



A first checklist of the alien-dominated vegetation in Italy

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Subject editor: Gianluigi Bacchetta ♦ Received 27 January 2020 ♦ Accepted 20 May 2020 ♦ Published 12 June 2020

Abstract

This study provides a first step toward the knowledge of the alien-dominated and co-dominated plant communities present in Italy. The first ever checklist of the alien phytocoenoses described or reported in literature for the Italian territory has been compiled, produced by data-mining in national and local thematic literature. The resulting vegetation-type draft-list has been checked in the light of the most recent syntaxonomic documentation and updated with regards to syntaxonomy and nomenclature, with special reference to the frame proposed in the Italian Vegetation Prodrome. The list includes 27 vascular and one bryophyte vegetation classes, hosting 194 low rank alien-dominated *syntaxa*. The different vegetation types detected for each syntaxonomic class and macro-vegetation group, defined by physiognomical and ecological attributes, are discussed.

Keywords

biodiversity, conservation, habitat, Invasive Alien Species, phytosociology, plant communities, syntaxonomy, threats

Introduction

Biological invasions are an ever-increasing global process arising from the intentional or accidental human-mediated introduction of species to areas outside their native range, overcoming natural dispersal mechanisms and biogeographic barriers (Richardson and Pyšek 2006; Blackburn et al. 2014; Seebens et al. 2018).

The consideration and debate on Invasive Alien Species (IAS) date back to long ago (Allan 1936; Egler 1942; Baker 1948; Elton 1958). In the last few decades, invasion science has emerged, becoming a relevant discipline of its own (Richardson et al. 2000; Richardson and Pyšek 2006; Richardson 2011), also as a consequence of the increasing rates of alien species' introductions at the global scale (Seebens et al. 2017, 2018) with no exception in Europe (DAISIE 2009) and in the Mediterranean (Hulme et al. 2008).

Biological invasions are nowadays widely recognized as an important component of human-induced global environmental change (Vitousek et al. 1997; Parmesan and Yohe 2003; Vilà et al. 2011; Vilà and Hulme 2017), being the second most common threat associated to species that underwent extinction in recent times (Bellard et al. 2016). Despite the accumulation of rigorous evidence of its importance to science and society, invasion biology has been the target of criticisms from scientists and academics who do not agree on the role of alien *taxa* as one of the greatest extinction threat at global scale (Da-

vis 2011; Richardson and Ricciardi 2013; Pearce 2015; Russell and Blackburn 2017; Ricciardi and Ryan 2018). However, it is renowned that the establishment and spread of IAS can drastically affect the native biodiversity by changing community composition, biotic interactions and other ecosystem processes (Vilà et al. 2011; Pyšek et al. 2012; Vilà and Hulme 2017), as well as by replacing it with common and widespread *taxa* (McKinney and Lockwood 1999; Hahs and McDonnell 2016), and can promote alternative successional trajectories that may dramatically affect the landscape (Williamson 1996; McKinney and Lockwood 1999; Weber 2003; Acosta et al. 2007; Del Vecchio et al. 2013; Gaertner et al. 2014; Stinca et al. 2015; Malavasi et al. 2018).

An alien organism needs to overcome geographical, environmental, and reproductive barriers to colonize a new region and spread over wide areas. In this process, some factors and traits can be more significant than others in explaining its success and therefore its invasiveness (Van Kleunen et al. 2015). Particularly, the concept of invasiveness of plant species has been integrated with that of propagule pressure and of "invasibility" of habitats and plant communities, i.e. the susceptibility of an environment to invasions by alien species, as different habitats and phytocoenoses may be more invasible than others and show different degrees of resistance/resilience (Rejmánek 1989; Lonsdale 1999; Rejmánek et al. 2005; Richardson and Pyšek 2006). All these concepts have been included in

the unifying theory of invasion syndromes (Perkins and Nowak 2013).

Patterns of distribution and abundance of IAS depend on a number of drivers including introduction history and pathways, life traits, availability of potentially invasible ecosystems, residence time, disturbance (Wilson et al. 2007; Carranza et al. 2010; Richardson et al. 2011; Comin et al. 2011; Dainese and Poldini 2012; Jucker et al. 2013). It is acknowledged that anthropogenic drivers play a crucial role in the establishment and spread of alien species (Pyšek and Richardson 2006; Pyšek et al. 2010a, 2010b; Bolpagni and Piotti 2015; Lazzaro et al. 2017; Stinca et al. 2017), however alien species showed to be largely constrained also by the same broad environmental factors acting on the native vegetation (Rouget et al. 2015). The same applies to IAS populations as well, as it has been demonstrated that what is good for natives is good for aliens too (Pyšek and Richardson 2006; Dalle Fratte et al. 2019).

Vascular plants are the most investigated taxonomic group in the field of invasive biology, and Europe devoted great efforts to their study, being the second continent (after North America) for investigative endeavour on plant invasions (Pyšek et al. 2009; Early et al. 2016). However, in spite of a huge scientific production about alien species numbers, ecology, impacts and distribution (for a synthesis at the European level, see DAISIE 2009 and Galasso et al. 2018 for Italy), their patterns and co-occurrence dynamics have only recently started to be in the spotlight, together with the factors driving alien plant assemblages (Hui et al. 2013; Pyšek and Chytrý 2014; Rouget et al. 2015).

A number of studies addressed the key role of cover and dominance of alien species to understand the invasion patterns (Lundholm and Larson 2004; Smith et al. 2004; Crall et al. 2006; Chytrý et al. 2008, 2009; Pyšek et al. 2010a, 2010b). This seems to some extent more important than alien species number, suggesting the hypothesis that the more an alien *taxon* becomes dominant in a vegetation type, the stronger the impact on native species diversity might be. This approach gave a pulse to research on alien plant species assemblages, and recently investigations on IAS have been addressed also at the community scale and benefitted from the currently available large databases of vegetation plots (see, e.g., Dengler et al. 2011; Landucci et al. 2012; Del Vecchio et al. 2015; Chytrý et al. 2016; Sperandii et al. 2018; Bonari et al. 2019).

A study by Chytrý et al. (2008) demonstrated that the habitat type is a reliable predictor of the level of plant invasion. This paper took into account patterns of plant invasions across habitats at the European scale, suggesting precious insights for biodiversity conservation and habitat monitoring, especially when considering the acknowledged correspondence between syntaxonomic types and habitats *sensu* Dir. 92/43/EEC (European Commission 1992, 2013; Evans 2010; Biondi et al. 2012; Viciani et al. 2016). Indeed, alien species have been listed among the causes of habitat decline and loss also in the most recent

red-list assessments, both at European (Janssen et al. 2016) and Italian scale (Gigante et al. 2018).

Some authors started to qualify and quantify the role of alien species in different vegetation and habitat types, and to point out the functional role that alien species play in plant communities (Celesti-Grapow et al. 2010; Pyšek and Chytrý 2014; Prisco et al. 2016). Moreover, habitat misclassification can be favoured by the presence of alien species (Sarmati et al. 2019). However, to date only few studies focussed on alien-dominated plant communities and the role of aliens in natural assemblages. With notable exceptions (e.g. Jurko 1964; Hadač and Sofron 1980; Ubaldi 2003; Vítková and Kolbek 2010; Sîrbu and Oprea 2011; Allegrezza et al. 2019), the large majority of vegetation studies, especially outside Europe, are still mostly focused on natural and semi-natural phytocoenoses with few or no aliens (Chytrý et al. 2009; Pyšek and Chytrý 2014).

Despite a long-dated Italian tradition of phytosociological studies, a national synthesis of the alien-dominated (and co-dominated) plant communities thriving in Italy has never been produced yet. Therefore, the present research aims at taking the first step toward the filling of this knowledge gap.

The European and national projects dedicated to IAS, which involved and currently still involve research academic centres and institutions, are numerous in Italy. Some of them have helped to gather a significant amount of information on invasive plant and animal species. Among the most recent, addressing non-native plant species, their impact and their management, as well as the awareness of the general public on these topics, we can mention: LIFE ASAP (<http://www.lifeasap.eu>), LIFE GESTIRE IP2020 (<http://www.naturachevale.it/il-progetto/life-gestire-2020/>), LIFE REDUNE (<http://www.liferedune.it>), MARITTIMO ALIEM (<http://interreg-maritime.eu/web/aliem>).

