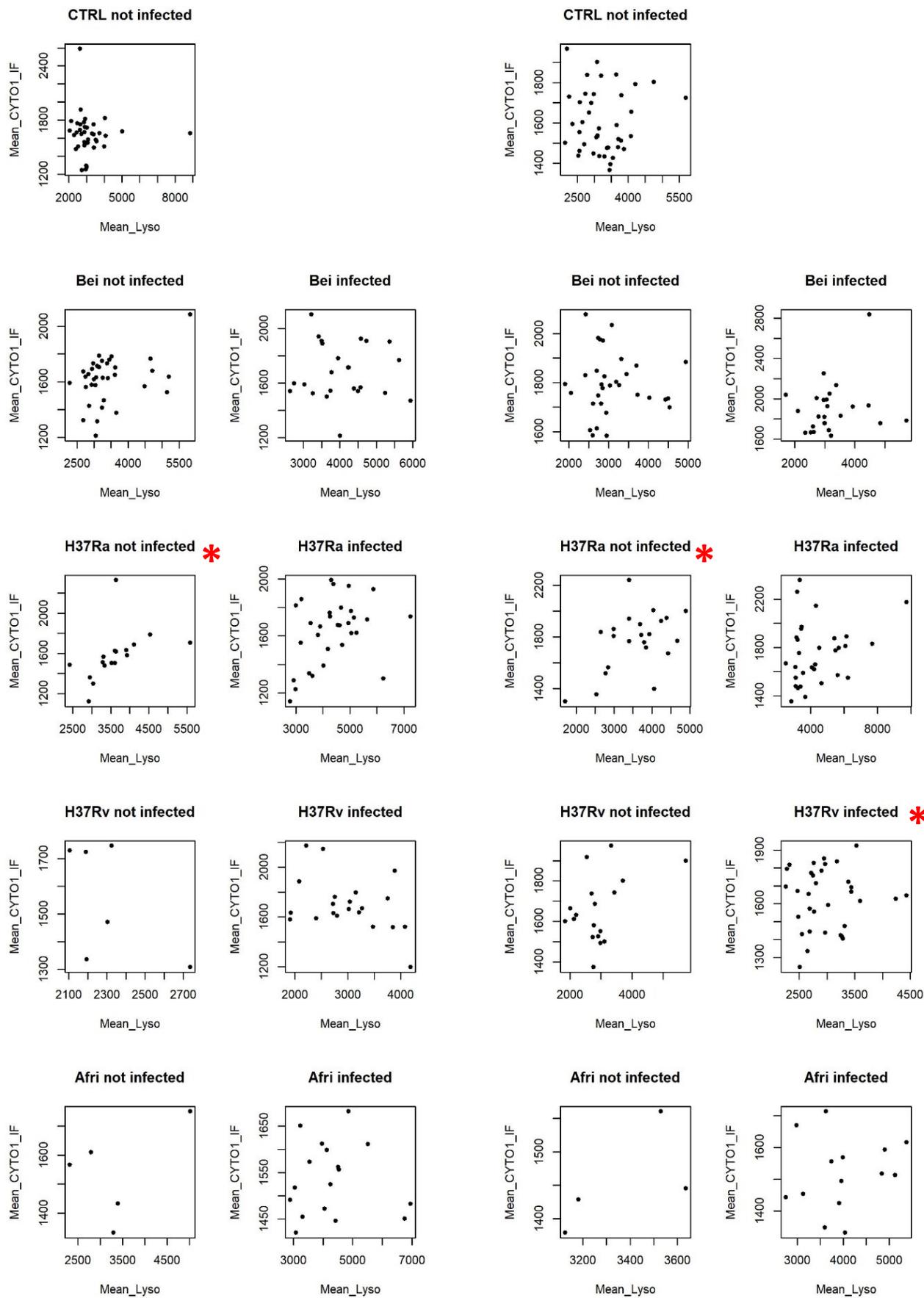


* Statistically significant correlation

Correlation between phagolysosomal acidification (LysoSensor) and TNF α (immunofluorescence)

M1 – 24 h p.i.

M2 – 24 h p.i.

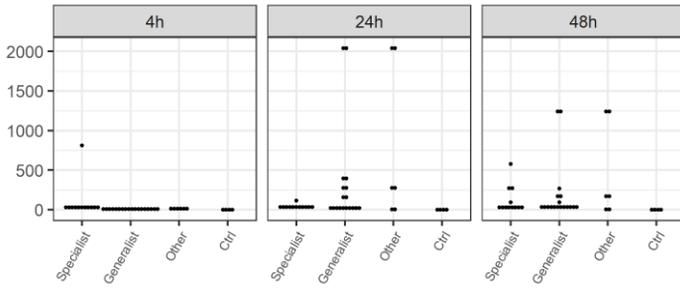


* Statistically significant correlation

Grouping: Geo-host adaptation

M1 – MIP-1β (CCL4)

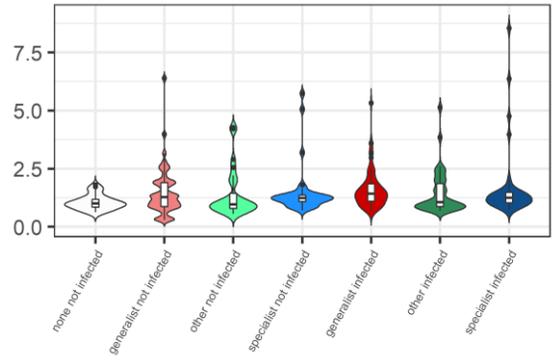
a) BioPlex (bulk)



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.8512	0.4674	0.0745
Time24h	0.5214	0.2749	0.0636
Time48h	0.6709	0.2749	0.0182
Geo_host_adaptationGeneralist	0.1332	0.251	0.5978
Time24h:Geo_host_adaptationGeneralist	0.5717	0.3549	0.1134
Time48h:Geo_host_adaptationGeneralist	0.0954	0.3549	0.7893

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.1332	0.251	0.5978
Specialist - Generalist	24h	-0.7049	0.251	0.007
Specialist - Generalist	48h	-0.2286	0.251	0.3667

b) Immunofluorescence (single-cell) – 24 h p.i.

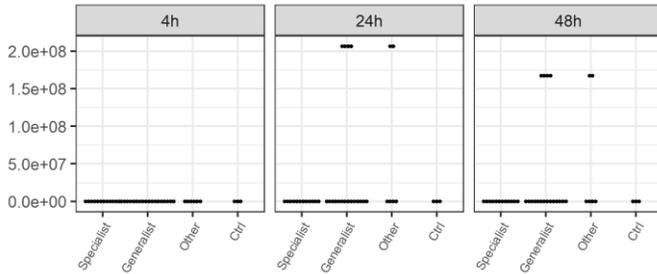


Par	Value	Std.Error	p-value
(Intercept)	0.0222	0.3033	0.9417
groupgeneralist not infected	0.4717	0.1851	0.0113
groupother not infected	0.3029	0.2097	0.1496
groupspecialist not infected	-0.0756	0.222	0.7338
groupgeneralist infected	0.5263	0.1832	0.0044
groupother infected	0.2395	0.207	0.2483
groupspecialist infected	-0.1113	0.2207	0.6145

Contrast	estimate	SE	p-value
none not infected-generalist not infected	-0.4717	0.1851	0.0448
none not infected-other not infected	-0.3029	0.2097	0.2805
none not infected-specialist not infected	0.0756	0.222	0.7338
none not infected-generalist infected	-0.5263	0.1832	0.0329
none not infected-other infected	-0.2395	0.207	0.4138
none not infected-specialist infected	0.1113	0.2207	0.6687
generalist not infected-other not infected	0.1688	0.189	0.4874
generalist not infected-specialist not infected	0.5473	0.2163	0.0448
other not infected-specialist not infected	0.3785	0.2258	0.2599
generalist infected-other infected	0.2868	0.1845	0.2599
generalist infected-specialist infected	0.6376	0.2136	0.0329
other infected-specialist infected	0.3508	0.2216	0.2599
generalist not infected-generalist infected	-0.0546	0.0557	0.4874
other not infected-other infected	0.0634	0.0736	0.4874
specialist not infected-specialist infected	0.0357	0.0727	0.6687

M2 – MIP-1β (CCL4)

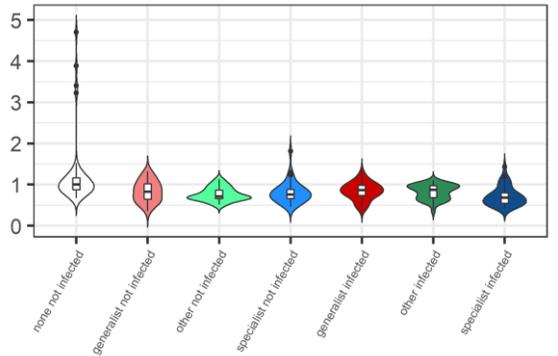
a) BioPlex (bulk)



Parameter	Estimate	Std.Error	p-value
(Intercept)	-1.0577	0.3966	0.0102
Time24h	1.2369	0.3517	9e-04
Time48h	0.83	0.3517	0.0221
Geo_host_adaptationGeneralist	0.5531	0.3211	0.091
Time24h:Geo_host_adaptationGeneralist	-0.3937	0.4541	0.3899
Time48h:Geo_host_adaptationGeneralist	-0.1473	0.4541	0.747

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.5531	0.3211	0.091
Specialist - Generalist	24h	-0.1594	0.3211	0.6217
Specialist - Generalist	48h	-0.4058	0.3211	0.212

b) Immunofluorescence (single-cell) – 24 h p.i.



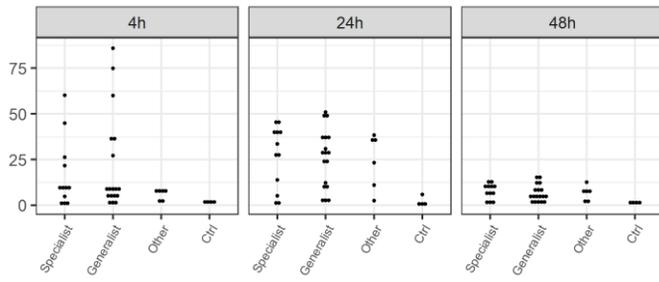
Par	Value	Std.Error	p-value
(Intercept)	-0.0087	0.1779	0.9612
groupgeneralist not infected	-0.2449	0.143	0.0877
groupother not infected	-0.2794	0.1628	0.0871
groupspecialist not infected	-0.105	0.1715	0.541
groupgeneralist infected	-0.2823	0.1428	0.0487
groupother infected	-0.2026	0.1619	0.2117
groupspecialist infected	-0.2201	0.1716	0.2006

Contrast	estimate	SE	p-value
none not infected-generalist not infected	0.2449	0.143	0.3133
none not infected-other not infected	0.2794	0.1628	0.3133
none not infected-specialist not infected	0.105	0.1715	0.7216
none not infected-generalist infected	0.2823	0.1426	0.3133
none not infected-other infected	0.2026	0.1619	0.4536
none not infected-specialist infected	0.2201	0.1716	0.4536
generalist not infected-other not infected	0.0344	0.1442	0.8695
generalist not infected-specialist not infected	-0.14	0.1631	0.5873
other not infected-specialist not infected	-0.1744	0.1719	0.5189
generalist infected-other infected	-0.0797	0.1428	0.7216
generalist infected-specialist infected	-0.0622	0.163	0.8111
other infected-specialist infected	0.0174	0.1712	0.9189
generalist not infected-generalist infected	0.0373	0.0333	0.4844
other not infected-other infected	-0.0767	0.0471	0.3133
specialist not infected-specialist infected	0.1151	0.0444	0.1494

Grouping: Geo-host adaptation

M1 – MCP-1

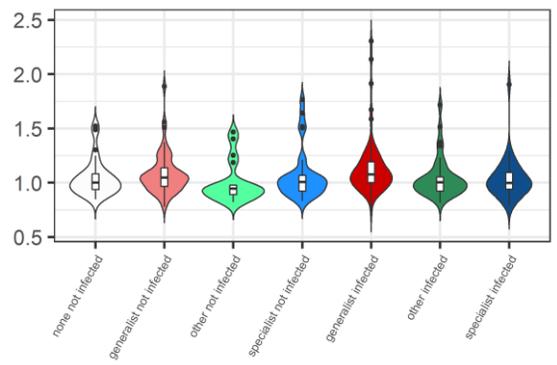
a) BioPlex (bulk)



Parameter	Estimate	Std.Error	p-value
(Intercept)	1.3888	0.6863	0.0483
Time24h	0.9816	0.371	0.0108
Time48h	-0.06	0.371	0.8721
Geo_host_adaptationGeneralist	0.5813	0.3387	0.0922
Time24h:Geo_host_adaptationGeneralist	-0.3471	0.4789	0.4719
Time48h:Geo_host_adaptationGeneralist	-0.5931	0.4789	0.2213

contrast	Time	ratio	SE	p-value
Specialist / Generalist	4h	0.5592	0.1894	0.0922
Specialist / Generalist	24h	0.7912	0.268	0.4924
Specialist / Generalist	48h	1.0119	0.3427	0.9724

b) Immunofluorescence (single-cell) – 24 h p.i.

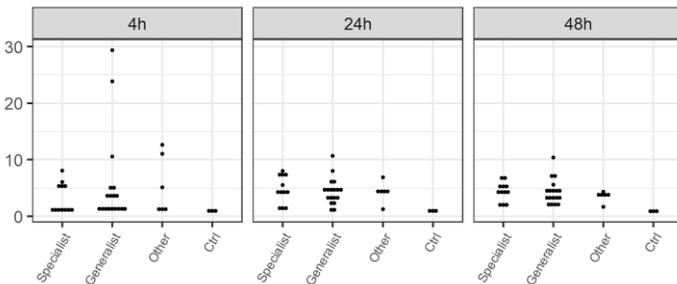


Par	Value	Std.Error	p-value
(Intercept)	0.003	0.047	0.9498
groupgeneralist not infected	0.0471	0.0579	0.4169
groupprother not infected	-0.0679	0.0691	0.3266
groupspecialist not infected	-0.0236	0.0674	0.726
groupgeneralist infected	0.0709	0.0575	0.2184
groupprother infected	0.0038	0.0665	0.9541
groupspecialist infected	0.0037	0.0664	0.955

Contrast	estimate	SE	p-value
none not infected-generalist not infected	-0.0471	0.0579	0.6254
none not infected-other not infected	0.0679	0.0691	0.5443
none not infected-specialist not infected	0.0236	0.0674	0.9075
none not infected-generalist infected	-0.0709	0.0575	0.5245
none not infected-other infected	-0.0038	0.0665	0.999
none not infected-specialist infected	-0.0037	0.0664	0.999
generalist not infected-other not infected	0.115	0.0609	0.4497
generalist not infected-specialist not infected	0.0707	0.0589	0.5245
other not infected-specialist not infected	-0.0442	0.0699	0.7194
generalist infected-other infected	0.0671	0.0576	0.5245
generalist infected-specialist infected	0.0672	0.0573	0.5245
other infected-specialist infected	1e-04	0.0664	0.999
generalist not infected-generalist infected	-0.0239	0.0185	0.5245
other not infected-other infected	-0.0717	0.0314	0.3465
specialist not infected-specialist infected	-0.0274	0.0271	0.5443

M2 – MCP-1

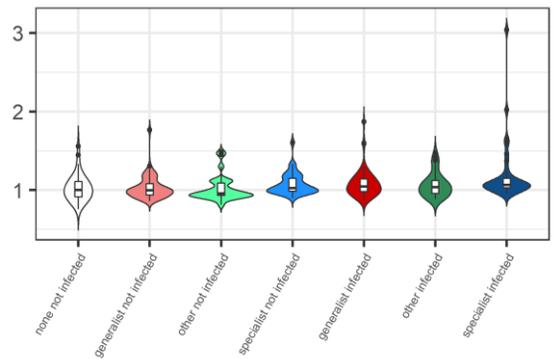
a) BioPlex (bulk)



Parameter	Estimate	Std.Error	p-value
(Intercept)	0.7511	0.3205	0.023
Time24h	0.4851	0.3106	0.1246
Time48h	0.5348	0.3106	0.0912
Geo_host_adaptationGeneralist	0.2181	0.2836	0.4454
Time24h:Geo_host_adaptationGeneralist	-0.2826	0.401	0.4842
Time48h:Geo_host_adaptationGeneralist	-0.2724	0.401	0.5

contrast	Time	ratio	SE	p-value
Specialist / Generalist	4h	0.8041	0.228	0.4454
Specialist / Generalist	24h	1.0667	0.3025	0.8209
Specialist / Generalist	48h	1.0559	0.2994	0.8487

b) Immunofluorescence (single-cell) – 24 h p.i.



