

Targeting cancer vasculature via endoglin/CD105: a novel antibody-based diagnostic and therapeutic strategy in solid tumours

Ester Fonsatti¹, Hugues J.M. Nicolay^{1,2}, Maresa Altomonte¹, Alessia Covre^{1,2}, and Michele Maio^{1,2*}

¹Division of Medical Oncology and Immunotherapy, Department of Oncology, Istituto Toscano Tumori, University Hospital of Siena, Strada delle Scotte 14, 53100 Siena, Italy; and ²Cancer Bioimmunotherapy Unit, Department of Medical Oncology, Centro di Riferimento Oncologico, Istituto di Ricovero e Cura a Carattere Scientifico 33081 Aviano, Italy

Received 19 March 2009; revised 14 September 2009; accepted 5 October 2009; online publish-ahead-of-print 7 October 2009

Time for primary review: 33 days

Endoglin/CD105 is well acknowledged as being the most reliable marker of proliferation of endothelial cells, and it is overexpressed on tumour neovasculature. Our current knowledge of its structure, physiological role, and tissue distribution suggests that targeting of endoglin/CD105 is a novel and powerful diagnostic and therapeutic strategy in human malignancies, through the imaging of tumour-associated angiogenesis and the inhibition of endothelial cell functions related to tumour angiogenesis. Among biotherapeutic agents, monoclonal antibodies have shown a major impact on the clinical course of human malignancies of different histotypes. Along this line, the potential efficacy of anti-endoglin/CD105 antibodies and their derivatives for clinical purposes in cancer is supported by a large body of available pre-clinical *in vitro* and *in vivo* data. In this review, the main findings supporting the translation of antibody-based endoglin/CD105 targeting from pre-clinical studies to clinical applications in human cancer are summarized and discussed.

Keywords Angiogenesis • Cancer • Endoglin/CD105 • Monoclonal antibodies • Therapy

1. Introduction

A large amount of experimental evidence has shown that endoglin/ CD105 is expressed on endothelial cells of both mature and immature blood vessels,¹⁻³ and that it is overexpressed in vascular endothelial cells of tissues undergoing angiogenesis such as regenerating and inflamed tissues or tumours.³⁻⁶ Furthermore, levels of endoglin/CD105 positively correlate with the extent of endothelial cell proliferation^{4,5} and with the expression of proliferation markers in tumour endothelia.⁶ In addition, endoglin/CD105 has been suggested to be the most suitable marker available to quantify tumour angiogenesis.^{1,3,5,7,8} Lastly, intratumour microvessel density (IMVD) assessed by endoglin/CD105 staining strongly correlates with prognosis in cancer patients.^{1,3,9,10} Altogether, these findings support the role of endoglin/ CD105 as an optimal marker of proliferation of endothelial cells and its emerging clinical potential as prognostic, diagnostic, and therapeutic vascular target in human cancer.^{1,4,11,12}

2. Structure

Endoglin/CD105 is a disulphide-linked homodimeric transmembrane protein $^{13-15}$ of 180 $k\mathrm{Da}^{16,17}$ with short intracellular and

transmembrane domains and a large extracellular region.^{13–15} The latter consists of an orphan and a zona pellucida (ZP) domain, with an arginine-glycine-aspartic acid (RGD) binding motif,^{15,18,19} while the intracellular region presents several potential phosphorylation sites.²⁰ Four potential N-linked glycosylation sites and a region of O-linked glycosylation rich in serine and threonine have been described.¹⁵

A high amino acid sequence homology was observed among human, porcine, and murine endoglin/CD105 proteins^{21,22} with major differences in the extracellular domain.²² The RGD tripeptide was detected only in the human protein. Furthermore, endoglin/CD105 shows homology with the TGF- β receptor (T β R) type III betaglycan in the transmembrane and cytoplasmic domains.^{2,23}

The analysis of the three-dimensional structure of the extra-cellular region of endoglin/CD105 by single-particle electron microscopy identified three well-defined domains for each monomer region, including the two ZP regions and one orphan domain.¹⁹