The present work is the result of a research agreement between SISV (the Italian Society for Vegetation Science) and ISPRA (the Italian Institute for Environmental Protection and Research), with financial support from the Italian Ministry of Environment, aimed at supporting the implementation of Regulation EU 2014/1143 (updated by EU Reg. 2017/1263) on the prevention and management of the introduction and spread of invasive alien species. In particular, the general agreement focused on: i) the update of the Database of Italian Alien Species (DIAS) with reference to their impacts on the ecosystems and the most threatened habitats and ii) the identification of the alien-dominated or co-dominated plant communities occurring in Italy. The outcomes of the latter are here presented. The research on alien plant communities and habitats was carried out through the collaboration of a wide working group of experts led by a Coordinating Committee composed by SISV members.

Methods

A dedicated SISV working team, formed by national and local experts, collected all the thematic literature related to terrestrial and freshwater alien-dominated and co-dominated vegetation. On the basis of this bibliographic dataset, a selection of all vegetation data was carried out, with special attention to nomenclature and syntaxonomic classification. All the existing national and regional vegetation databases (e.g. LiSy – <http://www.scienzadellavegetazione.it/sisv/lisy/index.jsp>; Poldini et al. 1985; Poldini 1991, 2002, 2009; Gallizia Vuerich et al. 1999; Brullo et al. 2001; Gigante et al. 2012; Landucci et al. 2012; Evangelista et al. 2016), together with the regional bibliographic sources (e.g. Poldini 1989; Poldini and Vidali 1989; Poldini et al. 1991, 1999), were consulted. Data concerning each phytocoenosis were selected if one or more alien species played a substantial role in the analysed vegetation unit. In particular, the SISV Coordinating Committee collected and checked the information and selected the data whenever:

1. the analysed low rank *syntaxon* (association, subassociation, phytocoenon) was dominated or co-dominated by one or more alien plant species; a cover value ≥ 3 according to the "Braun-Blanquet" scale (Braun-Blanquet 1979) for alien species cover in each relevé has been set as threshold; if, instead of the "Braun-Blanquet" scale, the "Pignatti" scale was used in the bibliographical reference (Pignatti and Mengarda 1962), the cover value threshold was set to ≥ 2 , i.e. 20-40%;
2. the alien species name was included in the name of the *syntaxon*, by that implying that it is a characteristic/differential/diagnostic or somehow important *taxon* for the definition of the *syntaxon*, or even dominating and determining its vertical structure (as stated in Art. 10b of the International Code of Phytosociological Nomenclature: Theurillat et al. 2020).

Starting from this selected dataset, a draft syntaxonomic list was produced. The names of the plant communities and their syntaxonomic attributions at higher ranks have been acknowledged (and are here reported) in the same form as published by the original Authors. In case of inconsistencies or discrepancies, a specific comment has been provided.

The syntaxonomic scheme has then been updated in accordance with the Italian Vegetation Prodrome (Biondi et al. 2014; <http://www.prodromo-vegetazione-italia.org/>), which however does not take into considerations *syntaxa* below the alliance level. Relevant differences and discrepancies with other syntaxonomic frames, especially regarding the EuroVegChecklist (Mucina et al. 2016), have been commented.

Plant nomenclature in the text follows the Checklists of the vascular flora of Italy (Bartolucci et al. 2018; Galasso et al. 2018) and later updates reported in the "Portal to the flora of Italy" (<http://dryades.units.it/floritaly/index.php>), to which we referred also to identify the species to be considered as aliens to Italy, including archaeophytes (e.g. *Arundo donax*, *Acanthus mollis* subsp. *mollis*) as well. New hybridogenic species due to xeno-speciation events (e.g. some species of *Oenothera* or *Vitis*) were taken into account, when considered as alien *taxa* by Galasso et al. (2018). In case of species considered alien to an administrative region and native to another one, only the communities reported for the region where the species is alien have been recorded (e.g.: a community dominated by *Acanthus mollis* subsp. *mollis* was considered alien to Liguria but not to Sicily, where this species is considered as native, see Minissale et al. 2019).

Results and discussion

A comprehensive and annotated checklist of the alien-dominated and co-dominated plant communities occurring in Italy is provided in Appendix I. All phytocoenoses have been framed in an updated syntaxonomic scheme. A specific bibliographical list with references for all the *syntaxa* quoted in the checklist is available in Appendix II.

The checklist includes a total of 27 classes of vascular plant vegetation and one of bryophyte vegetation, including plant communities dominated or co-dominated by aliens to Italy. The total number of low rank *syntaxa* (associations/subassociations/communities) amounts to 194.

The number of communities for each class is reported in Fig. 1. As expected, the class with the highest number of alien-dominated plant communities resulted by far *Stellarietea mediae*, followed by *Artemisietae vulgaris*. The presence of some higher-rank *syntaxa* named after and mainly formed by IAS (e.g. the class *Robinietea*, the order *Nicotiano glaucae-Ricinetalia communis*) is worth to be noted, which highlights the coenological and physiognomic-structural autonomy of these communities. The 27 identified classes and some subordinate *syntaxa* have been grouped in clusters based on their physiognomy and ecology. The considered groups are as follows:

- Forest vegetation (*Querco-Fagetea*, *Quercetea ilicis*);
- Anthropogenic woody vegetation (*Robinietea* and part of *Rhamno-Prunetea*);
- Alluvial, marshy and riparian woody vegetation (*Alnetea*, *Salici-Populeta*, *Salicetea*, *Alnion incanae*);
- Perennial herbaceous hygrophilous and hygro-nitrophilous vegetation (*Galio-Urticetea*, *Filipendulo-Convolvuleta*, *Molinio-Arrhenatheretea*, *Phragmito-Magnocaricetea*);

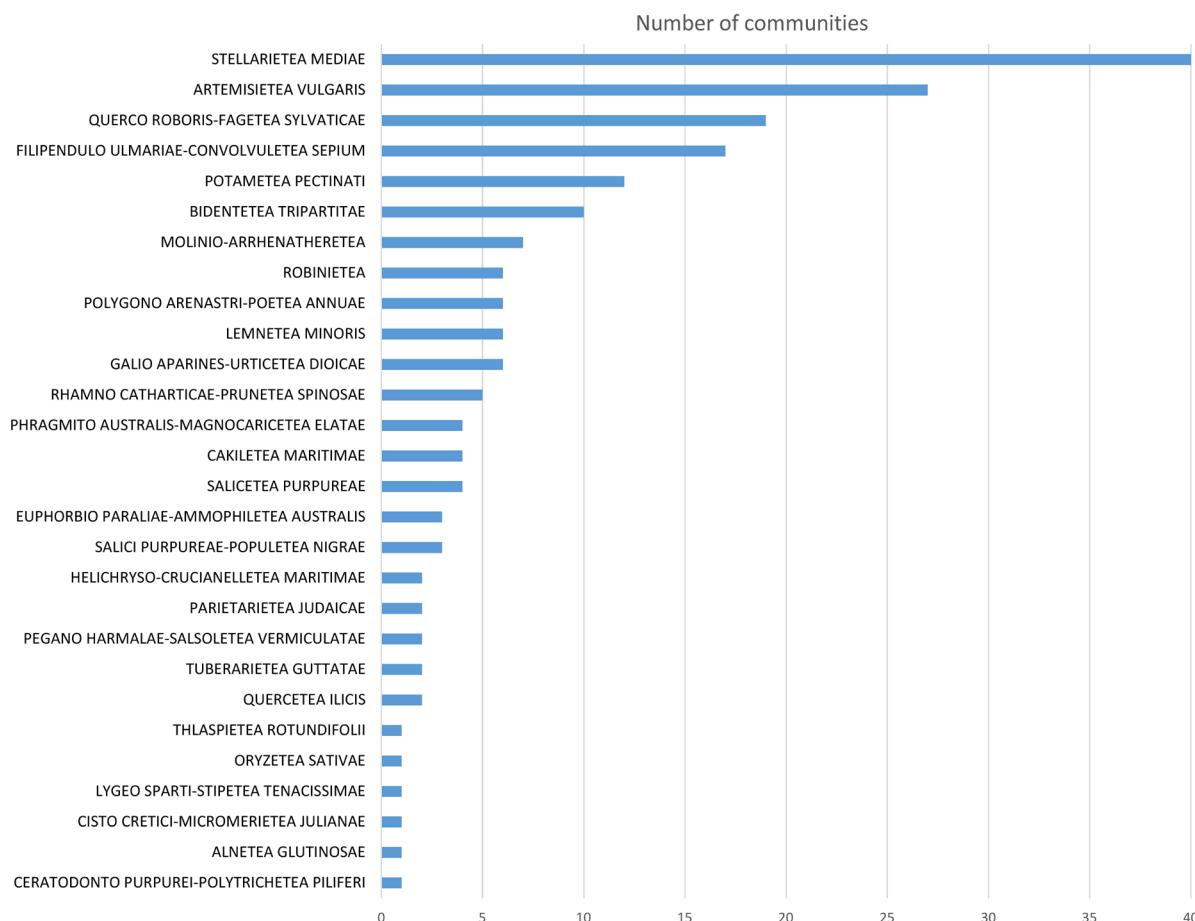


Figure 1. Number of alien-dominated communities for each syntaxonomic class of the checklist reported in Appendix I.