Par	Value	Std.Error	p-value
(Intercept)	0.0013	0.0304	0.9664
groupgeneralist not infected	0.0228	0.0249	0.3589
groupprother not infected	-0.0248	0.0299	0.4072
groupspecialist not infected	0.0361	0.0295	0.2223
groupgeneralist infected	0.0597	0.0245	0.0156
groupprother infected	0.0603	0.0276	0.0298
groupspecialist infected	0.054	0.0305	0.0779

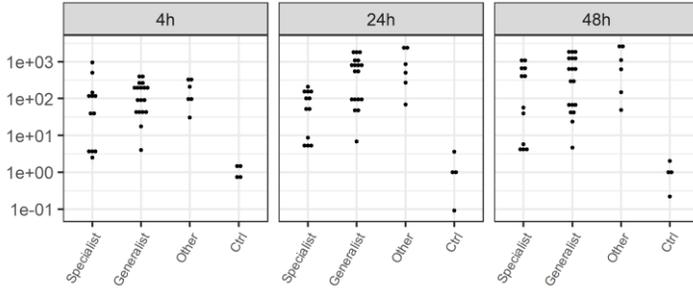
Contrast	estimate	SE	p-value
none not infected-generalist not infected	-0.0228	0.0249	0.5981
none not infected-other not infected	0.0248	0.0299	0.6107
none not infected-specialist not infected	-0.0361	0.0295	0.4168
none not infected-generalist infected	-0.0597	0.0245	0.1173
none not infected-other infected	-0.0603	0.0276	0.1488
none not infected-specialist infected	-0.054	0.0305	0.1845
generalist not infected-other not infected	0.0477	0.0277	0.1845
generalist not infected-specialist not infected	-0.0132	0.0279	0.7942
other not infected-specialist not infected	-0.0609	0.031	0.1599
generalist infected-other infected	-7e-04	0.0248	0.979
generalist infected-specialist infected	0.0057	0.0293	0.9073
other infected-specialist infected	0.0063	0.0303	0.9073
generalist not infected-generalist infected	-0.0369	0.019	0.1599
other not infected-other infected	-0.0852	0.0269	0.0262
specialist not infected-specialist infected	-0.018	0.026	0.6679

Cytokines

IL-1 β (logarithmic transformation)

M1

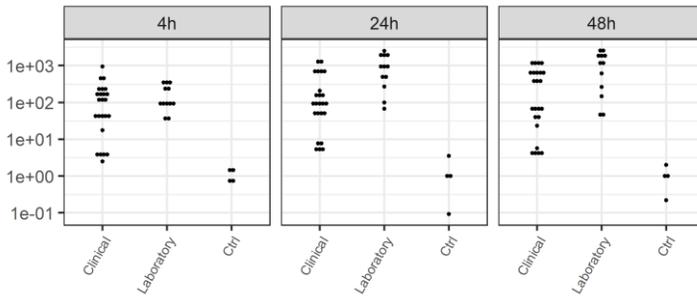
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	3.1332	0.898	0.001
Time24h	0.126	0.5026	0.803
Time48h	0.6053	0.5026	0.234
Geo_host_adaptationGeneralist	1.0797	0.4588	0.0225
Time24h:Geo_host_adaptationGeneralist	1.0129	0.6488	0.1247
Time48h:Geo_host_adaptationGeneralist	0.2476	0.6488	0.7043

contrast	Time	ratio	SE	p-value
Specialist / Generalist	4h	0.3397	0.1558	0.0225
Specialist / Generalist	24h	0.1234	0.0566	<0.0001
Specialist / Generalist	48h	0.2652	0.1217	0.0056

Lab_adaptation

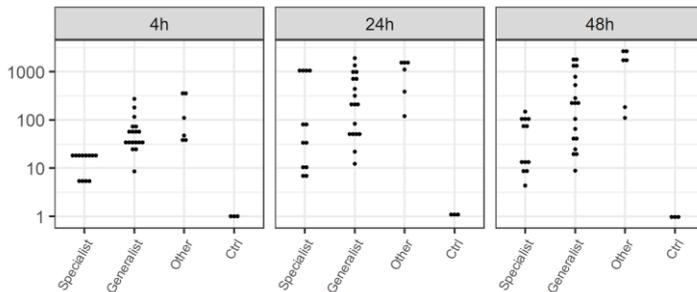


Parameter	Estimate	Std.Error	p-value
(Intercept)	3.5932	0.8327	1e-04
Time24h	0.4926	0.3461	0.1595
Time48h	0.5607	0.3461	0.1102
Lab_adaptationLaboratory	1.0004	0.4238	0.0214
Time24h:Lab_adaptationLaboratory	1.0395	0.5994	0.0877
Time48h:Lab_adaptationLaboratory	0.8204	0.5994	0.176

contrast	Time	ratio	SE	p-value
Clinical / Laboratory	4h	0.3677	0.1559	0.0214
Clinical / Laboratory	24h	0.13	0.0551	<0.0001
Clinical / Laboratory	48h	0.1619	0.0686	1e-04

M2

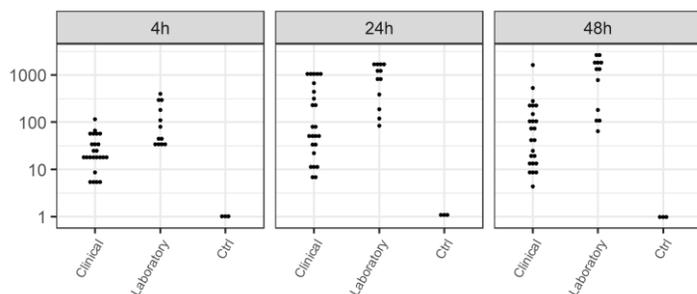
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	2.2805	0.6836	0.0016
Time24h	1.5483	0.4843	0.0024
Time48h	0.8123	0.4843	0.0996
Geo_host_adaptationGeneralist	1.4036	0.4421	0.0025
Time24h:Geo_host_adaptationGeneralist	-0.2928	0.6252	0.6415
Time48h:Geo_host_adaptationGeneralist	0.1804	0.6252	0.7741

contrast	Time	ratio	SE	p-value
Specialist / Generalist	4h	0.2457	0.1086	0.0025
Specialist / Generalist	24h	0.3393	0.1456	0.0152
Specialist / Generalist	48h	0.2051	0.0907	8e-04

Lab_adaptation



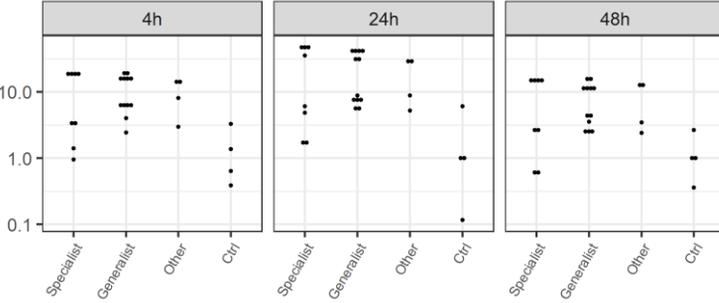
Parameter	Estimate	Std.Error	p-value
(Intercept)	2.8781	0.6266	<0.0001
Time24h	1.2575	0.3153	2e-04
Time48h	0.7271	0.3153	0.0244
Lab_adaptationLaboratory	1.347	0.3862	9e-04
Time24h:Lab_adaptationLaboratory	0.6441	0.5462	0.2427
Time48h:Lab_adaptationLaboratory	1.111	0.5462	0.0462

contrast	Time	ratio	SE	p-value
Clinical / Laboratory	4h	0.26	0.1004	9e-04
Clinical / Laboratory	24h	0.1365	0.0527	<0.0001
Clinical / Laboratory	48h	0.0856	0.0331	<0.0001

IL-1ra (logarithmic transformation)

M1

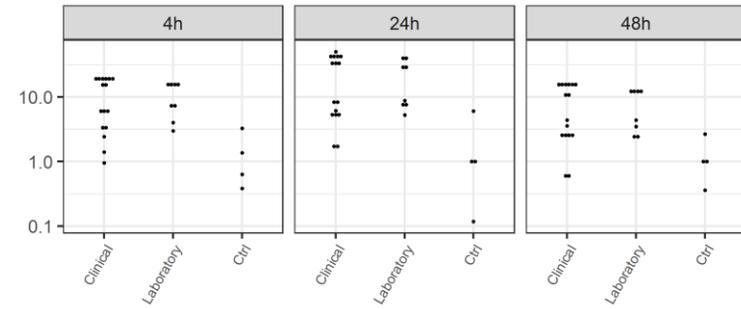
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	1.4289	0.6462	0.0333
Time24h	0.5749	0.2723	0.0416
Time48h	-0.3663	0.2723	0.1868
Geo_host_adaptationGeneralist	0.5735	0.2486	0.0268
Time24h:Geo_host_adaptationGeneralist	-0.068	0.3516	0.8476
Time48h:Geo_host_adaptationGeneralist	-0.0159	0.3516	0.9641

contrast	Time	ratio	SE	p-value
Specialist / Generalist	4h	0.5636	0.1401	0.0268
Specialist / Generalist	24h	0.6032	0.15	0.0493
Specialist / Generalist	48h	0.5726	0.1424	0.031

Lab_adaptation

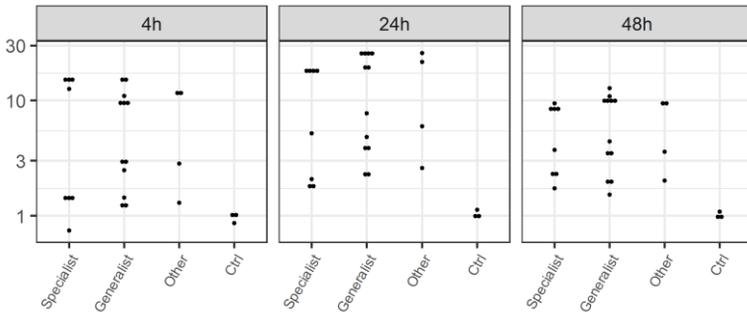


Parameter	Estimate	Std.Error	p-value
(Intercept)	1.7107	0.6045	0.0069
Time24h	0.529	0.2101	0.0154
Time48h	-0.3688	0.2101	0.0859
Lab_adaptationLaboratory	0.2718	0.2574	0.2964
Time24h:Lab_adaptationLaboratory	-0.0253	0.364	0.9449
Time48h:Lab_adaptationLaboratory	-0.0273	0.364	0.9405

contrast	Time	ratio	SE	p-value
Clinical / Laboratory	4h	0.762	0.1961	0.2964
Clinical / Laboratory	24h	0.7815	0.2011	0.3431
Clinical / Laboratory	48h	0.7831	0.2015	0.347

M2

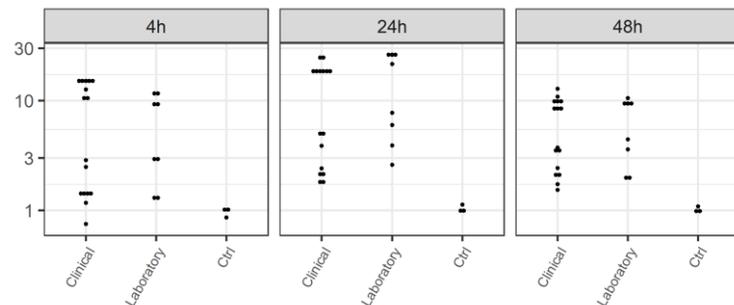
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.8433	0.563	0.1427
Time24h	0.747	0.2766	0.0104
Time48h	0.46	0.2766	0.1047
Geo_host_adaptationGeneralist	0.3013	0.2525	0.2404
Time24h:Geo_host_adaptationGeneralist	0.1747	0.3571	0.6276
Time48h:Geo_host_adaptationGeneralist	-0.1963	0.3571	0.5859

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.3013	0.2525	0.2404
Specialist - Generalist	24h	-0.476	0.2525	0.0673
Specialist - Generalist	48h	-0.105	0.2525	0.6799

Lab_adaptation



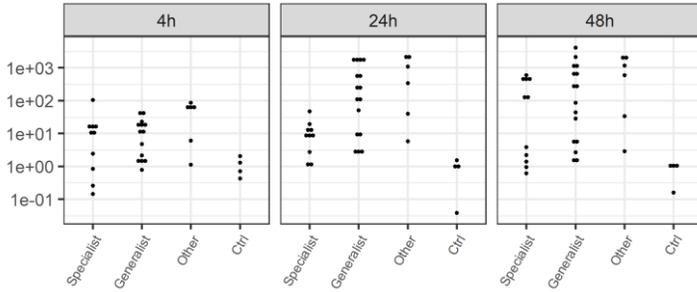
Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.6735	0.5481	0.2254
Time24h	0.7071	0.1945	7e-04
Time48h	0.312	0.1945	0.1154
Lab_adaptationLaboratory	0.0765	0.2382	0.7494
Time24h:Lab_adaptationLaboratory	0.4138	0.3368	0.2254
Time48h:Lab_adaptationLaboratory	-0.0204	0.3368	0.952

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.0765	0.2382	0.7494
Clinical - Laboratory	24h	-0.4904	0.2382	0.0452
Clinical - Laboratory	48h	-0.0562	0.2382	0.8146

IL-6 (logarithmic transformation)

M1

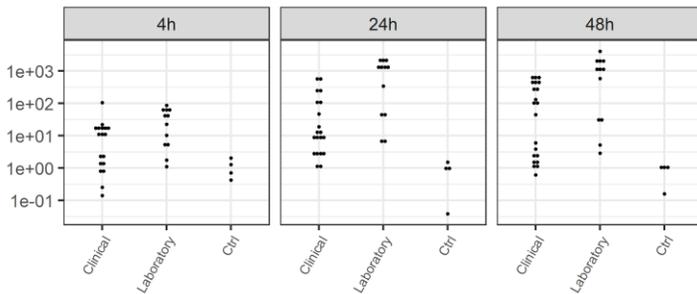
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.4217	0.3601	0.247
Time24h	0.1159	0.3681	0.7541
Time48h	0.5162	0.3681	0.1669
Geo_host_adaptationGeneralist	0.0083	0.336	0.9805
Time24h:Geo_host_adaptationGeneralist	0.6241	0.4752	0.1949
Time48h:Geo_host_adaptationGeneralist	0.174	0.4752	0.7158

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.0083	0.336	0.9805
Specialist - Generalist	24h	-0.6324	0.336	0.0655
Specialist - Generalist	48h	-0.1823	0.336	0.5899

Lab_adaptation

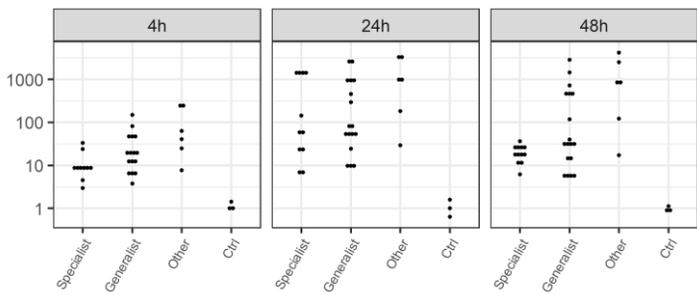


Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.7019	0.2885	0.0178
Time24h	0.4316	0.231	0.0664
Time48h	0.6115	0.231	0.0102
Lab_adaptationLaboratory	0.7652	0.2829	0.0088
Time24h:Lab_adaptationLaboratory	0.2533	0.4001	0.529
Time48h:Lab_adaptationLaboratory	0.0454	0.4001	0.9101

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.7652	0.2829	0.0088
Clinical - Laboratory	24h	-1.0185	0.2829	6e-04
Clinical - Laboratory	48h	-0.8106	0.2829	0.0057