The gene coding for endoglin/CD105 maps on human chromosome $9q34 \rightarrow qter^{24}$ and cloning of its promoter demonstrated a strong and selective activity in endothelial cells.^{25,26,27} Alternative spliced transcript variants encoding two different isoforms of endoglin/CD105 (L- and S-CD105) have been described.^{28,29} The two

* Corresponding author. Tel: +39 0577 586335; fax: +39 0577 586303, Email: mmaio@cro.it

Published on behalf of the European Society of Cardiology. All rights reserved. © The Author 2009. For permissions please email: journals.permissions@oxfordjournals.org.

forms differ in the length of the cytoplasmic domain and show a different pattern of tissue distribution.^{27,28,29} L-CD105, the longer variant, is predominantly expressed in endothelial cells²⁹ and consists of 658 amino acids residues^{15,21} with a higher extent of phosphorylation compared with S-CD105.²⁹

3. Expression

Studies on the cellular and tissue distribution of endoglin/CD105 suggest its profound functional involvement in angiogenesis, vascular development, and homeostasis. Concerning the role of endoglin/CD105 in the development of the cardiovascular system, endoglin/CD105 was exclusively found on vascular endothelia of human embryos at 4–8 weeks of gestation, and it was transiently up-regulated during heart septation and heart valves formation.³⁰ Furthermore, an altered expression of endoglin/CD105 is observed in human foetuses with cardiac defects.³¹ The involvement of endoglin/CD105 in vascular development is enforced by the observation that endoglin/CD105 *null* mice die during early gestation, due to structural alterations in the primitive vascular plexus of the yolk sac that prevents the formation of normal mature vessels, as reported for transforming growth factor (TGF)- β 1, TGF- β receptor (T β R) type I activin receptor-like kinases (ALK)-1, and ALK-5 *null* mice.³²

In human adult tissues, endoglin/CD105 expression is mainly restricted to vascular endothelial and stromal cells,^{1,4} while it is weakly detectable on activated monocytes, macrophages, erythroid precursors, fibroblasts, mesangial cells, follicular dendritic cells, and melanocytes,^{1,33} but it is highly expressed on syncytiotrophoblasts of term placenta.³⁴

In cultured endothelial cells, the highest levels of endoglin/CD105 were detectable in cells showing levels of RNA, DNA, and proteins consistent with cellular proliferation and activation.⁴ Consistently, levels of endoglin/CD105 expression correlated with the rate of cellular proliferation and density in cultures of human umbilical vein endothelial cells (HUVEC),^{1,5} and high levels of endoglin/CD105 associated with other markers of proliferation such as cyclin A and Ki-67 in tumour endothelium.⁶ Supporting these in vitro data, and highly suggestive for its involvement in tumour angiogenesis, an up-regulated expression of endoglin/CD105 was found on endothelia of angiogenetic blood vessels within tumour tissues^{4,5,7,8} where, at variance with the pan-endothelial markers CD31 and CD34, it was also detected in neovessels with strong remodelling activity and in immature neovessels.^{3,35} Among tumours of different histotype, staining of endoglin/CD105 was invariably observed in peri- and intratumoural blood vessels and on tumour stromal components;^{1,5,36} and, on the other hand, only a weak expression of endoglin/CD105 was detected in the cytoplasm of neoplastic cells of selected histotypes.^{1,5,36}

4. Functions

Biochemical and biological functions of endoglin/CD105 are under active investigation, and new data continue to emerge on its functional role within the TGF- β receptor complex, on its modulation of cellular responses to TGF- β , and on its involvement in vascular physiology and angiogenesis.^{12,14}

As reported for betaglycan, endoglin/CD105 is a T β R type III auxiliary receptor^{12,16,37} that modulates the signalling response to TGF- β , a pleiotropic cytokine involved in tumour development and It has been demonstrated that endoglin/CD105 requires the association with T β R type II to bind ligands,^{17,23,38} and that it can interact with T β R type I or type II in the absence of ligand.³⁸ Based on different experimental data, it has been proposed that through its interactions with the T β R type I and type II, endoglin/CD105 regulates their phosphorylation status and subsequently their signalling ability.³⁸ However, endoglin/CD105 can also act independently of the TGF- β signalling pathway.¹²