- Perennial ruderal herbaceous vegetation (*Artemisieta*);
- Annual ruderal herbaceous vegetation (*Stellarieta*, *Polygo no arenastri-Poetea annuae*);
- Annual herbaceous hygro-nitrophilous vegetation (*Bidenteta*);
- Hydrophilic freshwater vegetation (*Potameta*, *Lemnetea*);
- Psammophilous vegetation (*Euphorbio-Ammophileta*, *Cakileta*, *Helichryso-Crucianelleta*).

Results of the community rates per class groups are shown in Fig. 2. The group including the classes of annual ruderal herbaceous vegetation (*Stellarieta mediae* and *Polygo no arenastri-Poetea annuae*) hosts about 25% of the total number of the detected alien-dominated communities. *Stellarieta mediae* is by definition characterized by high rates of alien plant species, especially archaeophytes (<http://www.prodromo-vegetazione-italia.org/>). However, data analysis showed that this explains only part of the story. Actually, the involved alien species and genera are mainly represented by neophytes (e.g. *Robinia pseudoacacia*, *Artemisia* sp.pl., *Amaranthus* sp.pl., *Erigeron* sp.pl., *Euphorbia* sp.pl., *Solidago* sp.pl.). Indeed, it is known that neophytes mostly occur in strongly anthropogenic areas, whose habitats appear to be not only the most invaded, but also the most invasible (Pyšek et al. 2002,

2005; Deutschewitz et al. 2003; Kühn et al. 2003; Chytrý et al. 2008). On the contrary, relatively low- or non-anthropized ecosystems, such as nutrient-poor environments or montane habitats, are least or not invaded (Chytrý et al. 2008; Angiolini et al. 2019). For these reasons, heavily human-impacted environments (i.e. arable lands and fallow fields, urban and industrial areas, aquatic and riparian habitats) show the highest levels of neophyte invasion, as already suggested by previous studies, at least in continental areas (Kowarik 1995; Walter et al. 2005; Richardson and Pyšek 2006; Chytrý et al. 2009; Myśliwy 2014) and, as a consequence, the highest number of alien-dominated plant communities (Bolpagni and Piotti 2015).

Also the coastal areas are highly impacted by IAS (Acosta et al. 2007; Carboni et al. 2010; Del Vecchio et al. 2013, 2015; Lazzaro et al. 2017). Indeed, the close connection between invasibility, propagule pressure and habitat disturbance is a widely accepted relationship (Di Castri 1990; Vitousek et al. 1997; Pino et al. 2006; Perkins and Nowak 2013). This is indirectly confirmed in this study, by the fact that many of the less represented groups of alien-dominated communities refer to scarcely invasible environments, such as scree (*Thlaspietea rodundifolii*) or Mediterranean grass-dominated vegetation (*Lygeo-Stipetea*). Unexpectedly, our data seem to suggest that psammophilous coastal vegetation experiences low rates of

alien community occurrence (Figs. 1, 2). However, to correctly interpret this outcome, it should be considered that the collected data might outline a biased picture due to a lack of syntaxonomic investigation and classification for some communities. In support of this hypothesis, it should be noted that many alien coastal communities dominated by *Yucca* sp.pl., *Agave* sp.pl., *Opuntia* sp.pl. or *Acacia* sp.pl. have been noted although not syntaxonomically described yet (Carboni et al. 2010; Santoro et al. 2012; Del Vecchio et al. 2013; Lazzaro et al. 2014, 2015, 2017; Stinca et al. 2017). Moreover, low numbers of alien-dominated communities described for a particular environment do not imply low rates of invasion, as even few IAS can spread dramatically to the detriment of native plant communities. For instance, the neo-phytocoenoses formed by just two species of *Carpobrotus* are nowadays massively distributed along the coasts of the whole Europe (Souza-Alonso et al. 2019), possibly leading to a peculiar coenological diversity which, however, has never been formalized in syntaxonomic terms yet.

Disturbance regimes are often related to fluctuation in resource availability, indicated as a key driver for invasions, in relation to the intermittent resource enrichment or release (Alpert et al. 2000; Davis et al. 2000; Richardson and Pyšek 2006). For this reason, besides strongly anthropized habitats, also aquatic and riparian ecosystems are particularly susceptible and extremely vulnerable to biological invasions and generally show high rates of alien species, due to their intrinsically medium-high levels of disturbance (Di Castri 1990; Rauchich and Reader 1999; Pyšek and Prach 1993; Alpert et al. 2000; Bolpagni 2013; Bolpagni and Piotti 2015). As it is known, alien hydrophytes often become dominant in these environments, and give rise to species-poor alien communities. This is confirmed by our results (Figs. 1, 2) where, despite the relative scarcity of ecological niches in aquatic environments, the percentage of alien hydrophytes communities is relevant (Lazzaro et al. 2019). This can have severe consequences considering that alien aquatic plants, as primary producers and often structural component of the ecosystems, can drastically transform the structure of freshwater habitats and their water quality (Valley and Bremigan 2002; Rommens et al. 2003; Perna and Burrows 2005; Ricciardi and MacIsaac 2011; Brundu et al. 2012; Brundu 2015; Ceschin et al. 2016, 2019).

Other habitats also linked to water (and to the related high levels of disturbance) show a high number of alien-dominated communities. It is the case of riparian non-woody habitats, or of annual and perennial herbaceous hygrophilous and hygro-nitrophilous vegetation types (Fig. 2), which reach high percentages (the latter more than 18%).

Also the alien communities attributed by many authors to *Querco-Fagetea* have a prevalent mesohygrophilous character (Appendix I). The most relevant woody invasive alien species, also found in the anthropogenic neophytic woody vegetation (e.g. *Robinia pseudoacacia*, *Ailanthus altissima*, *Amorpha fruticosa*, and many others)

can be defined as “ecosystem engineers”, i.e. species that shape habitats and/or cause changes to their state and resources availability (Vitousek 1986; Schmitz et al. 1997; Jones et al. 1997; Crooks 2002; Bañou 2009; Bañou and Vilá 2009; Djurdjevic et al. 2011; Benesperi et al. 2012; Cierjacks et al. 2013; Vítková et al. 2017; Lazzaro et al. 2018). The replacement of native species by alien plants, even when apparently ecologically equivalent, almost always negatively affects the ecosystems, especially if those species act as “ecosystem engineers” (Brown et al. 2006; Wilson and Ricciardi 2009; Lazzaro et al. 2018; Sitzia et al. 2018; Uboni et al. 2019).

In the checklist we also recorded a bryophyte community dominated by the alien *Campylopus introflexus*, which in Europe is considered a neophytic moss, introduced from the Southern hemisphere and rapidly expanding (Hill et al. 2006). It was detected along Mediterranean ponds and neighbouring wood glades (Cogoni et al. 2002; Puglisi et al. 2016; Poponessi et al. 2016, 2018) and its distribution deserves to be monitored.

Few considerations can be made on the number of alien species involved in the communities reported in the checklist. According to Galasso et al. (2018), there are 791 non-native naturalized species in the Italian territory, 221 of which are considered invasive in at least one region. Our checklist shows that, based on the current knowledge, less than one hundred of these species (precisely 88) perform a dominant or co-dominant role in the considered plant communities. Only five of these (*Ailanthus altissima*, *Elodeia nuttallii*, *Lagarosiphon major*, *Myriophyllum aquaticum*, *Cenchrus setaceus*) are considered IAS of Union Concern (see https://ec.europa.eu/environment/nature/invasivealien/list/index_en.htm), however this number grows considerably when downscaling at the national level, with 76 of them (i.e. more than 86% of the total) to be considered as IAS in Italy (Galasso et al. 2018). Again, this can be traced back to the fact that most of these IAS are neophytes, many have been introduced in relatively recent times, and probably there was not enough time to give rise to such a degree of invasion to be relevant at European scale yet. Additionally, also at the national scale these invasions are often very localized or at very early stages, and their study from the phytosociological point of view is still a minor topic (although emergent).