M2

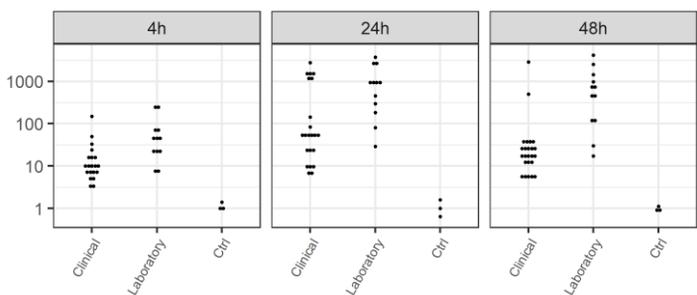
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.95	0.3338	0.0064
Time24h	1.2182	0.3444	9e-04
Time48h	0.6859	0.3444	0.0518
Geo_host_adaptationGeneralist	0.467	0.3144	0.1436
Time24h:Geo_host_adaptationGeneralist	-0.311	0.4446	0.4874
Time48h:Geo_host_adaptationGeneralist	-0.0452	0.4446	0.9194

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.467	0.3144	0.1436
Specialist - Generalist	24h	-0.156	0.3144	0.6219
Specialist - Generalist	48h	-0.4218	0.3144	0.1856

Lab_adaptation



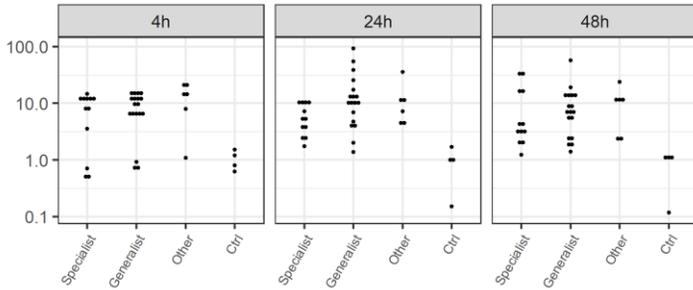
Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.9666	0.277	9e-04
Time24h	0.9737	0.2007	<0.0001
Time48h	0.5709	0.2007	0.006
Lab_adaptationLaboratory	0.9	0.2459	5e-04
Time24h:Lab_adaptationLaboratory	0.0265	0.3477	0.9394
Time48h:Lab_adaptationLaboratory	0.2729	0.3477	0.4355

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.9	0.2459	5e-04
Clinical - Laboratory	24h	-0.9265	0.2459	4e-04
Clinical - Laboratory	48h	-1.1729	0.2459	<0.0001

IL-8 (logarithmic transformation)

M1

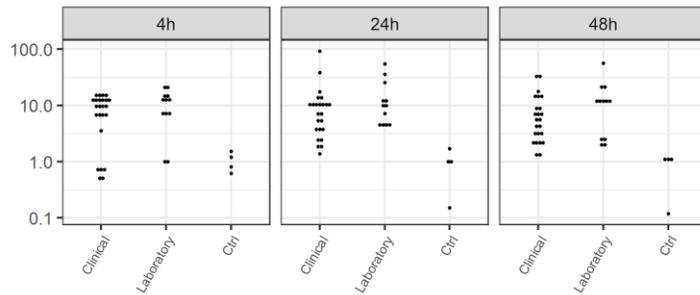
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	1.1811	0.4599	0.0132
Time24h	0.3432	0.5055	0.5003
Time48h	0.354	0.5055	0.487
Geo_host_adaptationGeneralist	0.4805	0.4615	0.3027
Time24h:Geo_host_adaptationGeneralist	0.3484	0.6526	0.5958
Time48h:Geo_host_adaptationGeneralist	-0.0511	0.6526	0.9378

contrast	Time	ratio	SE	p-value
Specialist / Generalist	4h	0.6185	0.2854	0.3027
Specialist / Generalist	24h	0.4365	0.2015	0.0784
Specialist / Generalist	48h	0.6509	0.3004	0.3565

Lab_adaptation

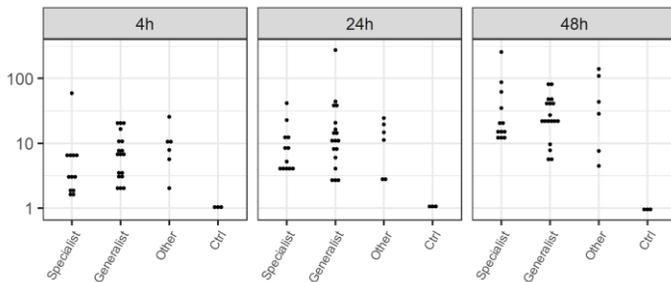


Parameter	Estimate	Std.Error	p-value
(Intercept)	1.3995	0.3552	2e-04
Time24h	0.4703	0.3583	0.1941
Time48h	0.207	0.3583	0.5655
Lab_adaptationLaboratory	0.4661	0.4388	0.2922
Time24h:Lab_adaptationLaboratory	0.1835	0.6206	0.7684
Time48h:Lab_adaptationLaboratory	0.299	0.6206	0.6316

contrast	Time	ratio	SE	p-value
Clinical / Laboratory	4h	0.6274	0.2753	0.2922
Clinical / Laboratory	24h	0.5222	0.2292	0.1437
Clinical / Laboratory	48h	0.4653	0.2042	0.0861

M2

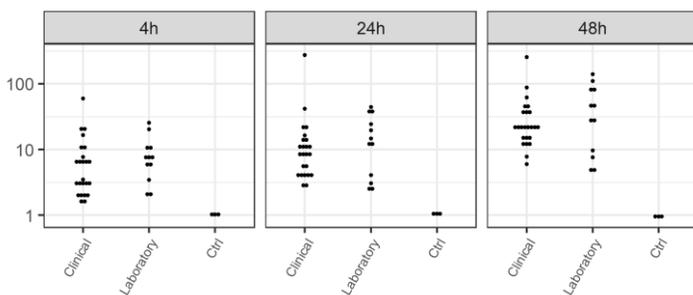
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.8703	0.3132	0.0076
Time24h	0.6888	0.3625	0.0631
Time48h	1.7044	0.3625	<0.0001
Geo_host_adaptationGeneralist	0.4926	0.3309	0.1427
Time24h:Geo_host_adaptationGeneralist	-0.1275	0.4679	0.7863
Time48h:Geo_host_adaptationGeneralist	-0.4968	0.4679	0.2934

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.4926	0.3309	0.1427
Specialist - Generalist	24h	-0.3651	0.3309	0.2751
Specialist - Generalist	48h	0.0042	0.3309	0.9899

Lab_adaptation



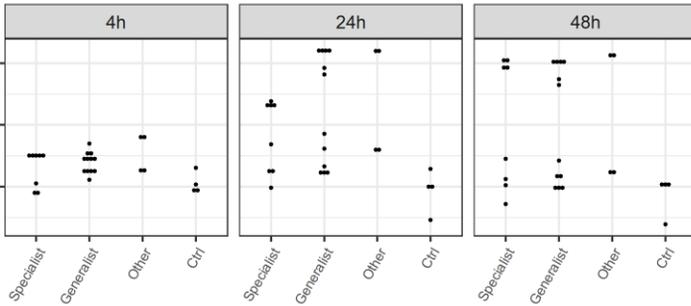
Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.6421	0.2705	0.0207
Time24h	0.583	0.2577	0.0271
Time48h	1.4078	0.2577	<0.0001
Lab_adaptationLaboratory	0.3615	0.3156	0.2565
Time24h:Lab_adaptationLaboratory	-0.1414	0.4464	0.7525
Time48h:Lab_adaptationLaboratory	-0.1382	0.4464	0.7578

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.3615	0.3156	0.2565
Clinical - Laboratory	24h	-0.2201	0.3156	0.4882
Clinical - Laboratory	48h	-0.2232	0.3156	0.482

IL-10 (logarithmic transformation)

M1

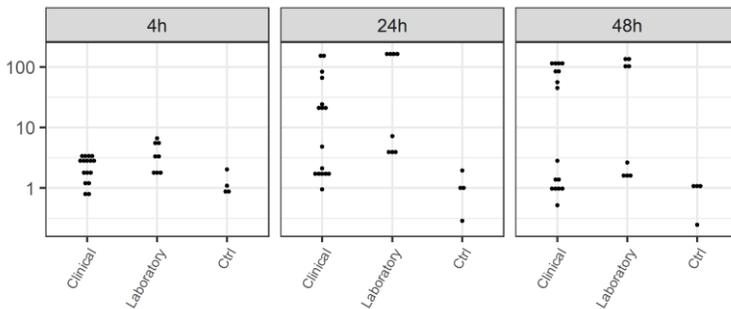
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.7685	0.5475	0.1687
Time24h	0.5941	0.3544	0.1021
Time48h	0.3549	0.3544	0.3231
Geo_host_adaptationGeneralist	0.4255	0.3235	0.1965
Time24h:Geo_host_adaptationGeneralist	0.104	0.4576	0.8214
Time48h:Geo_host_adaptationGeneralist	-0.3401	0.4576	0.4619

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.4255	0.3235	0.1965
Specialist - Generalist	24h	-0.5296	0.3235	0.1102
Specialist - Generalist	48h	-0.0854	0.3235	0.7933

Lab_adaptation

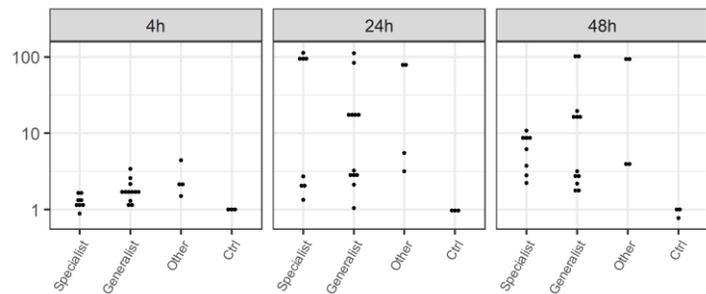


Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.6924	0.51	0.1811
Time24h	0.5825	0.2147	0.0093
Time48h	0.1375	0.2147	0.5252
Lab_adaptationLaboratory	0.465	0.263	0.0837
Time24h:Lab_adaptationLaboratory	0.4544	0.3719	0.228
Time48h:Lab_adaptationLaboratory	0.0271	0.3719	0.9423

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.465	0.263	0.0837
Clinical - Laboratory	24h	-0.9194	0.263	0.0011
Clinical - Laboratory	48h	-0.492	0.263	0.0677

M2

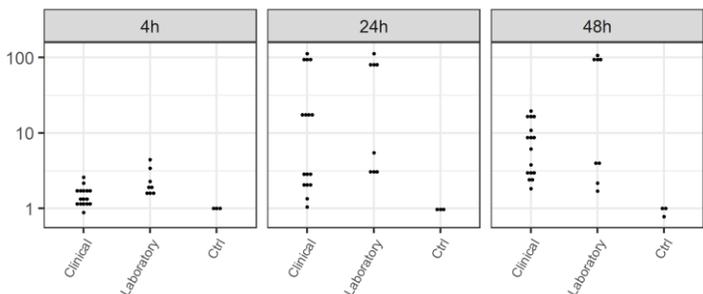
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-1.3338	0.3722	0.001
Time24h	1.6934	0.3488	<0.0001
Time48h	1.5947	0.3488	1e-04
Geo_host_adaptationGeneralist	0.6186	0.3184	0.0597
Time24h:Geo_host_adaptationGeneralist	-0.7651	0.4503	0.0977
Time48h:Geo_host_adaptationGeneralist	-0.5838	0.4503	0.2028

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.6186	0.3184	0.0597
Specialist - Generalist	24h	0.1465	0.3184	0.6481
Specialist - Generalist	48h	-0.0348	0.3184	0.9136

Lab_adaptation



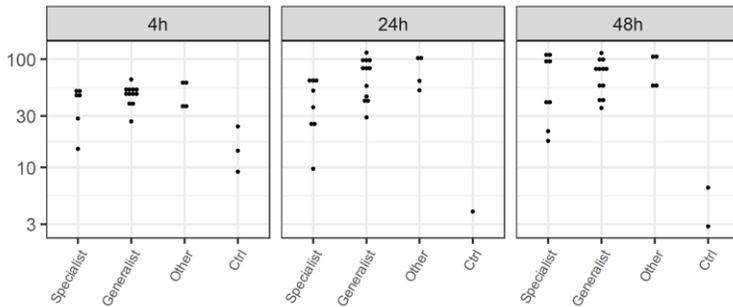
Parameter	Estimate	Std.Error	p-value
(Intercept)	-1.1752	0.3293	9e-04
Time24h	1.2767	0.2369	<0.0001
Time48h	1.3542	0.2369	<0.0001
Lab_adaptationLaboratory	0.653	0.2902	0.0293
Time24h:Lab_adaptationLaboratory	-0.1926	0.4104	0.6411
Time48h:Lab_adaptationLaboratory	-0.3247	0.4104	0.4329

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.653	0.2902	0.0293
Clinical - Laboratory	24h	-0.4604	0.2902	0.1195
Clinical - Laboratory	48h	-0.3283	0.2902	0.2638

IL-15 (logarithmic transformation)

M1

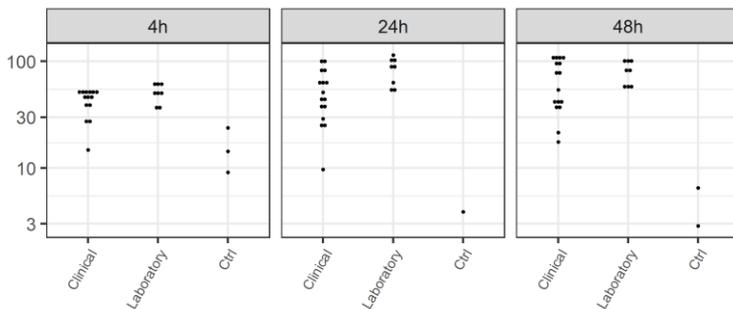
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-1.1803	0.5086	0.0259
Time24h	0.4752	0.2854	0.1043
Time48h	1.0331	0.2854	9e-04
Geo_host_adaptationGeneralist	0.8558	0.2605	0.0022
Time24h:Geo_host_adaptationGeneralist	0.1315	0.3684	0.7232
Time48h:Geo_host_adaptationGeneralist	-0.4369	0.3684	0.2432

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.8558	0.2605	0.0022
Specialist - Generalist	24h	-0.9873	0.2605	5e-04
Specialist - Generalist	48h	-0.4189	0.2605	0.1163

Lab_adaptation

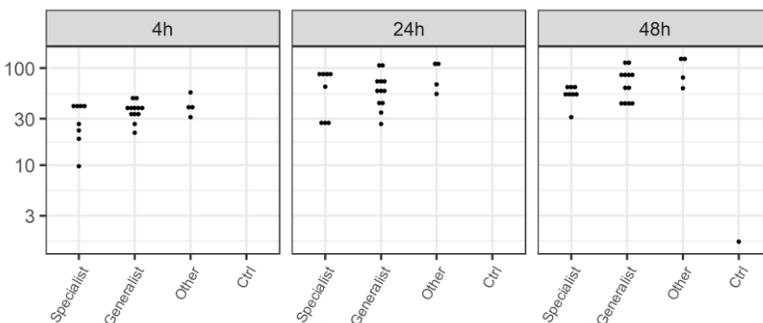


Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.9407	0.4765	0.0544
Time24h	0.4927	0.1892	0.0123
Time48h	0.8665	0.1892	<0.0001
Lab_adaptationLaboratory	0.6391	0.2317	0.0083
Time24h:Lab_adaptationLaboratory	0.4892	0.3276	0.1422
Time48h:Lab_adaptationLaboratory	-0.0112	0.3276	0.9728

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.6391	0.2317	0.0083
Clinical - Laboratory	24h	-1.1283	0.2317	<0.0001
Clinical - Laboratory	48h	-0.6278	0.2317	0.0094