In endothelial cells, two T β R type I pathways with opposite effects have been identified: the ALK-5 that induces Smad 2/3 phosphorylation and the ALK-1 that induces Smad 1/5 phosphorylation.^{39,40} The latter is known to promote endothelial cell proliferation, migration, and tube formation, while the first inhibits these cellular responses to TGF- β .⁴⁰ Evidence suggests that endoglin/CD105 is required for TGF- β /ALK1 signalling and indirectly inhibits TGF- β / ALK5 signalling.^{40,41}

Down-regulation of endoglin/CD105 expression through small interfering RNA demonstrated that endoglin/CD105 promotes TGF- β -induced Smad 1/5 phosphorylation, and proliferation and migration of murine endothelial cells via ALK-1 receptor. In fact, it has been shown that loss of endoglin/CD105 abrogates ALK-1 signalling and endothelial cells proliferation, while endoglin/CD105 haploin-sufficiency leads to a down-regulation of surface ALK-5 expression, probably as an adaptation mechanism.³⁹ These results suggest that endoglin/CD105 is required for efficient TGF- β /ALK-1 signalling and that it acts as a modulator factor of the balance between TGF- β /ALK-1 and TGF- β /ALK-5 signalling pathways³⁹ (*Figure 1*).

Available data on the interaction of endoglin/CD105 with T β R type I and type II suggest that the extracellular and the cytosolic domains of endoglin/CD105 play a distinct role in the TGF- β receptor signalling,^{14,17,23,38} and that the association of CD105 with ALK-1 is crucial for the response of endothelial cells to TGF- β .^{39,42,43} Along this line, distinct levels of endoglin/CD105 differentially modulate cellular responses to TGF- β including: cellular proliferation, adhesion and migration, homotypic cell aggregation, and expression of selected matrix components (i.e. plasminogen activator inhibitor-1, collagen, fibronectin, lumican).^{1,23,37,44-48} In particular, the deletion of endoglin/CD105 enhances the ability of TGF- β 1 to suppress growth, migration, or microvessels formation in HUVEC and smooth muscle cells, as well as apoptosis induced by hypoxia or TGF- β 1 in endothelial cells.^{37,45,49} Consistently, reduced proliferation and migration, increased basal or TGF- β 1-mediated collagen synthesis, impaired capillary tube formation, and vascular endothelial growth factor (VEGF) secretion are reported in endothelial cells derived from endo $glin/CD105 \pm$ adult mice.⁵⁰

Supporting its role in vascular homeostasis and integrity, endoglin/ CD105 was identified as a component of endothelial nitric oxide synthase (eNOS) activation pathway and a modulator of COX-2 expression and activity.^{51–54} Thus, endoglin/CD105 can also regulate the vascular tone by maintaining the fine balance between eNOS and COX-2 in endothelial cells.⁵⁴ Mutations in the *endoglin/CD105* gene

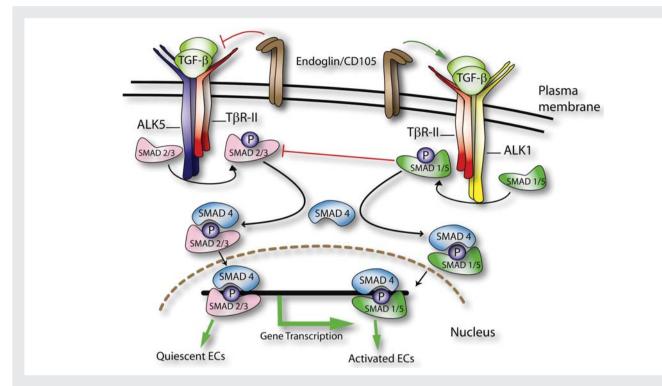


Figure I A schematic hypothetical role of endoglin/CD105 in TGF- β /ALK-1 and TGF- β /ALK-5 signalling pathways in endothelial cells. Available data in literature suggest that the levels of expression of endoglin/CD105 on endothelial cells (ECs) can affect the response of endothelial cells to TGF- β modulating their proliferation. In endothelial cells, TGF- β can activate two type I receptor pathways with opposite effects: ALK-5 inducing Smad 2/3 phosphorylation and ALK-1 promoting Smad 1/5 phosphorylation. Endoglin/CD105 binds TGF- β by associating with TGF- β signalling receptors type (T β R)-II. This association results in an altered phosphorylation state of T β R-II promoting endothelial cells proliferation via TGF- β /ALK-1 signalling pathway and inducing an indirect inhibition of ALK-5 signalling pathway. Upon activation, phosphorylated Smads form heteromeric complexes with the common mediator Smad 4, which in the nucleus act as transcription factor complexes regulating the transcriptional activity of target genes.