At the same time, the continuous rate of introduction-naturalisation-invasion of new alien plants is an ongoing process that should be detected just in early stages, in order to prevent serious damage to native biodiversity. The numbers here reported raise the alarm for planning conservation biodiversity studies, monitoring protocols and management activities.

Conclusion

The here presented first checklist of the alien-dominated plant communities in Italy should not be considered exhaustive. It is the first step toward a better understanding

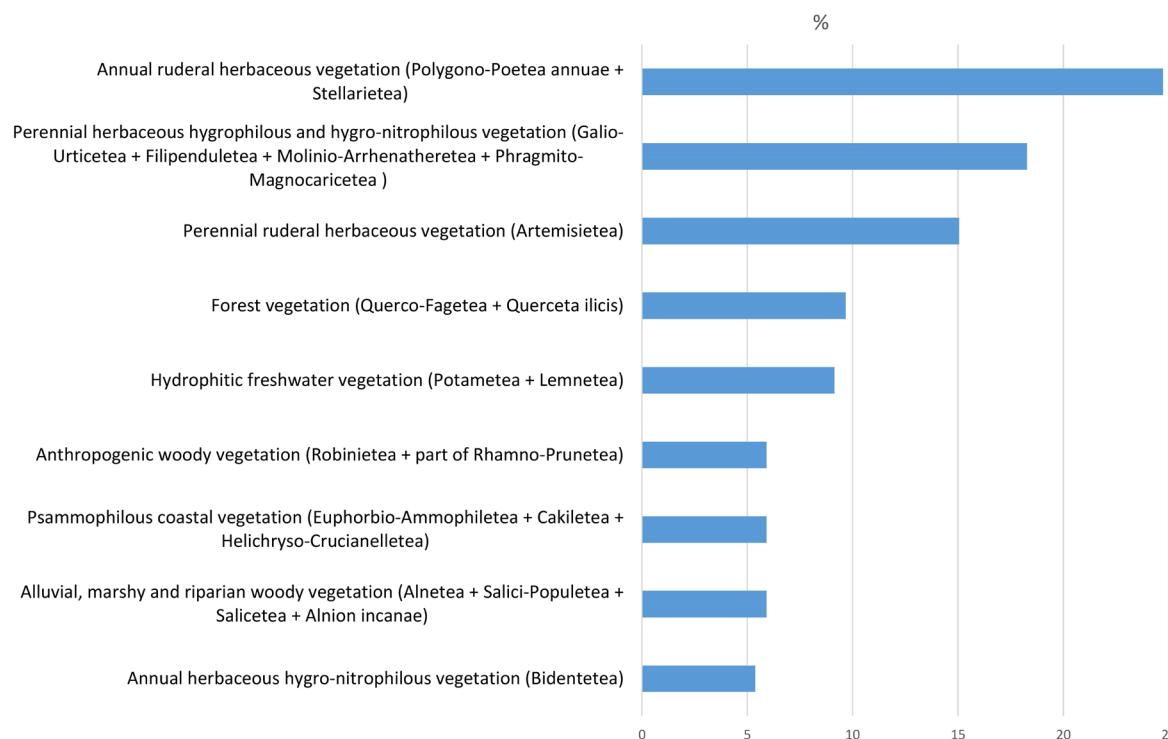


Figure 2. Percentages of alien-dominated communities in physiognomically and ecologically homogeneous groups of classes and subordinate *syntaxa*, based on the checklist reported in Appendix I.

of distribution, ecology and invasion processes of alien species at community level in this country. Our review represents a screenshot of the current knowledge and suggests that a serious lack of investigation for certain vegetation types (and probably for certain areas of the country) has to be highlighted.

The present checklist of the alien vegetation in Italy can represent a very useful tool, not only for stimulating further studies and investigations but also for prevention, management and monitoring purposes. As emphasized by Olaczek (1982), the phytosociological school, taking into account the whole floristic composition of plant communities, was one of the first approaches able to detect the effects of alien species on the diversity of natural phytocoenoses and to include these new communities in the syntaxonomical vegetation classification. In fact, the “floristic and phylogenetic homogenization” (e.g. Pino et al. 2009) and the “degeneration of phytocoenoses” by means of a progressive modification of structure and floristic composition due to the alien species invasion, concepts expressed by Olaczek (1982) and Faliński (1998a, 1998b), can transform a native plant community into an anthropogenic one, or even in a “novel ecosystem” (Lugo 2015), susceptible to be classified in a new syntaxonomic frame.

This becomes particularly important when considering the close link between plant communities syntaxonomically described and Natura 2000 habitat types, as listed in the Annex I to the Directive 92/43/EEC (European Commission 1992, 2013). It is mandatory for Member States to conserve Annex I habitats in Europe in a favourable

conservation status (Evans 2012; Gigante et al. 2016). Undoubtedly, a better understanding of the processes by which a plant community and a habitat type are firstly invaded and then dominated by alien species, together with the comprehension of the successional (and syntaxonomic) implications of those processes, can effectively support the monitoring and management of biodiversity and protected areas.

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Appendices

Appendix I – Alien-dominated and co-dominated plant communities in Italy (*Syntaxa* authors are abbreviated according to Izco 2002)

Class	Order	Alliance	Association/Phytocoenon	Reference number in the specific bibliographical list of Supplement 1
2 LEMNETEA MINORIS O. Bolòs & Masclans 1955				
		2.1 LEMNETALIA MINORIS O. Bolòs & Masclans 1955		
			<i>Lemna minuta</i> community	115
		2.1.1 Lemnion minoris O. Bolòs & Masclans 1955		
			<i>Azollo filiculoidis</i> - <i>Lemnetum minuscolaef</i> Felzines & Loiseau 1991	82, 83, 90
			<i>Ceratophyllo-Azolletum filiculoidis</i> Nedelcu 1967	56, 58
			<i>Lemna minuta</i> community	110
			<i>Lemnetum minuto-gibbae</i> Liberman Cruz, Pedrotti & Venanzoni 1988	56
			<i>Lemno-Azolletum filiculoidis</i> Br.-Bl. 1952	40, 115
3 POTAMETEA PECTINATI Klika in Klika & V. Novák 1941				
		3.1 POTAMETALIA PECTINATI Koch 1926		
		3.1.1 Potamion pectinati (Koch 1926) Libbert 1931		
			<i>Callitricho-Elodeetum canadensis</i> Passarge 1964 ex Passarge 1994	39
			<i>Elodea canadensis</i> and <i>Potamogeton crispus</i> community	91
			<i>Elodea nuttallii</i> community	28
			<i>Elodeo-Potametum crispi</i> (Pignatti 1953) Passarge 1994	40
			<i>Elodeo-Ranunculetum</i> Richard 1975	114
			<i>Lagarosiphon major</i> community	28
			<i>Myriophyllum aquaticum</i> community	64
			<i>Potametum crispi</i> Soó 1927 <i>Myriophyllum aquaticum</i> variant	64
			<i>Potametum lucentis</i> Hueck 1931 <i>Lagarosiphon major</i> variant	116
		3.1.2 Nymphaeion albae Oberdorfer 1957		
			<i>Hydrocotyletum ranunculoidis</i> Corbetta & Lorenzoni 1976	42