M2

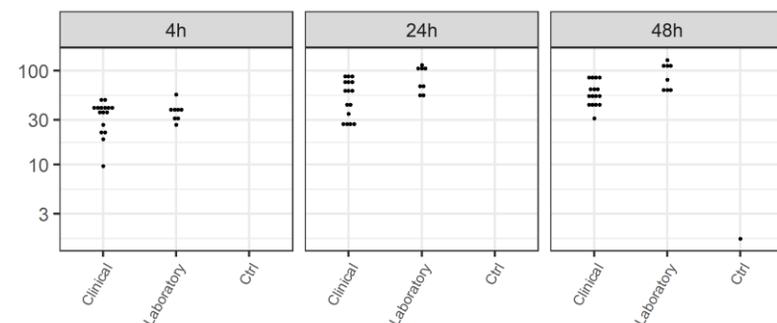
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	3.2263	0.2128	<0.0001
Time24h	0.6077	0.1759	0.0014
Time48h	0.706	0.1759	3e-04
Geo_host_adaptationGeneralist	0.3118	0.1606	0.0598
Time24h:Geo_host_adaptationGeneralist	-0.1739	0.2271	0.4486
Time48h:Geo_host_adaptationGeneralist	-0.1281	0.2271	0.5762

contrast	Time	ratio	SE	p-value
Specialist / Generalist	4h	0.7321	0.1175	0.0598
Specialist / Generalist	24h	0.8711	0.1399	0.3958
Specialist / Generalist	48h	0.8321	0.1336	0.2597

Lab_adaptation



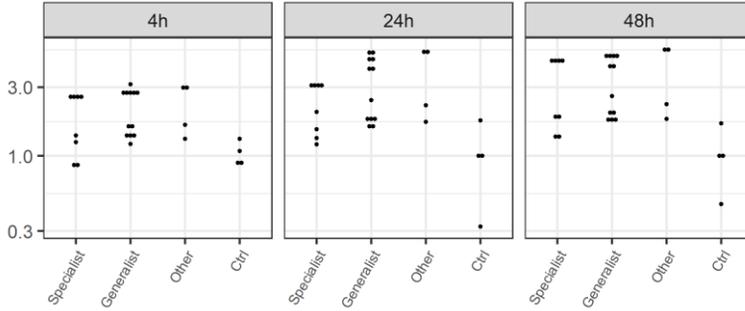
Parameter	Estimate	Std.Error	p-value
(Intercept)	3.3962	0.1865	<0.0001
Time24h	0.424	0.1092	3e-04
Time48h	0.5719	0.1092	<0.0001
Lab_adaptationLaboratory	0.1606	0.1338	0.2359
Time24h:Lab_adaptationLaboratory	0.3305	0.1892	0.0873
Time48h:Lab_adaptationLaboratory	0.2642	0.1892	0.1693

contrast	Time	ratio	SE	p-value
Clinical / Laboratory	4h	0.8516	0.1139	0.2359
Clinical / Laboratory	24h	0.6119	0.0818	6e-04
Clinical / Laboratory	48h	0.6539	0.0875	0.0027

FGF basic(logarithmic transformation)

M1

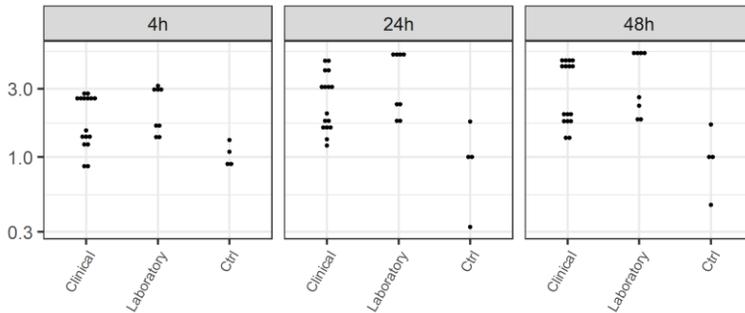
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	0.3589	0.2914	0.2259
Time24h	0.2868	0.0874	0.0023
Time48h	0.4621	0.0874	<0.0001
Geo_host_adaptationGeneralist	0.2224	0.0798	0.0084
Time24h:Geo_host_adaptationGeneralist	0.0455	0.1129	0.6893
Time48h:Geo_host_adaptationGeneralist	-0.0766	0.1129	0.5016

contrast	Time	ratio	SE	p-value
Specialist / Generalist	4h	0.8006	0.0639	0.0084
Specialist / Generalist	24h	0.765	0.0611	0.0018
Specialist / Generalist	48h	0.8643	0.069	0.0759

Lab_adaptation

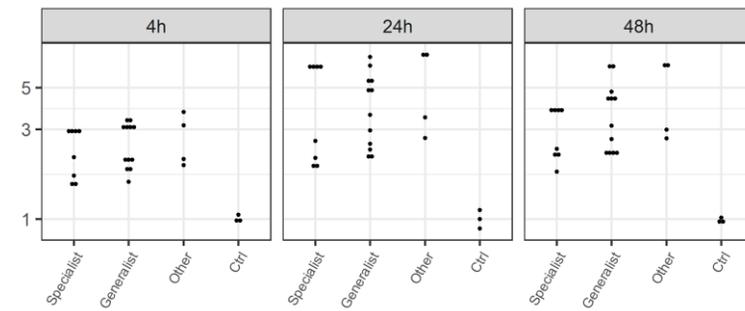


Parameter	Estimate	Std.Error	p-value
(Intercept)	0.4512	0.2905	0.1272
Time24h	0.2929	0.0617	<0.0001
Time48h	0.4168	0.0617	<0.0001
Lab_adaptationLaboratory	0.1875	0.0756	0.0168
Time24h:Lab_adaptationLaboratory	0.0984	0.1069	0.3618
Time48h:Lab_adaptationLaboratory	-4e-04	0.1069	0.9971

contrast	Time	ratio	SE	p-value
Clinical / Laboratory	4h	0.829	0.0627	0.0168
Clinical / Laboratory	24h	0.7513	0.0568	4e-04
Clinical / Laboratory	48h	0.8293	0.0627	0.017

M2

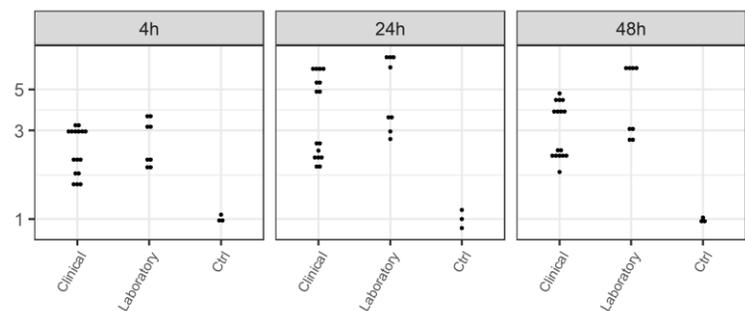
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	0.7161	0.2443	0.0058
Time24h	0.4077	0.098	2e-04
Time48h	0.2303	0.098	0.0242
Geo_host_adaptationGeneralist	0.0969	0.0894	0.2858
Time24h:Geo_host_adaptationGeneralist	-0.0125	0.1265	0.9219
Time48h:Geo_host_adaptationGeneralist	0.0875	0.1265	0.4931

contrast	Time	ratio	SE	p-value
Specialist / Generalist	4h	0.9077	0.0812	0.2858
Specialist / Generalist	24h	0.9191	0.0822	0.3516
Specialist / Generalist	48h	0.8316	0.0744	0.0463

Lab_adaptation



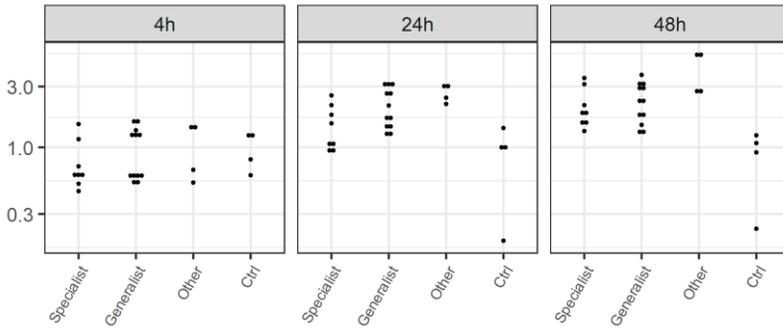
Parameter	Estimate	Std.Error	p-value
(Intercept)	0.7533	0.2404	0.003
Time24h	0.3584	0.054	<0.0001
Time48h	0.2344	0.054	1e-04
Lab_adaptationLaboratory	0.1082	0.0661	0.1084
Time24h:Lab_adaptationLaboratory	0.2054	0.0935	0.033
Time48h:Lab_adaptationLaboratory	0.2325	0.0935	0.0166

contrast	Time	ratio	SE	p-value
Clinical / Laboratory	4h	0.8974	0.0593	0.1084
Clinical / Laboratory	24h	0.7308	0.0483	<0.0001
Clinical / Laboratory	48h	0.7113	0.047	<0.0001

G-CSF (logarithmic transformation)

M1

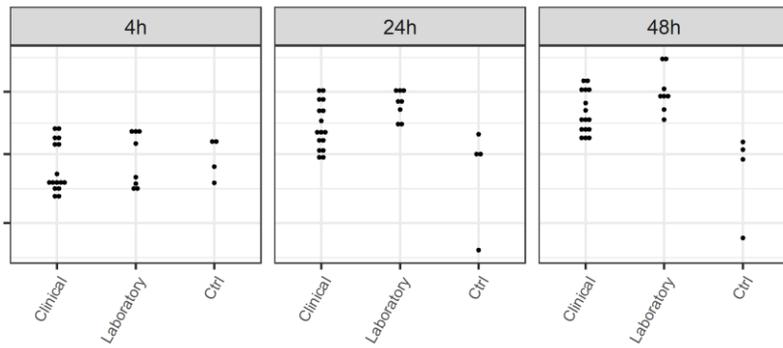
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.2671	0.2404	0.2738
Time24h	0.7224	0.0987	<0.0001
Time48h	1.0556	0.0987	<0.0001
Geo_host_adaptationGeneralist	0.2886	0.0901	0.0028
Time24h:Geo_host_adaptationGeneralist	0.064	0.1274	0.6181
Time48h:Geo_host_adaptationGeneralist	-0.1739	0.1274	0.1805

contrast	Time	ratio	SE	p-value
Specialist / Generalist	4h	0.7493	0.0675	0.0028
Specialist / Generalist	24h	0.7028	0.0633	4e-04
Specialist / Generalist	48h	0.8916	0.0803	0.2107

Lab_adaptation

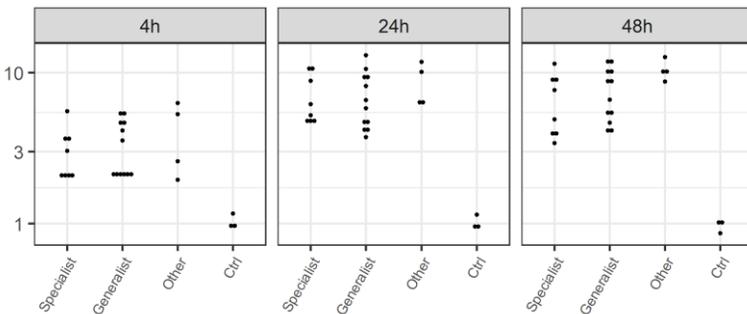


Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.1257	0.2112	0.5547
Time24h	0.736	0.1067	<0.0001
Time48h	0.9548	0.1067	<0.0001
Lab_adaptationLaboratory	0.1719	0.1307	0.195
Time24h:Lab_adaptationLaboratory	0.1599	0.1849	0.3915
Time48h:Lab_adaptationLaboratory	0.105	0.1849	0.5728

contrast	Time	ratio	SE	p-value
Clinical / Laboratory	4h	0.8421	0.1101	0.195
Clinical / Laboratory	24h	0.7176	0.0938	0.0146
Clinical / Laboratory	48h	0.7581	0.0991	0.0396

M2

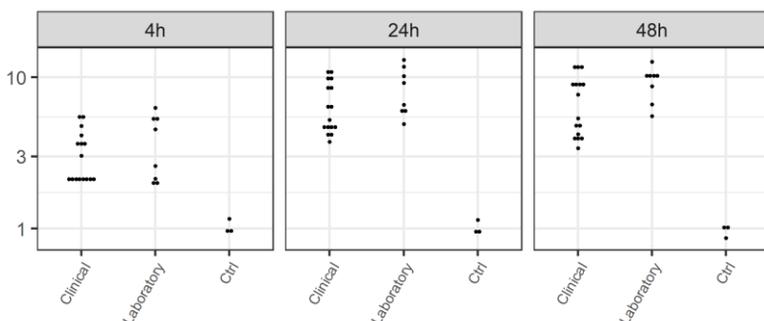
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.9208	0.4711	0.0582
Time24h	1.5376	0.1944	<0.0001
Time48h	1.362	0.1944	<0.0001
Geo_host_adaptationGeneralist	0.3551	0.1774	0.0527
Time24h:Geo_host_adaptationGeneralist	-0.3234	0.2509	0.2055
Time48h:Geo_host_adaptationGeneralist	0.017	0.2509	0.9462

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.3551	0.1774	0.0527
Specialist - Generalist	24h	-0.0317	0.1774	0.859
Specialist - Generalist	48h	-0.3721	0.1774	0.0428

Lab_adaptation



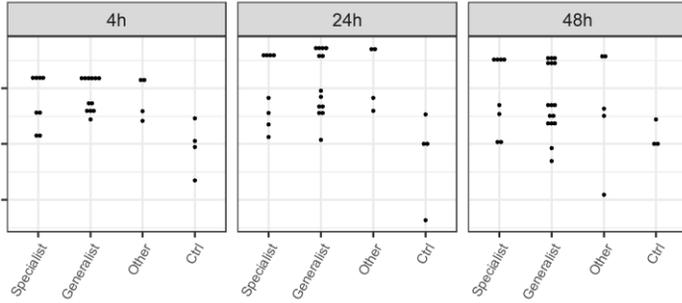
Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.7541	0.4436	0.0959
Time24h	1.1527	0.136	<0.0001
Time48h	1.2307	0.136	<0.0001
Lab_adaptationLaboratory	0.1201	0.1666	0.4747
Time24h:Lab_adaptationLaboratory	0.4738	0.2355	0.0501
Time48h:Lab_adaptationLaboratory	0.4992	0.2355	0.0395

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.1201	0.1666	0.4747
Clinical - Laboratory	24h	-0.5939	0.1666	9e-04
Clinical - Laboratory	48h	-0.6192	0.1666	5e-04

IFN γ (logarithmic transformation)

M1

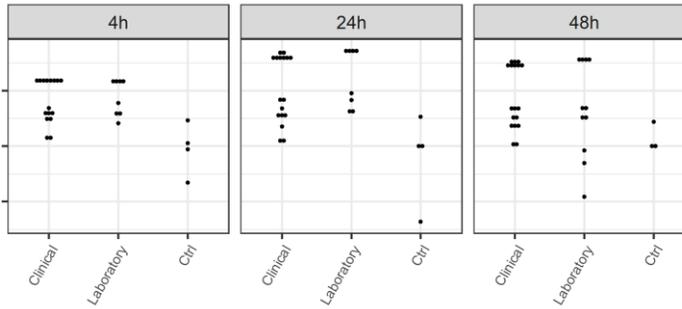
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.218	0.4333	0.617
Time24h	0.2724	0.1579	0.0905
Time48h	0.0968	0.1579	0.5426
Geo_host_adaptationGeneralist	0.0992	0.1441	0.4944
Time24h:Geo_host_adaptationGeneralist	0.161	0.2038	0.4331
Time48h:Geo_host_adaptationGeneralist	0.1051	0.2038	0.6083

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.0992	0.1441	0.4944
Specialist - Generalist	24h	-0.2602	0.1441	0.0769
Specialist - Generalist	48h	-0.2043	0.1441	0.1624

Lab_adaptation

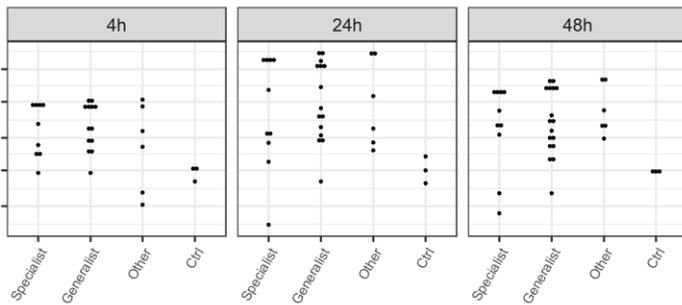


Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.1804	0.4267	0.6739
Time24h	0.3023	0.1088	0.0072
Time48h	0.1221	0.1088	0.2657
Lab_adaptationLaboratory	0.0454	0.1332	0.7344
Time24h:Lab_adaptationLaboratory	0.2353	0.1884	0.2163
Time48h:Lab_adaptationLaboratory	0.1518	0.1884	0.4232