are associated to hereditary hemorrhagic telangiectasia (HHT) type 1,^{19,55,56} an hereditary disease characterized by arteriovenous malformations and recurrent bleeding. It was in fact reported that endoglin/ CD105 haploinsufficiency is the cause of defective vascular repair in HHT type 1 that may in part explain the heterogeneity of symptoms observed among members of the same family.⁵⁷

Inhibition of endothelial cell apoptosis is crucial during angiogenesis and vasculogenesis, and it is well-known that pro-angiogenetic factors also promote survival of endothelial cells. In this setting, endoglin/ CD105 was shown to protect hypoxic endothelial cells from apoptosis either in the presence or absence of TGF- β 1.⁴⁹ Supporting this finding, an inverse correlation between the extent of apoptosis and IMVD assessed by anti- endoglin/CD105 antibodies was reported in non-small cell lung cancer patients.⁵⁸ Thus, in tumour angiogenesis, endoglin/CD105 may also play a role in activating anti-apoptotic signals in hypoxic endothelial cells.

At variance with the extensive knowledge on the activity of endoglin/CD105 in endothelial cells, limited and conflicting data are available on the functional role of endoglin/CD105 on neoplastic cells. *In vitro* studies reported that overexpression of endoglin/CD105 on metastatic prostate cancer^{59,60} and oesophageal squamous carcinoma cells⁶¹ decreased their invasiveness, while this ability was improved by the up-regulation of endoglin/CD105 in breast cancer cells.⁶² These findings, and its overall limited expression on neoplastic cells,^{1,5,36} clearly suggest that additional studies are required to unveil the possible functional significance of endoglin/CD105 on tumour cells of different histotype.

5. Tumour angiogenesis

A number of studies have reported that the assessment of IMVD by endoglin/CD105 staining represents a good prognostic indicator in different malignancies. Endoglin/CD105-positive blood vessels count negatively correlated with overall survival, disease-free survival, or presence of distant metastases in a wide range of human tumours.^{9,10,12,36,63,64} Additionally, IMVD score by anti-endoglin/ CD105 mAb was shown to positively correlate with Gleason score in prostate cancer patients³ and with the tumour stage in squamous cell carcinomas of the oral cavity.⁶⁵ These studies also indicated that blood vessels count by endoglin/CD105 staining is a more informative marker of prognosis as compared with staining by other pan-endothelial markers.^{1,9,10,12} Consistently, an increase in IMVD was assessed by endoglin/CD105 mAb during progressive stages of colorectal carcinogenesis from low- to high-grade dysplasia, and from high-grade dysplasia to carcinoma.⁶⁶ This evidence provided indirect support to the role of endoglin/CD105 in tumour angiogenesis and to the usefulness of endoglin/CD105 targeting for antiangiogenetic therapy of cancer.^{1,12}

To specifically investigate the role of endoglin/CD105 in tumour angiogenesis and growth, endoglin/CD105 haploinsufficient

(endoglin/CD105+/-) and control littermates (endoglin/CD105+/+) mice were subcutaneously inoculated with Lewis lung carcinoma cells. The results of these studies indicated that endoglin/CD105 deficiency decreased tumour vascularization and growth in mice, further supporting the idea that endoglin/CD105 is involved in tumour angiogenesis.⁶⁷ In addition, a significant decrease in the levels of eNOS and of phosphorylated eNOS was reported in *endoglin/CD105+/-* mice compared with control mice, suggesting that this deficiency might be responsible of the decreased angiogenesisdependent tumour yascularization in *endoglin/CD105+/-* mice.⁶⁷ Instead, the lower tumour vascularization in *endoglin/CD105+/-* mice was not associated with lower levels of the major angiogenetic factors, such as hypoxia inducible factors and VEGF, advising that the involvement of endoglin/CD105 in tumour angiogenesis occurs downstream of these factors.⁶⁷

Unexpectedly, an increased number of *endoglin/CD105*-positive vessels and an up-regulation of *endoglin/CD105* gene expression within tumours were reported in mice following anti-angiogenetic treatment with anti-VEGF mAb⁶⁸ or with anti-VEGF Receptor-2⁶⁹ mAb, respectively, likely as a consequence of increased hypoxia induced by the treatment.