Class	Order	Alliance	Association/Phytocoenon	Reference number in the specific bibliographical list of Supplement 1
3.1.3		Ranunculion aquatilis Passarge 1964		
		<i>Callitrichete stagnalis</i> and <i>Myriophyllum aquaticum</i> community	64	
		<i>Lemno-Callitrichetum cophocaruae</i> (Mierwald 1988) Passarge 1992	64	
		<i>Myriophyllum aquaticum</i> variant		
12		BIDENTETEA TRIPARTITAE Tüxen, Lohmeyer & Preising ex Von Rochow 1951		
12.1		<i>BIDENTETALIA TRIPARTITAE</i> Br.-Bl. & Tüxen ex Klika in Klika & Hadac 1944		
12.1.1		Bidention tripartitae Nordhagen 1940		
		<i>Bidens frondosus</i> community	57	
		<i>Bidenti-Polygonetum mitis</i> (Von Rochow 1951) Tüxen 1979	126	
		<i>Bidens frondosa</i> variant		
		<i>Bidenti-Polygonetum mitis</i> (Von Rochow 1951) Tüxen 1979	15	
		<i>echinochloetosum crus-galli</i> Baldoni & Biondi 1993		
		<i>Polygonetum hydropiperis</i> Passarge 1965 <i>Bidens frondosus</i> facies	61	
		<i>Xanthio italicii-Polygonetum persicariae</i> O. Bolòs 1957	61, 60	
		<i>Xanthio italicii-Persicarietum maculosae</i> O. Bolòs 1957 nom. mut. propos.	59	
		<i>Abutilon theophrasti</i> variant		
		<i>Xanthium orientale</i> subsp. <i>italicum</i> community	46	
12.1.2		Chenopodion rubri (Tüxen 1960) Hilbig & Jage 1972		
		<i>Cyperetum esculenti</i> Wisskirchen 1995	62	
			1, 7, 13, 20, 29, 38,	
		<i>Polygono-Xanthietum italicii</i> Pirola & Rossetti 1974	43, 44, 46, 53, 55,	
			63, 68, 70, 75, 76,	
			81, 92, 94, 95	
		<i>Polygono-Xanthietum italicii</i> Pirola & Rossetti 1974	4	
		<i>Ambrosia artemisiifolia</i> variant		
16		PHRAGMITO AUSTRALIS-MAGNOCARICETEA ELATAE Klika in Klika & V. Novák 1941		
16.1		<i>PHRAGMITETALIA AUSTRALIS</i> Koch 1926		
16.1.1		Phragmition communis Koch 1926		
		<i>Scirpetum maritimi</i> (Christiansen 1934) Tüxen 1937	60	
		<i>Paspalum distichum</i> variant		
16.3		<i>MAGNOCARICETALIA ELATAE</i> Pignatti 1953		
16.3.1		Magnocaricion elatae Koch 1926		
		<i>Cyperus eragrostis</i> community	16	
		<i>Cyperus glomeratus</i> community	43	
16.5		<i>NASTURTIO OFFICINALIS-GLYCERIETALIA FLUITANTIS</i> Pignatti 1953		
16.5.1		Glycerio fluitantis-Sparganion neglecti Br.-Bl. & Sissingh in Boer 1942		
		<i>Eleocharitetum palustris</i> Schennikov 1919	25	
		<i>paspalotosum paspaloidis</i> Biondi et al. 2002		
20		EUPHORBIO PARALIAE-AMMOPHILETEA AUSTRALIS Géhu & Rivas-Martínez in Rivas-Martínez, Asensi, Díaz-Garretas, Molero, Valle, Cano, Costa & T.E. Díaz 2011		
20.1		<i>AMMOPHILETALIA AUSTRALIS</i> Br.-Bl. 1933		
20.1.1		Ammophilion australis Br.-Bl. 1933 em. Géhu & Géhu-Franck 1988		
		<i>Carpobrotus acinaciformis</i> community	88	

Class	Order	Alliance	Association/Phytocoenon	Reference number in the specific bibliographical list of Supplement 1
			<i>Carpobrotus edulis</i> community	88
			<i>Xanthietum-Ammophiletum</i> Pignatti 1953	91
21	CAKILETEA MARITIMAE	Tüxen & Preising ex Br.-Bl. & Tüxen 1952		
21.1	EUPHORBIETALIA PEPLIS	Tüxen 1950		
21.1.1	Euphorbion peplis	Tüxen 1950		
			<i>Cakilo-Xanthietum italicici</i> Pignatti 1953	1, 9, 12, 38, 91, 93
			<i>Salsolo kali-Cakiletum maritimae</i> Costa et Mansanet 1981 corr. Rivas-Martínez et al. 1992 <i>xanthietosum</i> (Pignatti 1953) Géhu & Scoppola 1984	17, 24, 48, 70, 91, 106
			<i>Xanthio italicici-Cenchretum incerti</i> Biondi, Brugia Paglia, Allegrezza & Ballelli 1992	24, 100, 109
			<i>Xanthio italicici-Cenchretum longispini</i> Poldini et al. 1999	100, 106
22	HELICHRYSO-CRUCIANELLETEA MARITIMAE	(Sissingh 1974) Géhu, Rivas-Martínez & Tüxen 1973 em. Sissingh 1974		
22.1	HELICHRYSO-CRUCIANELLETALIA MARITIMAE	Géhu, Rivas-Martínez & Tüxen 1973 em. Sissingh 1974		
22.1.1	Crucianellion maritimae	Rivas Goday & Rivas-Martínez 1958		
			<i>Crucianello-Helichrysetum microphylli</i> Bartolo, Brullo, De Marco, Dinelli, Signorello & Spampinato 1992 <i>Carpobrotus acinaciformis</i> variant	125
			<i>Ephedro-Helichrysetum microphylli</i> Valsecchi & Bagella 1991 <i>Carpobrotus acinaciformis</i> variant	125
31	PARIETARIETEA JUDAICAE	Oberdorfer 1977		
31.1	TORTULO-CYMBALARIELTALIA	Segal 1969		
31.1.1	Parietario judaicæ	Segal 1969		
			<i>Cheiranthetum cheirii</i> Segal 1962	46
			<i>Erigeronetum karvinskianii</i> Oberdorfer 1969	46, 54, 70
33	THLASPIETEA ROTUNDIFOLII	Br.-Bl. 1948		
33.6	EPILOBIETALIA FLEISCHERI	Moor 1958		
			<i>Oenothera biennis</i> and <i>Scrophularia canina</i> community	114
34	ARTEMISIETEA VULGARIS	Lohmeyer, Preising & Tüxen ex Von Rochow 1951		
			<i>Sporobolus vaginiflorus</i> community	104
			<i>Senecio mikanioides</i> community	101
			<i>Helianthus tuberosus</i> community	99, 104
			<i>Solidago gigantea</i> community	70, 120
			<i>Senecio inaequidens</i> community	99
34.1	ARCTIO LAPPAE-ARTEMISIETALIA VULGARIS	Dengler 2002		
34.1.1	Arction lappae	Tüxen 1937		
			<i>Artemisietum verlotorii</i> Lang 1973	41, 99, 101
			<i>Arundo donax</i> community	43
			<i>Saponario-Artemisietum verlotorum</i> Biondi & Baldoni 1993	15, 63, 72
			<i>Sileno albae-Acanthetum mollis</i> Biondi, Allegrezza & Filigheddu 1990	21

Class	Order	Alliance	Association/Phytocoenon	Reference number in the specific bibliographical list of Supplement 1
			34.2 AGROPYRETALIA INTERMEDII-REPENTIS Oberdorfer, Müller & Görs in Müller & Görs 1969	
			34.2.1 Convolvulo arvensis-Agropyron repens Görs 1966	
			<i>Artemisia verlotiorum</i> community	13, 127
			<i>Sorghum halepense</i> community	46
			34.4 BRACHYPODIO RAMOSI-DACTYLETALIA HISPANICAE Biondi, Filigheddu & Farris 2001	
			34.4.4 Bromo-Oryzopsis miliaceae O. Bolòs 1970	
			<i>Boerhaavia-Oryzopsis miliaceae</i> Brullo 1984	33
			34.5 ONOPORDETALIA ACANTHII Br.-Bl. & Tüxen ex Klika in Klika & Hadač 1944	
			<i>Reynoutria japonica</i> community	118
			<i>Artemisia verlotiorum</i> community	104
			<i>Helianthus tuberosus</i> community	29
			<i>Senecio inaequidens</i> community	29
			<i>Solidago gigantea</i> community	29
			34.5.1 Onopordion acanthii Br.-Bl. in Br.-Bl., Gajewski, Wraber & Walas 1936	
			<i>Erigeron canadense</i> and <i>Broussonetia papyrifera</i> community	91
			34.5.2 Dauco carotae-Melilotion albi Görs 1966	
			<i>Artemisia verlotiorum</i> community	47, 118
			<i>Artemisio absinthii-Senecionetum inaequidentis</i> Pirone 2001	45, 96
			<i>Echio-Melilotetum</i> Tüxen 1947	
			<i>Senecio inaequidens</i> and <i>Erigeron annuus</i> variant	30
			<i>Echio-Melilotetum</i> Tüxen 1947 <i>Oenothera biennis</i> (aggr.) variant	31
			<i>Erigeron annuus</i> community	78
			<i>Helianthus tuberosus</i> community	29
			<i>Oenothera biennis</i> community	123
			<i>Senecio inaequidens</i> community	29, 31, 129
			<i>Solidago gigantea</i> community	29
			34.6 CARTHAMETALIA LANATI Brullo in Brullo & Marcenò 1985	
			34.6.2 Onopordion illyrici Oberdorfer 1954	
			<i>Carduo pycnocephali-Nicotianetum glaucae</i> Biondi, Blasi, Brugipaglia, Fogu & Mossa 1994	23
36			36 ORYZETEA SATIVAE Miyawaki 1960	
			36.1 CYPERO DIFFORMIS-ECHINOCHLOETALIA ORYZOIDIS O. Bolòs & Masclans 1955	
			36.1.1 Oryzo sativae-Echinochloion oryzoidis O. Bolòs & Masclans 1955	
			<i>Heteranthera</i> sp.pl. community	39
37			37 PEGANO HARMALAE-SALSOLETEA VERMICULATAE Br.-Bl. & O. Bolòs 1958	
			37.2 NICOTIANO GLAUCAE-RICINETALIA COMMUNIS Rivas-Martínez, Fernández-González & Loidi 1999	
			37.2.1 Nicotiano glaucae-Ricinion communis Rivas-Martínez, Fernández-González & Loidi 1999	
			<i>Nicotianeo glaucae-Ricinetum communis</i> (Br.-Bl. & Maire 1924) de Foucault 1993	37, 87
			<i>Polycarpo-Nicotianetum glaucae</i> Sunding 1972	37