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.0454	0.1332	0.7344
Clinical - Laboratory	24h	-0.2807	0.1332	0.0391
Clinical - Laboratory	48h	-0.1972	0.1332	0.1436

M2

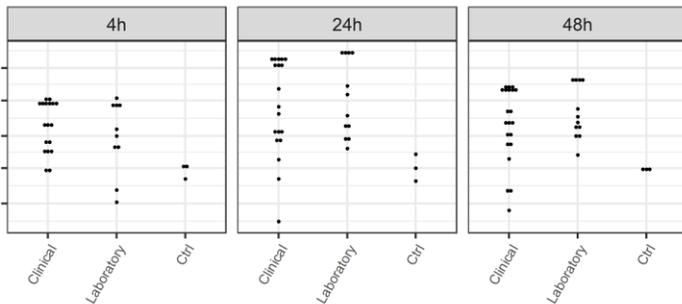
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.3213	0.4337	0.4621
Time24h	0.5423	0.185	0.005
Time48h	0.2887	0.185	0.1248
Geo_host_adaptationGeneralist	0.0188	0.1689	0.9117
Time24h:Geo_host_adaptationGeneralist	0.1128	0.2388	0.6386
Time48h:Geo_host_adaptationGeneralist	0.1161	0.2388	0.629

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.0188	0.1689	0.9117
Specialist - Generalist	24h	-0.1317	0.1689	0.4393
Specialist - Generalist	48h	-0.1349	0.1689	0.4281

Lab_adaptation



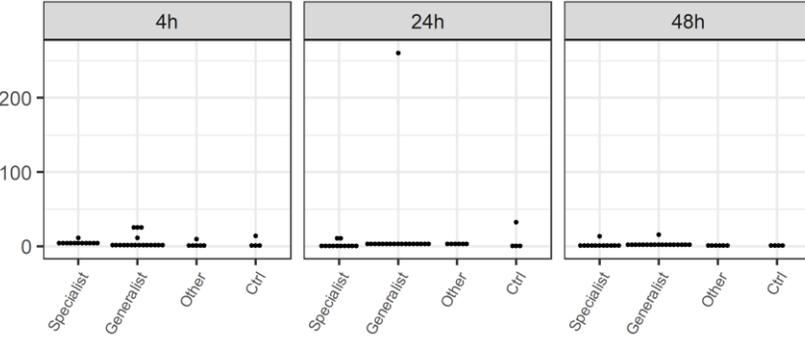
Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.3523	0.4187	0.4032
Time24h	0.4331	0.1186	5e-04
Time48h	0.3006	0.1186	0.0138
Lab_adaptationLaboratory	0.0092	0.1453	0.9497
Time24h:Lab_adaptationLaboratory	0.641	0.2055	0.0027
Time48h:Lab_adaptationLaboratory	0.321	0.2055	0.1232

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.0092	0.1453	0.9497
Clinical - Laboratory	24h	-0.6502	0.1453	<0.0001
Clinical - Laboratory	48h	-0.3302	0.1453	0.0264

IP-10 (logarithmic transformation)

M1

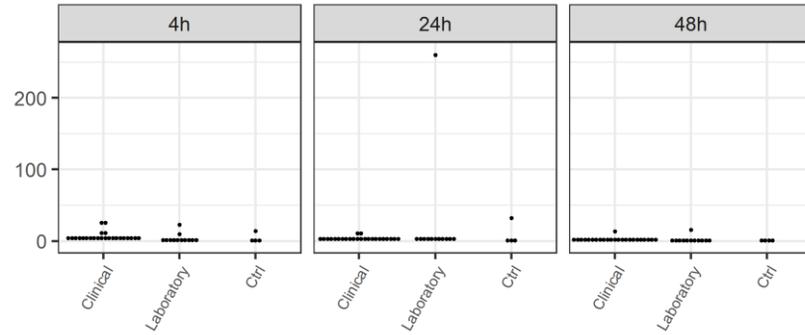
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.8833	1.2153	0.4707
Time24h	-0.1296	0.6331	0.8387
Time48h	-0.4979	0.6331	0.4353
Geo_host_adaptationGeneralist	0.8554	0.578	0.145
Time24h:Geo_host_adaptationGeneralist	0.4414	0.8174	0.5916
Time48h:Geo_host_adaptationGeneralist	-0.1404	0.8174	0.8643

contrast	Time	ratio	SE	p-value
Specialist / Generalist	4h	0.4251	0.2457	0.145
Specialist / Generalist	24h	0.2734	0.158	0.0292
Specialist / Generalist	48h	0.4892	0.2827	0.2217

Lab_adaptation

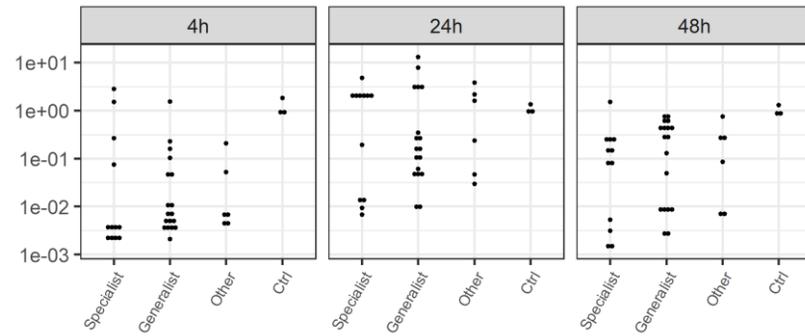


Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.3395	1.168	0.7723
Time24h	-0.2199	0.4391	0.6182
Time48h	-0.718	0.4391	0.107
Lab_adaptationLaboratory	-0.2087	0.5378	0.6993
Time24h:Lab_adaptationLaboratory	1.258	0.7605	0.1031
Time48h:Lab_adaptationLaboratory	0.4712	0.7605	0.5377

contrast	Time	ratio	SE	p-value
Clinical / Laboratory	4h	1.2321	0.6626	0.6993
Clinical / Laboratory	24h	0.3502	0.1883	0.0555
Clinical / Laboratory	48h	0.7691	0.4136	0.6271

M2

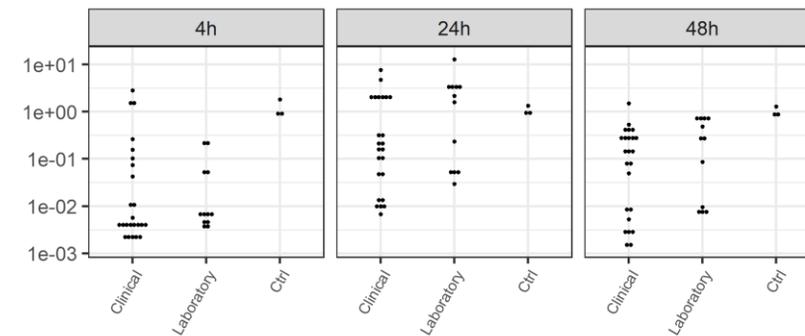
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-3.2589	1.1109	0.005
Time24h	2.4912	0.7194	0.0011
Time48h	0.7057	0.7194	0.3313
Geo_host_adaptationGeneralist	-0.323	0.6567	0.6249
Time24h:Geo_host_adaptationGeneralist	0.789	0.9287	0.9326
Time48h:Geo_host_adaptationGeneralist	0.7564	0.9287	0.4192

contrast	Time	ratio	SE	p-value
Specialist / Generalist	4h	1.3813	0.9071	0.6249
Specialist / Generalist	24h	1.2764	0.8383	0.7117
Specialist / Generalist	48h	0.6483	0.4258	0.5123

Lab_adaptation



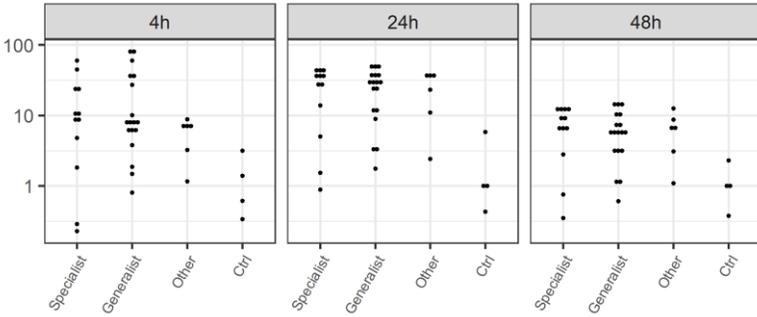
Parameter	Estimate	Std.Error	p-value
(Intercept)	-3.3825	0.9938	0.0012
Time24h	2.1167	0.485	<0.0001
Time48h	0.8923	0.485	0.0705
Lab_adaptationLaboratory	-0.3566	0.594	0.5594
Time24h:Lab_adaptationLaboratory	1.4685	0.84	0.0853
Time48h:Lab_adaptationLaboratory	0.903	0.84	0.2865

contrast	Time	ratio	SE	p-value
Clinical / Laboratory	4h	1.4284	0.8484	0.5594
Clinical / Laboratory	24h	0.3289	0.1954	0.0659
Clinical / Laboratory	48h	0.579	0.3439	0.3611

MCP-1 (logarithmic transformation)

M1

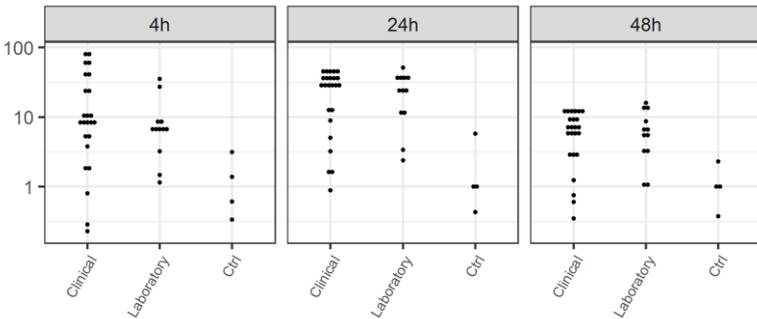
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	1.3888	0.6863	0.0483
Time24h	0.9816	0.371	0.0108
Time48h	-0.06	0.371	0.8721
Geo_host_adaptationGeneralist	0.5813	0.3387	0.0922
Time24h:Geo_host_adaptationGeneralist	-0.3471	0.4789	0.4719
Time48h:Geo_host_adaptationGeneralist	-0.5931	0.4789	0.2213

contrast	Time	ratio	SE	p-value
Specialist / Generalist	4h	0.5592	0.1894	0.0922
Specialist / Generalist	24h	0.7912	0.268	0.4924
Specialist / Generalist	48h	1.0119	0.3427	0.9724

Lab_adaptation

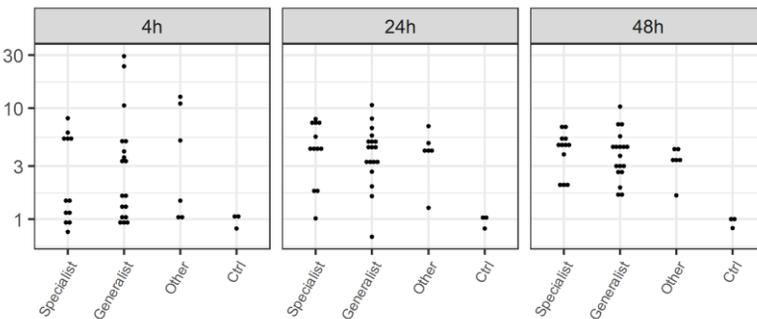


Parameter	Estimate	Std.Error	p-value
(Intercept)	1.6941	0.6324	0.0094
Time24h	0.7773	0.2573	0.0036
Time48h	-0.3977	0.2573	0.1273
Lab_adaptationLaboratory	-0.0707	0.3152	0.8232
Time24h:Lab_adaptationLaboratory	0.215	0.4457	0.6312
Time48h:Lab_adaptationLaboratory	0.1671	0.4457	0.709

contrast	Time	ratio	SE	p-value
Clinical / Laboratory	4h	1.0733	0.3383	0.8232
Clinical / Laboratory	24h	0.8656	0.2728	0.6487
Clinical / Laboratory	48h	0.9081	0.2862	0.7608

M2

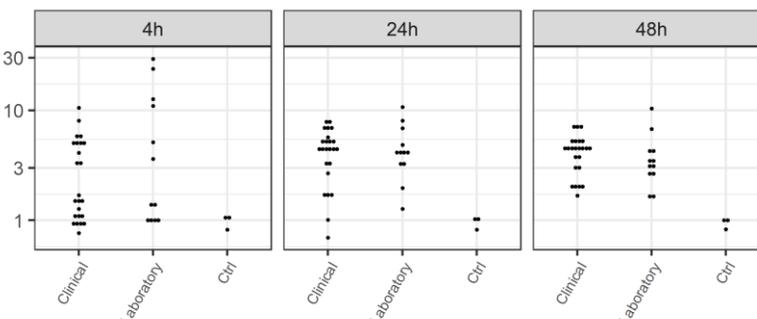
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	0.7511	0.3205	0.023
Time24h	0.4851	0.3106	0.1246
Time48h	0.5348	0.3106	0.0912
Geo_host_adaptationGeneralist	0.2181	0.2836	0.4454
Time24h:Geo_host_adaptationGeneralist	-0.2826	0.401	0.4842
Time48h:Geo_host_adaptationGeneralist	-0.2724	0.401	0.5

contrast	Time	ratio	SE	p-value
Specialist / Generalist	4h	0.8041	0.228	0.4454
Specialist / Generalist	24h	1.0667	0.3025	0.8209
Specialist / Generalist	48h	1.0559	0.2994	0.8487

Lab_adaptation



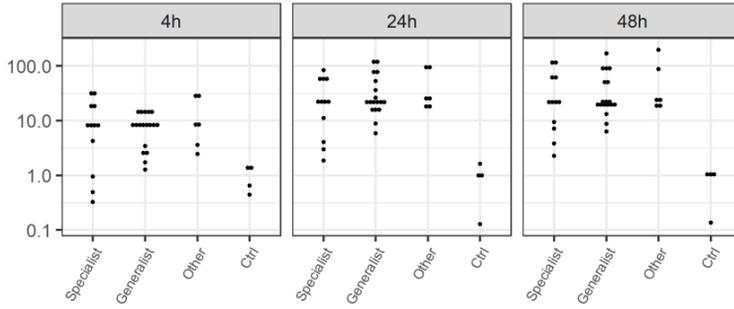
Parameter	Estimate	Std.Error	p-value
(Intercept)	0.809	0.2809	0.0054
Time24h	0.3495	0.2138	0.1071
Time48h	0.4657	0.2138	0.0332
Lab_adaptationLaboratory	0.3206	0.2619	0.2254
Time24h:Lab_adaptationLaboratory	-0.1925	0.3704	0.6051
Time48h:Lab_adaptationLaboratory	-0.4586	0.3704	0.2202

contrast	Time	ratio	SE	p-value
Clinical / Laboratory	4h	0.7257	0.19	0.2254
Clinical / Laboratory	24h	0.8797	0.2304	0.6263
Clinical / Laboratory	48h	1.148	0.3006	0.6001

MIP-1 α (logarithmic transformation)