6. Targeting of endoglin/CD105 for cancer imaging

Solid malignancies are generally highly vascularized, and endothelial cells lining the tumour vasculature proliferate much faster than endothelial cells of blood vessels in normal tissues.^{70,71} Thus, an ideal target for the imaging of tumour vasculature should be overexpressed in actively proliferating endothelial cells but it should be weakly expressed or undetectable in guiescent endothelial cells; additionally, it should be expressed in a large amount on the luminal surface of the blood vessels. Based on these notions, the evidence that levels of endoglin/CD105 expression correlate with the proliferation rate of endothelial cells,⁴⁻⁶ and that it is overexpressed in endothelia within neoplastic tissues compared with those of normal tissues, prompted investigations on endoglin/ CD105 as a potential target for the diagnostic imaging of solid tumours.^{5,72} The feasibility of endoglin/CD105 targeting for diagnostic applications was addressed in two distinct animal models and utilizing two different radio-labelled anti-endoglin/CD105 mAb. The results indicated that targeting of endoglin/CD105 is a useful and safe procedure for tumour imaging,^{5,72} regardless of tumour histotype and independently from endoglin/CD105 expression on neoplastic cells.⁵ In particular, the intravenous injection of the ¹²⁵I-labelled anti-endoglin/CD105 mAb MAEND3 efficiently imaged spontaneous mammary canine adenocarcinomas. The up-take of the mAb into the tumour areas was described as rapid and intense, without systemic side effects in injected animals during a 3-month follow-up after imaging procedures.⁵ Consistent with these initial findings, the intravenous administration of the ¹¹¹In-labelled anti-endoglin/CD105 mAb MJ7/18 effectively imaged allografts of melanoma in C57BL/6 mice.⁷² The autoradiography and the subsequent immunohistology showed that the highest levels of mAb were concentrated in the periphery of the tumour mass where vessel density is prominent, with a heterogeneous distribution in the tumour centre. The blood half-life of the antibody was reported to be less than 1 min.⁷²

Important clinical implications of endoglin/CD105 targeting in improving cancer diagnosis were further shown by an ex vivo study in human renal carcinoma.⁷³ The ⁹⁹Tcm-labelled anti-endoglin/ CD105 mAb E9 was perfused in the renal artery of freshly excised kidneys from seven patients diagnosed with renal carcinoma. The scintigraphy identified hot spots of radioactivity, which matched with the position of the neoplastic lesions. The specificity of the localization of the labelled anti-endoglin/CD105 mAb into the tumour mass was confirmed by the observation that a previous perfusion of unlabelled mAb completely blocked the localization of ⁹⁹Tcm-conjugated mAb. Noteworthy, the anti-endoglin/CD105 mAb identified two tumour masses previously undetected by pre-surgery magnetic resonance imaging scan.⁷³ Altogether, these imaging studies clearly indicate that in vivo anti-endoglin/CD105 mAb specifically and strongly target the tumour vasculature, and provide support to their usefulness also for therapeutic anti-angiogenic approaches in human cancer.

Targeting of endoglin/CD105 has also been suggested as a possible strategy to monitor the response to cancer therapy. Microbubbles are suitable contrast agents for the enhancement of ultrasound images. and avidin can be incorporated into the shell of microbubbles, acting as a direct ligand for biotinylated mAb. Utilizing this approach, microbubbles conjugated with the anti-endoglin/CD105 mAb MJ7/18 specifically bound in vitro to murine brain capillary endothelial cells expressing high levels of endoglin/CD105, but not to murine fibroblasts expressing low levels of endoglin/CD105.74 Being ultrasound a powerful non-invasive diagnostic tool, microbubbles conjugated to mAb MJ7/18 were also utilized to image and quantify vascular effects of cytotoxic therapy in a mouse model of pancreatic adenocarcinoma.⁷⁵ The results obtained showed a significantly decreased ultrasound signal in tumours treated with gemcitabine.⁷⁵ Thus, microbubbles conjugated with anti-endoglin/CD105 mAb may represent a foreseeable non-invasive tool for the imaging of tumour angiogenesis and for monitoring vascular effects of anti-cancer therapy.