Class	Order	Alliance	Association/Phytocoenon	Reference number in the specific bibliographical list of Supplement 1
38	POLYGONO ARENSTRI-POETEA ANNUAE	Rivas-Martínez 1975 corr.	Rivas-Martínez, Báscones, T.E. Díaz, Fernández-González & Loidi 1991	
			<i>Eleusinetum indicae</i> (Slavnic 1951) Pignatti 1953	30
			<i>Euphorbio-Oxalidetum corniculatae</i> Lorenzoni 1964	30
38.2	SAGINO APETALAE-POLYCARPETALIA TETRAPHYLLI	de Foucault 2010		
	38.2.1	Polycarpion tetraphylli	Rivas-Martínez 1975	
			<i>Eleusine indica</i> community	91
			<i>Eleusinetum indicae</i> (Slavnic 1951) Pignatti 1953	46, 70, 99, 104
			<i>Euphorbiatum maculatae</i> Poldini 1989	70, 99, 101, 104
			<i>Euphorbio-Oxalidetum corniculatae</i> Lorenzoni 1964	38, 99, 101
39	STELLARIETEA MEDIAE	Tüxen, Lohmeyer & Preising ex Von Rochow 1951		
39a.3	SOLANO NIGRI-POLYGONETALIA CONVOLVULI	(Sissingh in Westhoff, Dijk, Passchier & Sissingh 1946) O. Bolòs 1962		
	39a.3.1	Digitario ischaemi-Setarion viridis	Sissingh in Westhoff, Dijk, Passchier & Sissingh 1946	
			<i>Amarantho-Chenopodietum albi</i> (Morariu 1943) Soó 1957	101
			<i>Amarantho-Digitarietum sanguinalis</i> Pignatti 1953	67, 71, 91, 101
			<i>Chenopodium album</i> and <i>Amaranthus retroflexus</i> community	99
			<i>Cynodonto-Sorghetum halepensis</i> (Laban 1974) Kojic 1979	38
			<i>Echinochloo-Setarietum pumilae</i> Felföldy 1942 corr. Mucina 1993	105
			<i>xanthietosum italicii</i> Poldini et al. 1998	
			<i>Oxalido-Chenopodietum polyspermi</i> (Br.-Bl. 1921) Sissingh (1942) 1946 (*)	66, 67, 91
			<i>Oxalido-Chenopodietum polyspermi</i> (Br.-Bl. 1921) Sissingh (1942) 1946	66
			<i>Galinsoga parviflora</i> (*) subassociation	
			<i>Panico sanguinalis-Polygonetum persicariae</i> Pignatti 1953	14
			<i>sorghetosum halepensis</i> Baldoni 1995 (*)	
			<i>Panico sanguinalis-Polygonetum persicariae</i> Pignatti 1953	67
			<i>Acalypha virginica</i> (*) facies	
			<i>Panico sanguinalis-Polygonetum persicariae</i> Pignatti 1953	66
			<i>Echinochloa oryzoides</i> (*) facies	
			<i>Panico sanguinalis-Polygonetum persicariae</i> Pignatti 1953	66
			<i>Panicum capillare</i> (*) subassociation	
			<i>Panico sanguinalis-Polygonetum persicariae</i> Pignatti 1953	51
			<i>Datura stramonium</i> and <i>Portulaca oleracea</i> (*) variant	
			<i>Panico sanguinalis-Polygonetum persicariae</i> Pignatti 1953	51
			<i>Bolboschoenus maritimus</i> and <i>Paspalum distichum</i> (*) variant	
			<i>Euphorbio-Galinsogetum ciliatae</i> Passarge 1981	79
			<i>Galeopsido tetrahit-Galinsogetum parviflorae</i> Poldini et al. 1998	105
			<i>Galinsogo-Portulacetum</i> Br.-Bl. 1949 ex Pedrotti 1959	78, 91
			<i>Setario-Echinochloetum colonum</i> A. & O. Bolòs ex O. Bolòs 1956	34, 38
			<i>Setario-Galinsogetum parviflorae</i> (Beck 1941) Tüxen 1950 em. Müller & Oberdorfer	70
			<i>Setario ambiguae-Cyperetum rotundi</i> Brullo, Scelsi & Spampinato 2001	38
39a.3.3	Diplotaxion erucoidis	Br.-Bl. in Br.-Bl., Gajewski, Wraber & Walas 1936 em.		
			Brullo & Marcenò 1980	
			<i>Amaranthus retroflexus</i> community	86
39a.3.5	Fumarion wirtgenii-agrariae	Brullo in Brullo & Marcenò 1985		
			<i>Oxalis pes-caprae</i> community	85, 87

Class	Order	Alliance	Association/Phytocoenon	Reference number in the specific bibliographical list of Supplement 1
39b.1			<i>CHENOPODIETALIA MURALIS</i> Br.-Bl. in Br.-Bl., Gajewski, Wraber & Walas 1936	
39b.1.1			<i>Chenopodium muralis</i> Br.-Bl. in Br.-Bl., Gajewski, Wraber & Walas 1936	
			<i>Amarantho blitoidis-Chenopodietum ambrosoidis</i> O. Bolòs 1967	46
			<i>Amarantho muricati-Chenopodietum ambrosoidis</i> O. Bolòs 1967	34
			<i>Amarantho-Chenopodietum ambrosoidis</i> O. Bolòs 1967	34, 38, 99
			<i>Conyzetum albidae-canadensis</i> Baldoni & Biondi 1993	8
			<i>Lolium multiflorum</i> variant	
			<i>Conyzetum albido-canadensis</i> Baldoni & Biondi 1993	46, 101
			<i>Conzyo canadensis-Oenotheretum biennis</i> Biondi, Brugia paglia, Allegrezza & Ballelli 1992	24
			<i>Xanthio italicico-Daturetum stramoni</i> Fanelli 2002	46
			<i>Amaranthus deflexus</i> and <i>Polycarpon tetraphyllum</i> community	91
			<i>Datura stramonium</i> community	21
39b.2			<i>THERO-BROMETALIA</i> (Rivas Goday & Rivas-Martínez ex Esteve 1973) O. Bolòs 1975	
39b.2.1			<i>Echo plantaginei-Galactition tomentosae</i> O. Bolòs & Molinier 1969	
			<i>Erigeron canadensis</i> community	121
			<i>Galactito-Isatidetum canescens</i> Brullo 1983	32
39b.3			<i>SISYMBRIETALIA OFFICINALIS</i> J. Tüxen ex W. Matuszkiewicz 1962	
39b.3.1			<i>Sisymbrium officinalis</i> Tüxen, Lohmeyer & Preising ex Von Rochow 1951	
			<i>Artemisietum annuae</i> Fijalcowski 1967	30
			<i>Artemisietum annuae</i> Fijalcowski 1967 <i>ambrosietosum</i> Siniscalco & Montacchini 1989	118, 119
			<i>Conzyo-Lactucetum serriolae</i> Lohmeyer in Oberdorfer 1957	3, 31, 104
			<i>Datura stramonium</i> and <i>Malva neglecta</i> community	67, 91
			<i>Erigeron canadensis</i> community	70, 86
39b.3.2			<i>Hordeion leporini</i> Br.-Bl. in Br.-Bl., Gajewski, Wraber & Walas 1936 corr. O. Bolòs 1962	
			<i>Bassia scoparia</i> and <i>Chenopodium ambrosioides</i> community (**)	91
39b.4			<i>URTICO-SCROPHULARIETALIA PEREGRINAE</i> Brullo ex Biondi, Blasi, Casavecchia & Gasparri in Biondi, Allegrezza, Casavecchia, Galdenzi, Gasparri, Pesaresi, Vagge & Blasi 2014	
			<i>Oxalis pes-caprae</i> community	70
39b.4.1			<i>Veronic-Urticion urentis</i> Brullo in Brullo & Marcenò 1985	
			<i>Bromo-Brassicetum sylvestris</i> Brullo & Marcenò 1985	34
40			<i>GALIO APARINES-URTICETEA DIOICAE</i> Passarge ex Kopecký 1969	
			<i>Artemisia verlotiorum</i> community	30
40.1			<i>GALIO APARINES-ALLIARIETALIA PETIOLATAE</i> Oberdorfer ex Görs & Müller 1969	
			<i>Robinia pseudoacacia</i> community	39
40.1.1			<i>Petasition officinalis</i> Sill. 1933 em. Kopecký 1969	
			<i>Robinia pseudoacacia</i> community	5
40.1.2			<i>Geo-Alliarion</i> Lohmeyer & Oberdorfer ex Görs & Müller 1969	
			<i>Solidago canadensis</i> community	5, 89
40.1.5			<i>Allion triquetri</i> O. Bolòs 1967	
			<i>Acantho-Smyrnietum olusatris</i> Brullo & Marcenò 1985	34, 70
			<i>Acanthus mollis</i> community	70, 117