M1

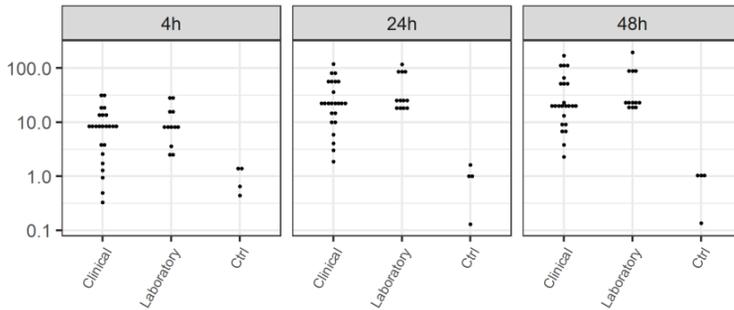
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	1.3076	0.5729	0.0267
Time24h	1.2163	0.2701	<0.0001
Time48h	1.3856	0.2701	<0.0001
Geo_host_adaptationGeneralist	0.3435	0.2466	0.1697
Time24h:Geo_host_adaptationGeneralist	0.246	0.3487	0.4838
Time48h:Geo_host_adaptationGeneralist	0.1089	0.3487	0.7561

contrast	Time	ratio	SE	p.value
Specialist / Generalist	4h	0.7093	0.1749	0.1697
Specialist / Generalist	24h	0.5546	0.1368	0.0206
Specialist / Generalist	48h	0.6361	0.1569	0.0724

Lab_adaptation

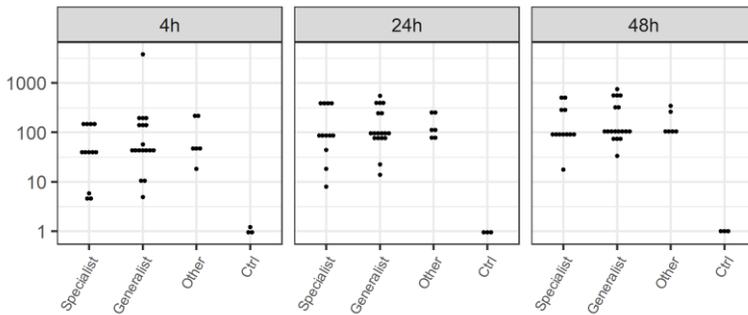


Parameter	Estimate	Std.Error	p-value
(Intercept)	1.4163	0.5382	0.0107
Time24h	1.3372	0.1814	<0.0001
Time48h	1.4548	0.1814	<0.0001
Lab_adaptationLaboratory	0.5009	0.2221	0.0276
Time24h:Lab_adaptationLaboratory	0.1245	0.3141	0.6933
Time48h:Lab_adaptationLaboratory	0.0332	0.3141	0.9161

contrast	Time	ratio	SE	p.value
Clinical / Laboratory	4h	0.606	0.1346	0.0276
Clinical / Laboratory	24h	0.535	0.1189	0.0065
Clinical / Laboratory	48h	0.5862	0.1302	0.0191

M2

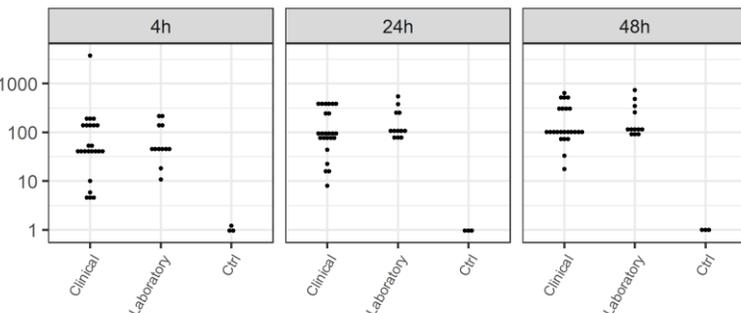
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	3.2355	0.5833	<0.0001
Time24h	1.0178	0.4495	0.0278
Time48h	1.4443	0.4495	0.0023
Geo_host_adaptationGeneralist	0.8325	0.4104	0.0477
Time24h:Geo_host_adaptationGeneralist	-0.5476	0.5803	0.3499
Time48h:Geo_host_adaptationGeneralist	-0.6548	0.5803	0.2645

contrast	Time	ratio	SE	p.value
Specialist / Generalist	4h	0.435	0.1785	0.0477
Specialist / Generalist	24h	0.7521	0.3086	0.4906
Specialist / Generalist	48h	0.8371	0.3435	0.6667

Lab_adaptation



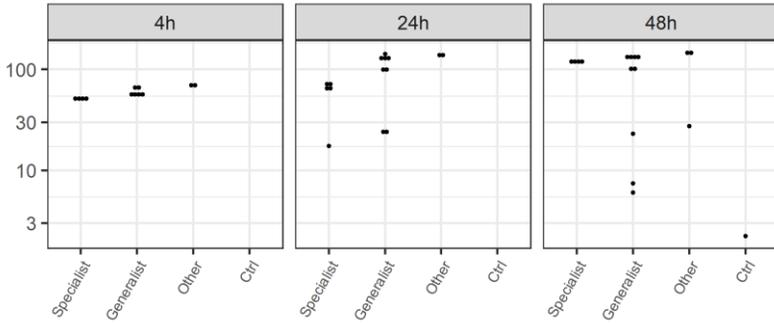
Parameter	Estimate	Std.Error	p-value
(Intercept)	3.7428	0.507	<0.0001
Time24h	0.5634	0.3037	0.0683
Time48h	0.9728	0.3037	0.0021
Lab_adaptationLaboratory	0.0968	0.372	0.7955
Time24h:Lab_adaptationLaboratory	0.4144	0.5261	0.4339
Time48h:Lab_adaptationLaboratory	0.1681	0.5261	0.7504

contrast	Time	ratio	SE	p.value
Clinical / Laboratory	4h	0.9077	0.3377	0.7955
Clinical / Laboratory	24h	0.5998	0.2231	0.1743
Clinical / Laboratory	48h	0.7673	0.2854	0.479

PDGF-bb (logarithmic transformation)

M1

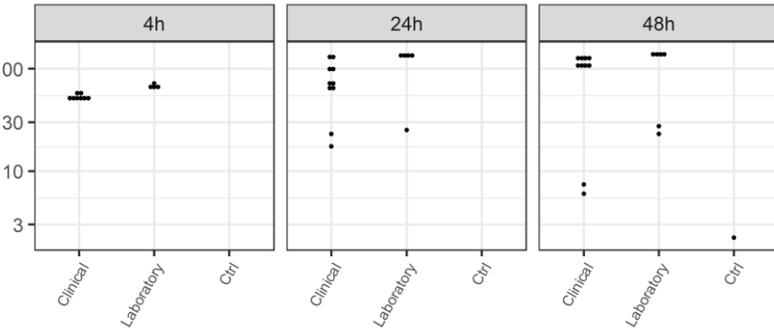
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.558	0.5215	0.2916
Time24h	0.2519	0.1985	0.2123
Time48h	0.3579	0.1985	0.0795
Geo_host_adaptationGeneralist	0.0681	0.1812	0.7094
Time24h:Geo_host_adaptationGeneralist	0.3353	0.2562	0.1988
Time48h:Geo_host_adaptationGeneralist	0.235	0.2562	0.365

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.0681	0.1812	0.7094
Specialist - Generalist	24h	-0.4034	0.1812	0.0322
Specialist - Generalist	48h	-0.3031	0.1812	0.1028

Lab_adaptation

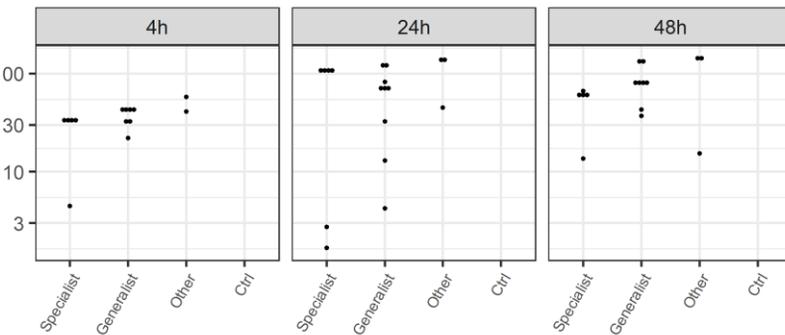


Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.5309	0.5114	0.3046
Time24h	0.3182	0.137	0.0246
Time48h	0.3875	0.137	0.0069
Lab_adaptationLaboratory	0.1071	0.1678	0.5265
Time24h:Lab_adaptationLaboratory	0.198	0.2372	0.4083
Time48h:Lab_adaptationLaboratory	0.3445	0.2372	0.1533

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.1071	0.1678	0.5265
Clinical - Laboratory	24h	-0.305	0.1678	0.0755
Clinical - Laboratory	48h	-0.4516	0.1678	0.0099

M2

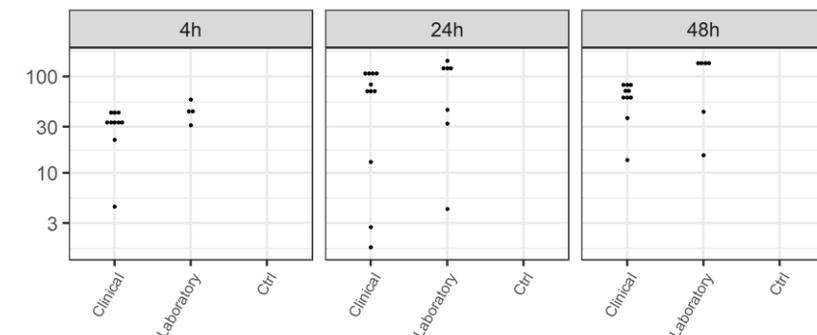
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.5241	0.5273	0.3267
Time24h	0.515	0.2487	0.0454
Time48h	0.1805	0.2487	0.4724
Geo_host_adaptationGeneralist	0.0283	0.227	0.9015
Time24h:Geo_host_adaptationGeneralist	-0.0309	0.321	0.9239
Time48h:Geo_host_adaptationGeneralist	0.3858	0.321	0.2371

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.0283	0.227	0.9015
Specialist - Generalist	24h	0.0026	0.227	0.991
Specialist - Generalist	48h	-0.4141	0.227	0.0762

Lab_adaptation



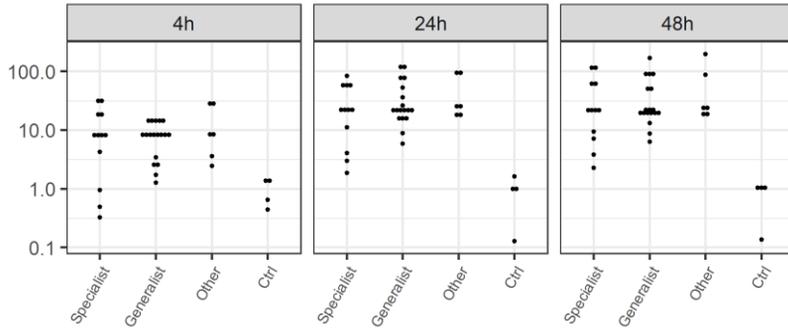
Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.5167	0.5097	0.316
Time24h	0.3338	0.1473	0.0282
Time48h	0.2434	0.1473	0.1053
Lab_adaptationLaboratory	-0.059	0.1805	0.7451
Time24h:Lab_adaptationLaboratory	0.6088	0.2552	0.0212
Time48h:Lab_adaptationLaboratory	0.6619	0.2552	0.0127

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	0.059	0.1805	0.7451
Clinical - Laboratory	24h	-0.5497	0.1805	0.0038
Clinical - Laboratory	48h	-0.6029	0.1805	0.0017

MIP-1 β (logarithmic transformation)

M1

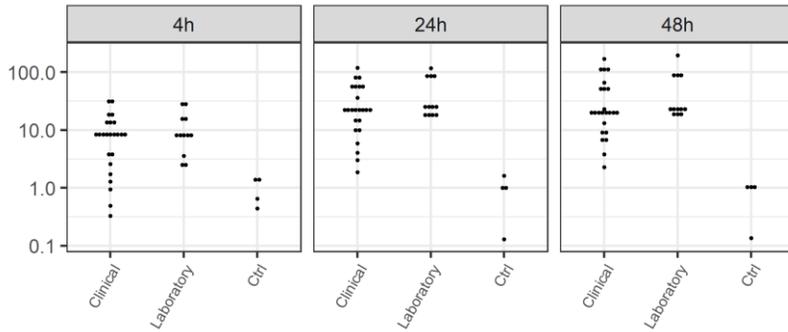
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	1.3076	0.5729	0.0267
Time24h	1.2163	0.2701	<0.0001
Time48h	1.3856	0.2701	<0.0001
Geo_host_adaptationGeneralist	0.3435	0.2466	0.1697
Time24h:Geo_host_adaptationGeneralist	0.246	0.3487	0.4838
Time48h:Geo_host_adaptationGeneralist	0.1089	0.3487	0.7561

contrast	Time	ratio	SE	p-value
Specialist / Generalist	4h	0.7093	0.1749	0.1697
Specialist / Generalist	24h	0.5546	0.1368	0.0206
Specialist / Generalist	48h	0.6361	0.1569	0.0724

Lab_adaptation

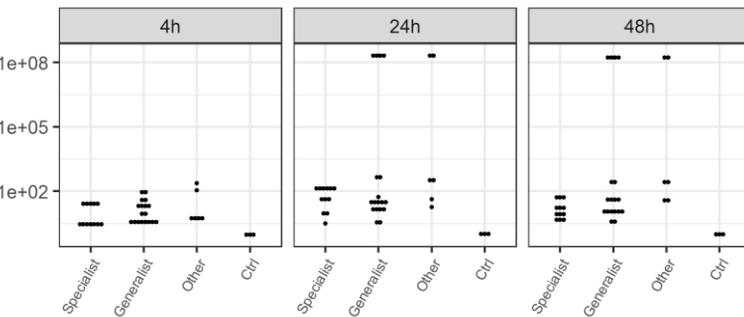


Parameter	Estimate	Std.Error	p-value
(Intercept)	1.4163	0.5382	0.0107
Time24h	1.3372	0.1814	<0.0001
Time48h	1.4548	0.1814	<0.0001
Lab_adaptationLaboratory	0.5009	0.2221	0.0276
Time24h:Lab_adaptationLaboratory	0.1245	0.3141	0.6933
Time48h:Lab_adaptationLaboratory	0.0332	0.3141	0.9161

contrast	Time	ratio	SE	p-value
Clinical / Laboratory	4h	0.606	0.1346	0.0276
Clinical / Laboratory	24h	0.535	0.1189	0.0065
Clinical / Laboratory	48h	0.5862	0.1302	0.0191

M2

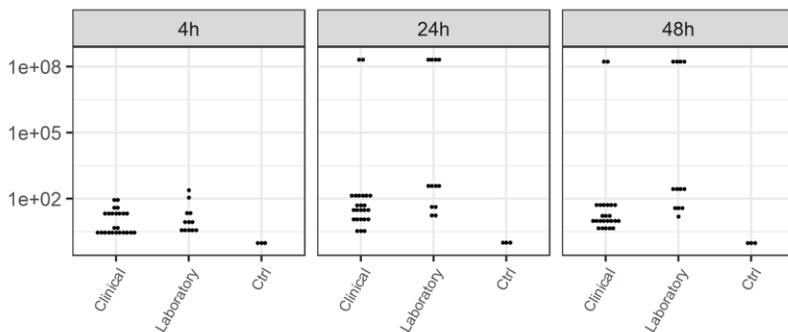
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-1.0577	0.3966	0.0102
Time24h	1.2369	0.3517	9e-04
Time48h	0.83	0.3517	0.0221
Geo_host_adaptationGeneralist	0.5531	0.3211	0.091
Time24h:Geo_host_adaptationGeneralist	-0.3937	0.4541	0.3899
Time48h:Geo_host_adaptationGeneralist	-0.1473	0.4541	0.747

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.5531	0.3211	0.091
Specialist - Generalist	24h	-0.1594	0.3211	0.6217
Specialist - Generalist	48h	-0.4058	0.3211	0.212

Lab_adaptation



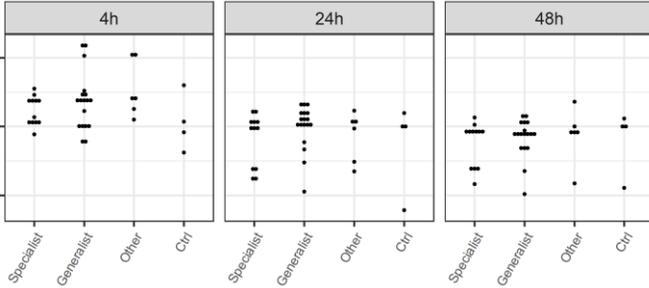
Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.8692	0.3472	0.0149
Time24h	0.8608	0.213	1e-04
Time48h	0.6255	0.213	0.0046
Lab_adaptationLaboratory	0.3031	0.2608	0.2496
Time24h:Lab_adaptationLaboratory	0.5193	0.3689	0.1641
Time48h:Lab_adaptationLaboratory	0.5802	0.3689	0.1207