Class	Order	Alliance	Association/Phytocoenon	Reference number in the specific bibliographical list of Supplement 1
41	FILIPENDULO ULMARIAE-CONVOLVULETEA	SEPIUM	Géhu & Géhu-Franck 1987	
41.1	CALYSTEGIETALIA	SEPIUM	Tüxen ex Mucina 1993 nom. mut. propos. Rivas-Martínez, T.E. Díaz, Fernandez-Gonzales, Izco, Loidi, Lousá & Penas 2002	
		<i>Reynoutria japonica</i> community		31
		<i>Helianthus tuberosus</i> community		31
		<i>Solidago canadensis</i> and <i>Erigeron annuus</i> community		31
		<i>Erigeron annuus</i> community		31
41.1.1	Calystegion sepium	Tüxen ex Oberdorfer 1957 nom. mut. propos. Rivas-Martínez, T.E. Díaz, Fernandez-Gonzales, Izco, Loidi, Lousá & Penas 2002		
		<i>Amorpha fruticosa</i> community		81
		<i>Artemisia verlotiorum</i> community		127
		<i>Arundini-Convolvuletum sepium</i> (Tüxen & Oberdorfer) O. Bolòs 1962		72
		<i>Artemisia verlotiorum</i> variant		
		<i>Arundini-Convolvuletum sepium</i> (Tüxen & Oberdorfer 1958) O. Bolòs 1962		11, 15, 34, 40, 46, 80, 99, 101
		<i>Arundo donax</i> community		43
		<i>Calystegio sylvaticae-Arundinetum donacis</i> Brullo, Scelsi & Spampinato 2001		35, 38, 73, 84, 85, 87
		<i>Calystegio-Asteretum lanceolati</i> (Holzner et al. 1978) Passarge 1993		63
		<i>Helianthus tuberosus</i> community		15, 63, 71, 127
		<i>Humulus scandens</i> community		39
		<i>Rubus caesius</i> and <i>Amorpha fruticosa</i> community		89
		<i>Solidago canadensis</i> community		81, 83
		<i>Solidago gigantea</i> community		39, 63, 127
50	TUBERARIETEA GUTTATAE (Br.-Bl. in Br.-Bl., Roussine & Nègre 1952)	Rivas Goday & Rivas-Martínez 1963 nom. mut. propos. Rivas-Martínez, T.E. Díaz, Fernández-González, Izco, Loidi, Lousa & Penas 2002		
50.2	MALCOLMIETALIA	Rivas Goday 1958		
50.3.2	Laguro ovati-Vulpion fasciculatae	Géhu & Biondi 1994		
		<i>Ambrosio coronopifoliae-Lophochloetum pubescens</i> Biondi, Brugiapaglia, Allegrezza & Ballelli 1992		95, 99, 102
		<i>Sileno coloratae-Vulpietum membranaceae</i> (Pignatti 1953) Géhu & Scoppola 1984 <i>ambrosietosum coronopifoliae</i> Pirone 2005		97
55	LYGEO SPARTI-STIPETEA TENACISSIMAE	Rivas-Martínez 1978 nom. conserv. propos. Rivas-Martínez, T.E. Díaz, Fernández-González, Izco, Loidi, Lousa & Penas 2002		
55.2	HYPARRHENIETALIA HIRTAE	Rivas-Martínez 1978		
55.2.1	Hyparrhenion hirtae	Br.-Bl., P. Silva & Rozeira 1956		
		<i>Penniseto setacei-Hyparrhenietum hirtae</i> Gianguzzi, Ilardi & Raimondo 1996		36, 52
56	MOLINIO-ARRHENATHERETEA	Tüxen 1937		
56.4	HOLOSCHOENETALIA VULGARIS	Br.-Bl. ex Tchou 1948		
56.4.4	Paspalo distichi-Agrostion semiverticillatae	Br.-Bl. in Br.-Bl., Roussine & Nègre 1952		
		<i>Aster squamatus</i> and <i>Inula viscosa</i> community		94
		<i>Loto tenuis-Paspaletum paspaloidis</i> Biondi, Casavecchia & Radetic 2002		25, 68
		<i>Paspalo distichi-Polypogonetum viridis</i> Br.-Bl. in Br.-Bl., Gajewski, Wraber & Walas 1936 nom. mut. propos. Rivas-Martínez et al. 2002 (= <i>Paspalo distichi-Agrostidetum verticillati</i> Br.-Bl. in Br.-Bl., Roussine & Nègre 1952)		15, 41, 46, 60, 61, 63, 68, 70, 76, 93, 94, 95, 99, 101, 115

Class	Order	Alliance	Association/Phytocoenon	Reference number in the specific bibliographical list of Supplement 1
			<i>Paspalo distichi-Polypogonetum viridis</i> Br.-Bl. in Br.-Bl., Gajewski, Wraber & Walas 1936 nom. mut. propos. Rivas-Martínez et al. 2002 facies with <i>Cyperus eragrostis</i>	63
			<i>Paspalum distichum</i> (= <i>paspalooides</i>) community	19, 81, 103
56.5	PLANTAGINETALIA MAJORIS	Tüxen ex Von Rochow 1951		
	56.5.1	Lolio perennis-Plantaginion majoris Sissingh 1969		
		<i>Juncetum macri</i> (Diemont et al. 1940) Tüxen 1950	78	
		<i>Eleusine indica</i> community	91	
61	CISTO RETICULI-MICROMERIETEA JULIANAE	Oberdorfer ex Horvatić 1958		
	61.1	CISTO RETICULI-ERICETALIA MANIPULIFLORAE	Horvatić 1958	
	61.1.1	Cisto cretici-Ericion manipuliflorae Horvatić 1958		
		<i>Opuntia ficus-indica</i> community	69, 85	
64	RHAMNO CATHARTICAE-PRUNETEA SPINOSAE	Rivas Goday & Borja ex Tüxen 1962		
	64.1	PRUNETALIA SPINOSAE	Tüxen 1952	
		<i>Robinia pseudoacacia</i> and <i>Rubus ulmifolius</i> community	6	
		<i>Robinia pseudoacacia</i> and <i>Sambucus nigra</i> community	70	
	64.1.1	Berberidion vulgaris Br.-Bl. 1950		
		" <i>Corno sanguineae-Ligustretum vulgaris</i> " sensu Biondi et al. 1999 non Horvat 1956 <i>amorphosum fruticosae</i> Biondi, Vagge, Baldoni & Taffetani 1999	27	
	64.1.2	Cytision sessilifolii Biondi in Biondi, Allegrezza & Guitian 1988		
		<i>Cercido siliquastri-Rhoetum corariae</i> Biondi, Allegrezza & Guitian 1988	22	
	64.3.1	Pruno-Rubion ulmifolii O. Bolòs 1954		
		<i>Clematido vitalbae-Arundinetum donacis</i> Biondi & Allegrezza 2004	18	
65	ALNETEA GLUTINOSAE	Br.-Bl. & Tüxen ex Westhoff, Dijk & Passchier 1946		
	65.1	SALICETALIA AURITAE	Doing ex Westhoff in Westhoff & Den Held 1969	
	65.1.1	Salicion cinereae Müller & Görs 1958		
		<i>Salicetum cinereae</i> Zolyomi 1931 <i>Amorpha fruticosa</i> variant	65	
68	SALICI PURPUREAE-POPULETEA NIGRAE	Rivas-Martínez & Cantó ex Rivas-Martínez, Báscones, T.E. Diaz, Fernández-González & Loidi 2001		
	68.1	POPULETALIA ALBAE	Br.-Bl. ex Tchou 1948	
		<i>Robinia pseudoacacia</i> community	122	
	68.1.1	Populion albae Br.-Bl. ex Tchou 1948		
		<i>Acer negundo</i> community	46	
		<i>Ailanthes altissima</i> and <i>Robinia pseudoacacia</i> community	6	
69	SALICETEA PURPUREAE	Moor 1958		
	69.1	SALICETALIA PURPUREAE	Moor 1958	
	69.1.1	Salicion albae Soó 1930		
		<i>Amorpha fruticosae-Salicetum albae</i> Poldini, Vidali, Bracco, Assini & Villani in Poldini, Vidali & Ganis 2011	13, 27, 48, 71, 107	
		<i>Sicyos angulatus</i> community	40, 112	
	69.1.2	Salicion triandrae Müller & Görs 1958		
		<i>Salicetum triandrae</i> (Malcuit 1929) Noirlalise 1955 <i>Amorpha fruticosa</i> variant	27	

Class	Order	Alliance	Association/Phytocoenon	Reference number in the specific bibliographical list of Supplement 1
69.1.4		Salicion incanae Aichinger 1933		
		<i>Salicetum incano-purpureae</i> Sillinger 1933 Amorpha fruticosa variant		27, 62
70	QUERCETEA ILICIS Br.-Bl. in Br.-Bl., Roussine & Nègre 1952			49
		<i>Sequoia sempervirens</i> community		
70.1	QUERCETALIA ILICIS Br.-Bl. ex Molinier 1934			
		Oleo sylvestris-Ceratonion siliquae Br.-Bl. ex Guinochet & Drouineau 1944		
		<i>Asparago acutifolii-Oleetum sylvestris</i> Bacchetta et al. 2003		117
		<i>Opuntia ficus-indica</i> variant		
71	QUERCO ROBORIS-FAGETEA SYLVATICA Br.-Bl. & Vlieger in Vlieger 1937			
		<i>Acacia melanoxylon</i> community		49
		<i>Euonymus europaeus</i> and <i>Robinia pseudoacacia</i> community		77
		<i>Festuca heterophylla</i> and <i>Robinia pseudoacacia</i> community		77
		<i>Arrhenatherum elatius</i> variant		
		<i>Festuca heterophylla</i> and <i>Robinia pseudoacacia</i> community		77
		<i>Rubus macrophyllus</i> variant		
		<i>Pinus canariensis</i> community		49
		<i>Pinus radiata</i> community		49
		<i>Robinia pseudoacacia</i> community		50
		<i>Robinia pseudoacacia</i> and <i>Prunus serotina</i> community		113
		<i>Sambucus nigra</i> and <i>Robinia pseudoacacia</i> community		77
		<i>Chelidonium majus</i> variant		
		<i>Sambucus nigra</i> and <i>Robinia pseudoacacia</i> community		77
		<i>Poa trivialis</i> variant		
		<i>Sambucus nigra</i> and <i>Robinia pseudoacacia</i> community		77
		<i>Rubus gr. discolor</i> variant		
71.1	FAGETALIA SYLVATICA Pawłowski in Pawłowski, Sokołowski & Wallisch 1928			
		Tilio platyphilli-Acerion pseudoplatani Klika 1955		
		<i>Robinia pseudoacacia</i> community		50
	71.1.6	Carpinion betuli Issler 1931		
		<i>Prunus serotina</i> community		113
	71.1.10	Alnion incanae Pawłowski in Pawłowski, Sokołowski & Wallisch 1928		
		<i>Amorpha fruticosa</i> community		40
		<i>Populus nigra</i> and <i>Robinia pseudoacacia</i> community		26
		<i>Robinia pseudoacacia</i> and <i>Rubus ulmifolius</i> community		55
71.2	QUERCETALIA ROBORIS Tüxen 1931			
		<i>Buddleja davidi</i> community		74
	71.2.1	Quercion roboris Malcuit 1929		
		<i>Robinia pseudoacacia</i> community		50
71.3	QUERCETALIA PUBESCENTI-PETRAEAE Klika 1933			
		Crataego laevigatae-Quercion cerridis Arrigoni 1997		
		<i>Sambuco nigrae-Robinietum pseudacaciae</i> Arrigoni 1997		10
75	ROBINIETEA Jurko ex Hadac & Sofron 1980			
75.1	CHELIDONIO-ROBINIETALIA Jurko ex Hadac & Sofron 1980			

Class	Order	Alliance	Association/Phytocoenon	Reference number in the specific bibliographical list of Supplement 1
75.1.1	Balloto nigrae-Robinion	Jurko ex Hadac & Sofron 1980		
		<i>Ailanthes altissima</i> community		88
75.1.2	Bryonio-Robinion	Ubaldi, Melloni & Cappelletti in Ubaldi 2003		
		<i>Ailanthes altissima</i> community		46
		<i>Bryonio-Robinietum</i> Ubaldi, Melloni & Cappelletti in Ubaldi 2003		124
		<i>Robinia pseudoacacia</i> community		46, 98, 128, 130
75.1.3	Lauro nobilis-Robinion pseudoacaciae	Allegrezza, Montecchiari, Ottaviani, Pelliccia & Tesei 2019		
		<i>Meliso altissimae-Robinietum pseudoacaciae</i> Allegrezza, Montecchiari, Ottaviani, Pelliccia & Tesei 2019		2
		<i>Rubio peregrinae-Robinietum pseudoacaciae</i> Allegrezza, Montecchiari, Ottaviani, Pelliccia & Tesei 2019		2

BRYOPHYTE COMMUNITIES

CERATODONTO PURPUREI-POLYTRICHETEA PILIFERI Mohan 1978

POLYTRICHETALIA PILIFERI von Hübschmann 1975

Campylopodium polytrichoides Giacomini 1951*Campylopus introflexus* community 108, 111

(*) In Poldini et al. (1998) the associations *Panico-Polygonetum persicariae* Pignatti 1953 and *Oxalido-Chenopodietum polyspermi* (Br.-Bl. 1921) Sissingh (1942) 1946 are considered syntaxonomic synonyms of *Echinochloo-Setarietum pumilae* Felföldy 1942 corr. Mucina 1993. The authors reached this result by elaborating the synthetic tables of only a part of the works published at national level with the name *Panico-Polygonetum* and *Oxalido-Chenopodietum*. Beside this, in Poldini et al. (1988) the numbers of tables and relevés taken into account for the analysis were not indicated. These authors did not also consider any subassociations and variants described. For these reasons, we prefer to report the names of the associations as they were indicated in the original works, without including them as synonyms in the name *Echinochloo-Setarietum*.

(**) This association was described for the north-Adriatic Italian coasts, published by Pignatti (1952-53) as " nom. prov.", therefore not validly, according to ICPN (Art. 3b, Weber et al., 2000). It cannot be attributed to *Atriplicion littoralis* sensu Nordhagen 1940, as already highlighted by Mucina et al. (2016, p. 137), according to whom it must be referred to ruderal communities of *Atriplicion Passarge 1978 (Sisymbrietalia)*. After examining the original table in Pignatti (1952-53), we agree with the comments of Mucina et al. (2016). Anyway, in the Italian Vegetation Prodrome, the alliance *Atriplicion Passarge 1978* is not reported, so we provisionally prefer to attribute this association to *Hordeion leporini*, the most similar alliance from the eco-coenological point of view present in the Italian Vegetation Prodrome.

Soon after the development and data analysis of this article, Pellizzari (2020) hypothesized a reinterpretation of *Cyperus glomeratus*, currently considered alien species

in Italy, as a probable native. We are currently sticking to the consolidated position of Galasso et al. (2018), pending a reassessment of the chorology of this species.

Appendix II – Specific bibliographical list with reference numbers for all the syntaxa quoted in the checklist of Appendix I

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