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.3031	0.2608	0.2496
Clinical - Laboratory	24h	-0.8224	0.2608	0.0025
Clinical - Laboratory	48h	-0.8833	0.2608	0.0012

RANTES (logarithmic transformation)

M1

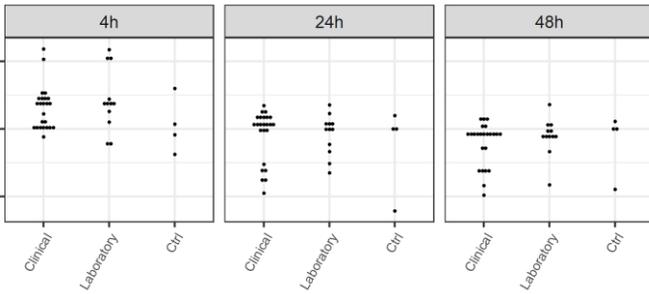
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	0.6918	0.3923	0.0838
Time24h	-0.9155	0.2712	0.0014
Time48h	-1.2338	0.2712	<0.0001
Geo_host_adaptationGeneralist	0.3405	0.2476	0.175
Time24h:Geo_host_adaptationGeneralist	0.0679	0.3502	0.8469
Time48h:Geo_host_adaptationGeneralist	-0.1782	0.3502	0.6131

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.3405	0.2476	0.175
Specialist - Generalist	24h	-0.4085	0.2476	0.1051
Specialist - Generalist	48h	-0.1624	0.2476	0.5149

Lab_adaptation

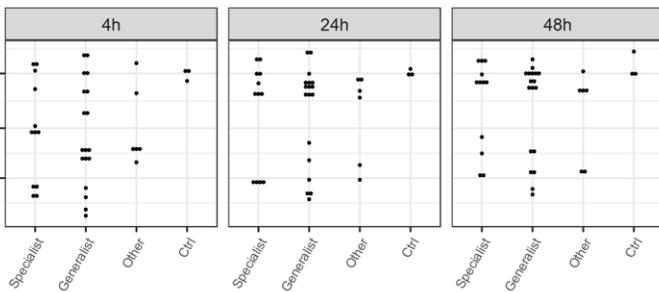


Parameter	Estimate	Std.Error	p-value
(Intercept)	0.8742	0.351	0.0154
Time24h	-0.8917	0.195	<0.0001
Time48h	-1.3747	0.195	<0.0001
Lab_adaptationLaboratory	0.1473	0.2388	0.5395
Time24h:Lab_adaptationLaboratory	-0.198	0.3377	0.5598
Time48h:Lab_adaptationLaboratory	0.0408	0.3377	0.9042

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.1473	0.2388	0.5395
Clinical - Laboratory	24h	0.0507	0.2388	0.8326
Clinical - Laboratory	48h	-0.1881	0.2388	0.4338

M2

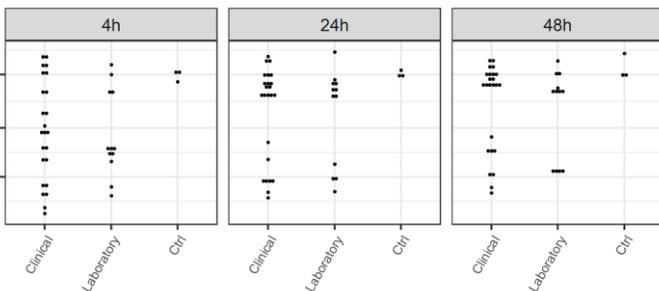
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	0.0341	0.5314	0.9491
Time24h	0.3003	0.2471	0.2298
Time48h	0.5398	0.2471	0.0335
Geo_host_adaptationGeneralist	-0.0249	0.2256	0.9127
Time24h:Geo_host_adaptationGeneralist	-0.0387	0.319	0.9039
Time48h:Geo_host_adaptationGeneralist	-0.1805	0.319	0.5739

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	0.0249	0.2256	0.9127
Specialist - Generalist	24h	0.0636	0.2256	0.7792
Specialist - Generalist	48h	0.2054	0.2256	0.3668

Lab_adaptation



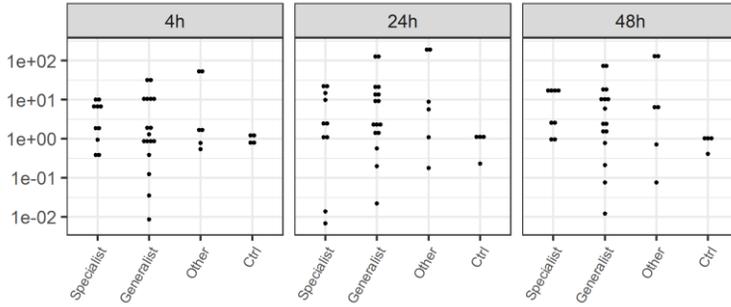
Parameter	Estimate	Std.Error	p-value
(Intercept)	0.1017	0.517	0.8447
Time24h	0.1852	0.1759	0.2964
Time48h	0.3968	0.1759	0.0276
Lab_adaptationLaboratory	-0.1775	0.2154	0.413
Time24h:Lab_adaptationLaboratory	0.1739	0.3046	0.57
Time48h:Lab_adaptationLaboratory	-0.0267	0.3046	0.9304

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	0.1775	0.2154	0.413
Clinical - Laboratory	24h	0.0036	0.2154	0.9868
Clinical - Laboratory	48h	0.2042	0.2154	0.3467

TNF α (logarithmic transformation)

M1

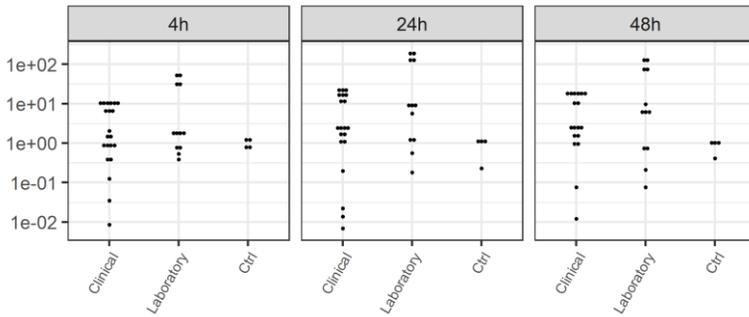
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.2225	0.4461	0.62
Time24h	0.1489	0.2033	0.4672
Time48h	0.076	0.2033	0.71
Geo_host_adaptationGeneralist	0.1279	0.1856	0.4938
Time24h:Geo_host_adaptationGeneralist	0.1954	0.2625	0.46
Time48h:Geo_host_adaptationGeneralist	0.1498	0.2625	0.5706

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.1279	0.1856	0.4938
Specialist - Generalist	24h	-0.3233	0.1856	0.0875
Specialist - Generalist	48h	-0.2778	0.1856	0.1407

Lab_adaptation

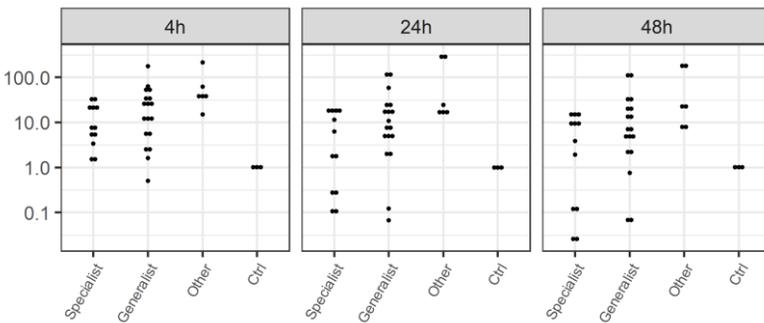


Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.2978	0.4328	0.4939
Time24h	0.176	0.1081	0.1083
Time48h	0.107	0.1081	0.326
Lab_adaptationLaboratory	0.4612	0.1323	9e-04
Time24h:Lab_adaptationLaboratory	0.2714	0.1872	0.152
Time48h:Lab_adaptationLaboratory	0.1501	0.1872	0.4254

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.4612	0.1323	9e-04
Clinical - Laboratory	24h	-0.7326	0.1323	<0.0001
Clinical - Laboratory	48h	-0.6113	0.1323	<0.0001

M2

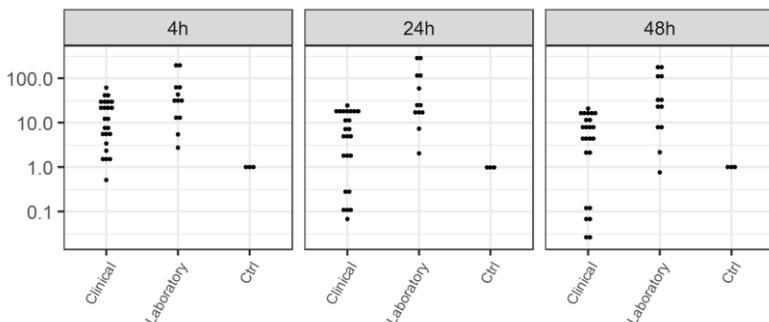
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.0157	0.4183	0.9703
Time24h	-0.397	0.3221	0.2233
Time48h	-0.5288	0.3221	0.1068
Geo_host_adaptationGeneralist	0.3967	0.294	0.1832
Time24h:Geo_host_adaptationGeneralist	0.1372	0.4158	0.7428
Time48h:Geo_host_adaptationGeneralist	0.176	0.4158	0.6739

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.3967	0.294	0.1832
Specialist - Generalist	24h	-0.5339	0.294	0.0753
Specialist - Generalist	48h	-0.5727	0.294	0.0569

Lab_adaptation



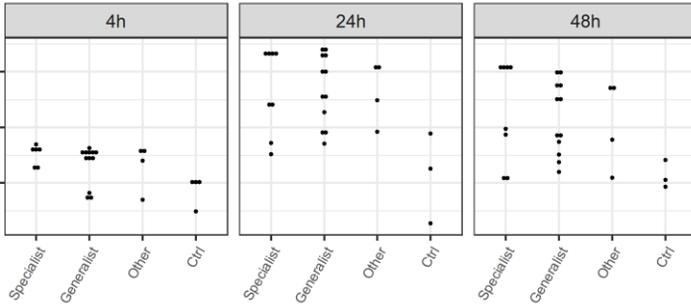
Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.0655	0.3515	0.8528
Time24h	-0.4378	0.1778	0.0166
Time48h	-0.5388	0.1778	0.0035
Lab_adaptationLaboratory	0.7376	0.2178	0.0012
Time24h:Lab_adaptationLaboratory	0.5497	0.308	0.0792
Time48h:Lab_adaptationLaboratory	0.4619	0.308	0.1387

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.7376	0.2178	0.0012
Clinical - Laboratory	24h	-1.2873	0.2178	<0.0001
Clinical - Laboratory	48h	-1.1995	0.2178	<0.0001

VEGF (logarithmic transformation)

M1

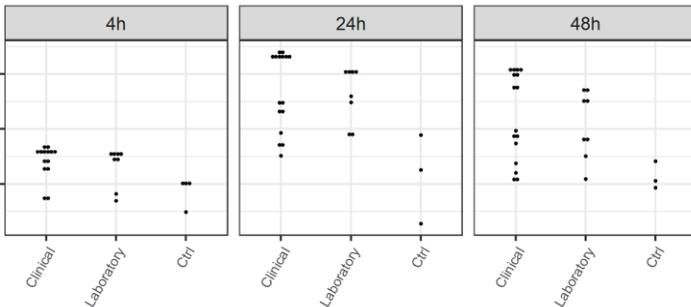
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-1.1476	0.4705	0.0196
Time24h	1.6351	0.1782	<0.0001
Time48h	1.0868	0.1782	<0.0001
Geo_host_adaptationGeneralist	0.1966	0.1627	0.2346
Time24h:Geo_host_adaptationGeneralist	-0.0845	0.2301	0.7154
Time48h:Geo_host_adaptationGeneralist	-0.2596	0.2301	0.2665

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.1966	0.1627	0.2346
Specialist - Generalist	24h	-0.112	0.1627	0.4953
Specialist - Generalist	48h	0.063	0.1627	0.7008

Lab_adaptation

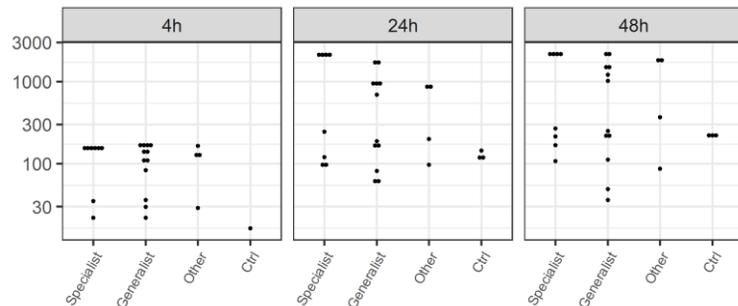


Parameter	Estimate	Std.Error	p-value
(Intercept)	-1.059	0.4622	0.0266
Time24h	1.6708	0.1246	<0.0001
Time48h	0.9798	0.1246	<0.0001
Lab_adaptationLaboratory	0.0884	0.1526	0.5651
Time24h:Lab_adaptationLaboratory	-0.2281	0.2159	0.2963
Time48h:Lab_adaptationLaboratory	-0.1895	0.2159	0.3845

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.0884	0.1526	0.5651
Clinical - Laboratory	24h	0.1396	0.1526	0.3651
Clinical - Laboratory	48h	0.1011	0.1526	0.5111

M2

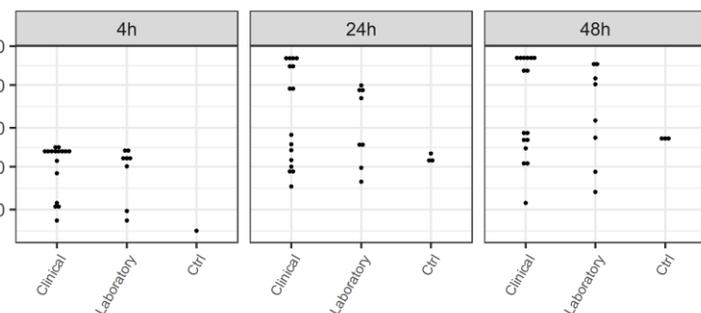
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.8316	0.5494	0.1386
Time24h	1.0045	0.2026	<0.0001
Time48h	1.3884	0.2026	<0.0001
Geo_host_adaptationGeneralist	-0.0697	0.1849	0.7082
Time24h:Geo_host_adaptationGeneralist	-0.2576	0.2615	0.3311
Time48h:Geo_host_adaptationGeneralist	-0.4135	0.2615	0.1223

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	0.0697	0.1849	0.7082
Specialist - Generalist	24h	0.3273	0.1849	0.085
Specialist - Generalist	48h	0.4833	0.1849	0.0129

Lab_adaptation



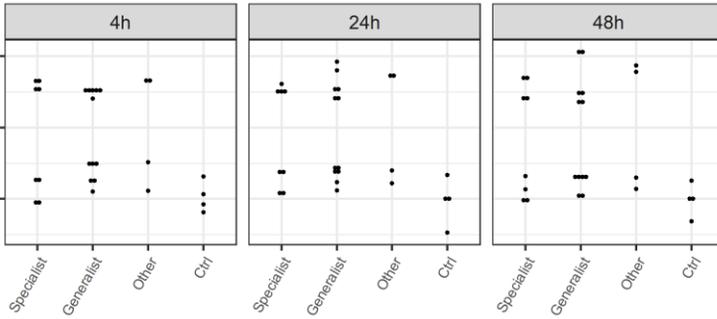
Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.8031	0.5438	0.1466
Time24h	0.8377	0.1492	<0.0001
Time48h	1.1834	0.1492	<0.0001
Lab_adaptationLaboratory	-0.2022	0.1828	0.2744
Time24h:Lab_adaptationLaboratory	0.0332	0.2585	0.8985
Time48h:Lab_adaptationLaboratory	-0.1473	0.2585	0.5716

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	0.2022	0.1828	0.2744
Clinical - Laboratory	24h	0.169	0.1828	0.36
Clinical - Laboratory	48h	0.3494	0.1828	0.0621

SDS-1 α (logarithmic transformation)

M1

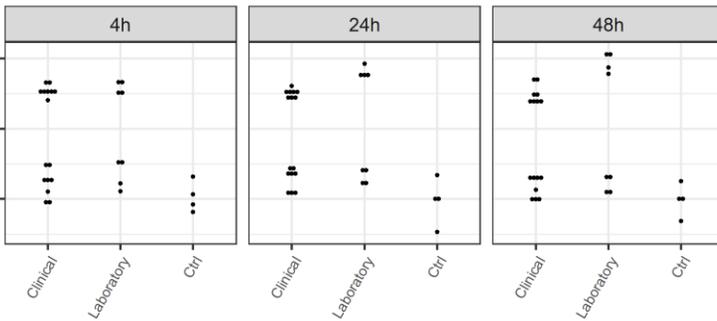
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.6082	0.5798	0.301
Time24h	0.3134	0.343	0.3669
Time48h	0.0983	0.343	0.7761
Geo_host_adaptationGeneralist	0.478	0.3132	0.1354
Time24h:Geo_host_adaptationGeneralist	-0.1994	0.4429	0.6552
Time48h:Geo_host_adaptationGeneralist	-0.1983	0.4429	0.6569

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.478	0.3132	0.1354
Specialist - Generalist	24h	-0.2786	0.3132	0.3794
Specialist - Generalist	48h	-0.2797	0.3132	0.3776

Lab_adaptation

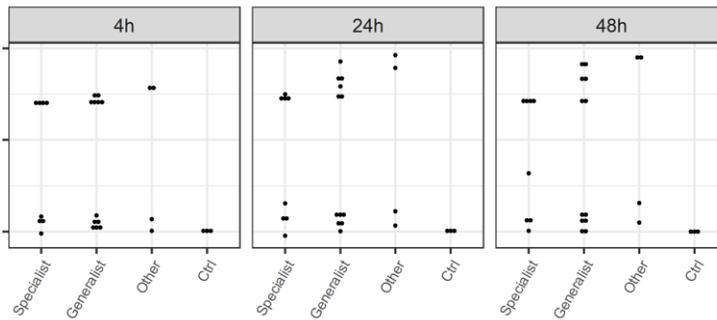


Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.4496	0.5522	0.4197
Time24h	0.1715	0.2307	0.461
Time48h	-0.0914	0.2307	0.6936
Lab_adaptationLaboratory	0.3502	0.2825	0.2214
Time24h:Lab_adaptationLaboratory	0.0703	0.3995	0.8612
Time48h:Lab_adaptationLaboratory	0.306	0.3995	0.4477

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.3502	0.2825	0.2214
Clinical - Laboratory	24h	-0.4205	0.2825	0.1435
Clinical - Laboratory	48h	-0.6562	0.2825	0.0247

M2

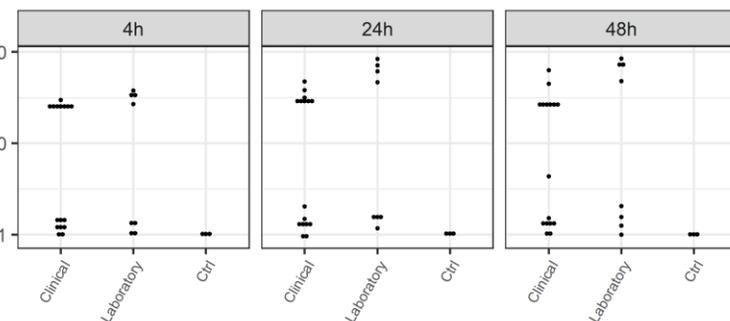
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.5888	0.59	0.3249
Time24h	0.2506	0.3115	0.4262
Time48h	0.2537	0.3115	0.4206
Geo_host_adaptationGeneralist	0.1978	0.2843	0.4911
Time24h:Geo_host_adaptationGeneralist	0.1468	0.4021	0.7171
Time48h:Geo_host_adaptationGeneralist	0.0464	0.4021	0.9087

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.1978	0.2843	0.4911
Specialist - Generalist	24h	-0.3446	0.2843	0.2332
Specialist - Generalist	48h	-0.2442	0.2843	0.396

Lab_adaptation



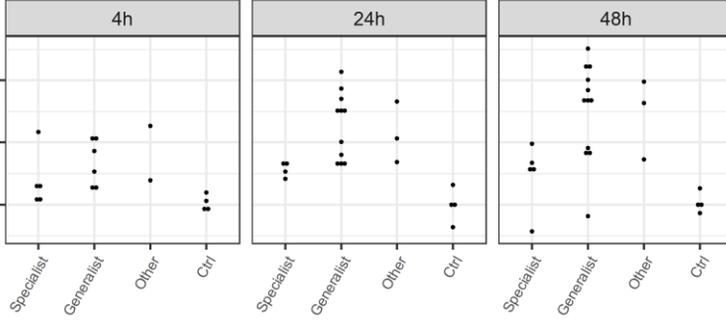
Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.524	0.5676	0.3607
Time24h	0.158	0.209	0.4534
Time48h	0.231	0.209	0.2747
Lab_adaptationLaboratory	0.1257	0.2559	0.6256
Time24h:Lab_adaptationLaboratory	0.4703	0.362	0.2003
Time48h:Lab_adaptationLaboratory	0.3237	0.362	0.3758

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.1257	0.2559	0.6256
Clinical - Laboratory	24h	-0.5961	0.2559	0.0243
Clinical - Laboratory	48h	-0.4494	0.2559	0.0858

IL-18 (logarithmic transformation)

M1

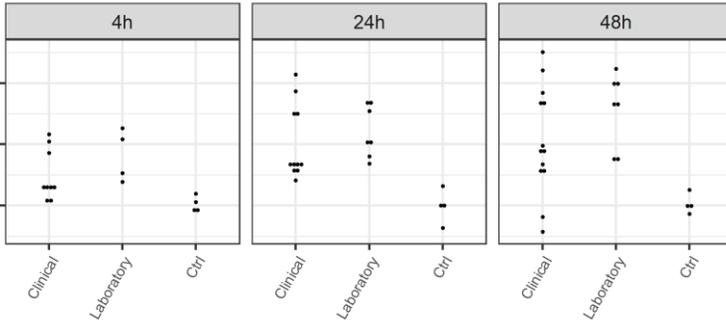
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.4429	0.2657	0.1039
Time24h	-0.053	0.3295	0.873
Time48h	0.0883	0.3295	0.7903
Geo_host_adaptationGeneralist	0.0281	0.3007	0.9261
Time24h:Geo_host_adaptationGeneralist	1.0793	0.4253	0.0155
Time48h:Geo_host_adaptationGeneralist	1.4113	0.4253	0.002

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.0281	0.3007	0.9261
Specialist - Generalist	24h	-1.1074	0.3007	7e-04
Specialist - Generalist	48h	-1.4394	0.3007	<0.0001

Lab_adaptation

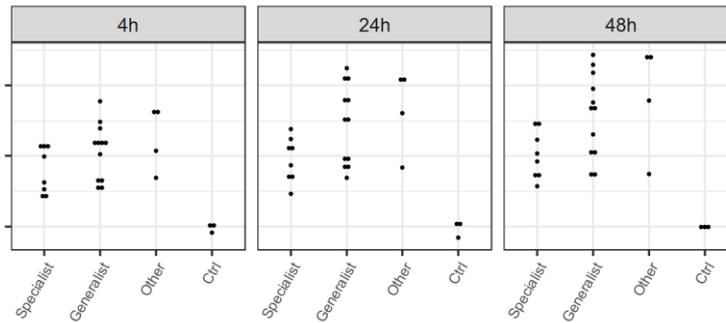


Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.4358	0.2491	0.0869
Time24h	0.4554	0.3	0.1359
Time48h	0.7678	0.3	0.0138
Lab_adaptationLaboratory	0.1101	0.3675	0.7658
Time24h:Lab_adaptationLaboratory	0.3462	0.5197	0.5086
Time48h:Lab_adaptationLaboratory	0.389	0.5197	0.4579

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.1101	0.3675	0.7658
Clinical - Laboratory	24h	-0.4563	0.3675	0.2206
Clinical - Laboratory	48h	-0.4991	0.3675	0.181

M2

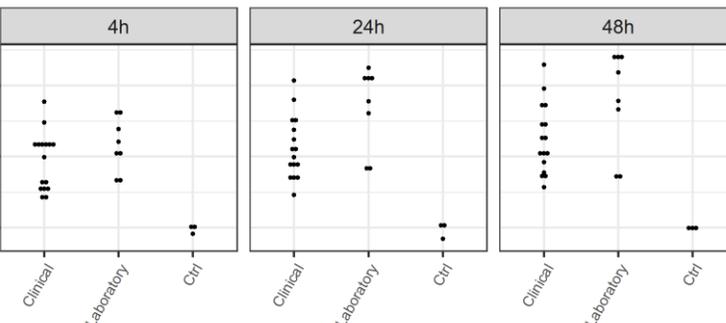
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-1.1197	0.4162	0.0106
Time24h	0.567	0.4155	0.1806
Time48h	0.7678	0.4155	0.0726
Geo_host_adaptationGeneralist	0.8471	0.3793	0.0317
Time24h:Geo_host_adaptationGeneralist	0.0445	0.5364	0.9344
Time48h:Geo_host_adaptationGeneralist	0.0519	0.5364	0.9235

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.8471	0.3793	0.0317
Specialist - Generalist	24h	-0.8915	0.3793	0.0242
Specialist - Generalist	48h	-0.899	0.3793	0.0231

Lab_adaptation



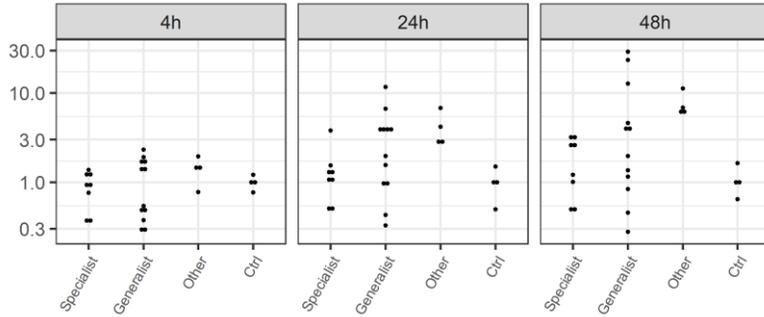
Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.8219	0.383	0.0372
Time24h	0.5042	0.2958	0.095
Time48h	0.7526	0.2958	0.0144
Lab_adaptationLaboratory	0.5731	0.3622	0.1204
Time24h:Lab_adaptationLaboratory	0.2571	0.5123	0.6181
Time48h:Lab_adaptationLaboratory	0.2004	0.5123	0.6975

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.5731	0.3622	0.1204
Clinical - Laboratory	24h	-0.8303	0.3622	0.0265
Clinical - Laboratory	48h	-0.7735	0.3622	0.0381

Gro- α (logarithmic transformation)

M1

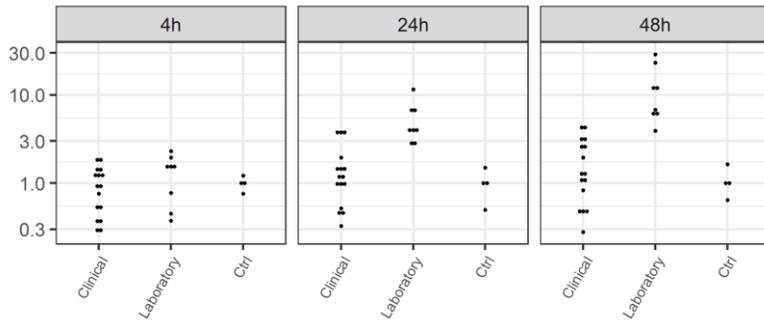
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.1115	0.4353	0.7992
Time24h	0.3827	0.4786	0.4291
Time48h	0.5643	0.4786	0.246
Geo_host_adaptationGeneralist	0.142	0.4369	0.747
Time24h:Geo_host_adaptationGeneralist	-0.4921	0.6179	0.4309
Time48h:Geo_host_adaptationGeneralist	0.5218	0.6179	0.4039

contrast	Time	ratio	SE	p-value
Specialist / Generalist	4h	0.8676	0.3791	0.747
Specialist / Generalist	24h	0.5304	0.2317	0.1551
Specialist / Generalist	48h	0.5149	0.225	0.1372

Lab_adaptation

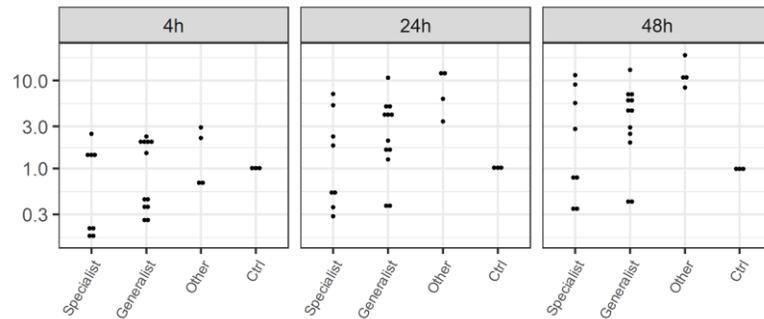


Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.0717	0.3046	0.815
Time24h	0.4123	0.2338	0.0845
Time48h	0.5163	0.2338	0.0323
Lab_adaptationLaboratory	0.3111	0.2864	0.283
Time24h:Lab_adaptationLaboratory	1.0089	0.405	0.0164
Time48h:Lab_adaptationLaboratory	1.5063	0.405	5e-04

contrast	Time	ratio	SE	p-value
Clinical / Laboratory	4h	0.7326	0.2098	0.283
Clinical / Laboratory	24h	0.2671	0.0765	<0.0001
Clinical / Laboratory	48h	0.1625	0.0465	<0.0001

M2

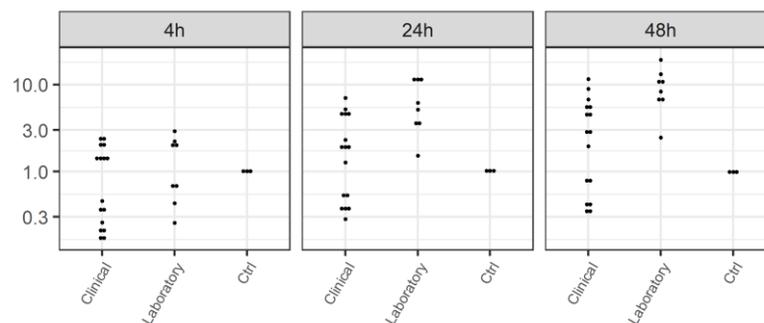
Geo_host_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.702	0.4994	0.1682
Time24h	0.8553	0.2804	0.0042
Time48h	1.2686	0.2804	1e-04
Geo_host_adaptationGeneralist	0.4624	0.2559	0.0789
Time24h:Geo_host_adaptationGeneralist	-0.1523	0.362	0.6764
Time48h:Geo_host_adaptationGeneralist	-0.1506	0.362	0.6798

contrast	Time	estimate	SE	p-value
Specialist - Generalist	4h	-0.4624	0.2559	0.0789
Specialist - Generalist	24h	-0.3102	0.2559	0.2333
Specialist - Generalist	48h	-0.3118	0.2559	0.2308

Lab_adaptation



Parameter	Estimate	Std.Error	p-value
(Intercept)	-0.6036	0.4145	0.1521
Time24h	0.6126	0.1738	0.001
Time48h	0.9337	0.1738	<0.0001
Lab_adaptationLaboratory	0.3972	0.2129	0.0685
Time24h:Lab_adaptationLaboratory	0.6271	0.3011	0.0428
Time48h:Lab_adaptationLaboratory	0.7349	0.3011	0.0186

contrast	Time	estimate	SE	p-value
Clinical - Laboratory	4h	-0.3972	0.2129	0.0685
Clinical - Laboratory	24h	-1.0243	0.2129	<0.0001
Clinical - Laboratory	48h	-1.132	0.2129	<0.0001