7. Targeting of endoglin/CD105 for cancer treatment

The well-acknowledged notion that angiogenesis supports primary tumour growth and its metastatic progression has prompted the investigation of different anti-cancer strategies aimed to inhibit the formation of new blood vessels and/or to disrupt tumour-associated blood vessels.^{76,77} In principle, these therapeutic modalities bear the advantage of overcoming several limitations commonly related to the targeting of transformed cells such as their inherent drug resistance, the poor delivery of drugs to neoplastic cells, and the need to target a biologically highly heterogeneous malignant cell population.^{76,78}

Among antibodies with anti-angiogenetic activity so far tested in cancer patients, only the humanized anti-VEGF mAb bevacizumab has received approval by Regulatory Agencies, though for selected clinical indications.^{79–81} Thus, different novel agents are currently under active clinical investigation for cancer treatment, with the intent to starve neoplastic cells by blocking their tumour blood supply. In this setting, targeting of proliferating endothelial cells is generally considered the most promising therapeutic strategy to control tumour angiogenesis.

Strong experimental support to the possible use of anti-endoglin/ CD105 mAb for therapeutic vascular targeting in cancer derives from *in vitro* studies demonstrating that different anti-endoglin/ CD105 antibodies inhibit proliferation, migration and adhesion of endothelial cells^{33,82,83} and/or induce apoptosis in human umbilical vein endothelial cells.⁶⁷ Moreover, selected anti-endoglin/CD105 mAb showed a synergistic activity with TGF- β in the inhibition of endothelial cells proliferation.⁸²

Diverse engineered antibodies directed to endoglin/CD105 have also been generated and utilized to target or to deliver pharmacological agents to endothelial cells *in vitro*. A bispecific antibody directed to endoglin/CD105 and CD3 was utilized to mediate killing of endoglin/ CD105-positive endothelial cells by cytotoxic T lymphocytes.⁸⁴ Single-chain Fv fragments directed to endoglin/CD105 were used to generate immunoliposome of encapsulated therapeutic drugs to target endothelial cells.⁸⁵ these complexes were able to internalize into and kill endothelial cells.⁸⁵ Nanobodies against endoglin/CD105 have also been recently described⁸⁶ and represent another promising tool for therapeutic applications of endoglin/CD105 since they are small, non-immunogenic, stable, highly soluble, and easy to produce.

The therapeutic potential of endoglin/CD105 targeting has been already extensively investigated at pre-clinical level also in vivo. Several studies demonstrated a long-lasting regression/suppression of tumour growth and metastasis in SCID mice by the immunotoxinconjugated,^{87,88} radiolabelled,⁸⁹ or naked anti-endoglin/CD105 mAb⁹⁰⁻⁹³ termed SN6f, SN6j, and SN6 k, likely mediated by the inhibition of tumour-associated angiogenesis, and/or by the destruction of tumour-associated vasculature. Furthermore, utilizing a human skin/ SCID mouse chimera model, the naked human anti-endoglin/CD105 mAb SN6f, SN6j, and SN6 k suppressed human blood vessels but poorly inhibited murine vessels in established tumours.⁹² Interestingly, this study also showed that the in vivo anti-tumour efficacy of tested anti-endoglin/CD105 mAb was not proportional to their antigenbinding avidities, and that the combined administration of SN6f and SN6 k, directed to non-overlapping epitopes of endoglin/CD105, showed an additive anti-tumour effect.⁹² Furthermore, the co-administration of mAb SN6j and cyclophosphamide had a synergistic anti-tumour efficacy and completely suppressed tumours in some chimeras.92

Growth suppression of established tumours of colon-26 murine colon carcinoma cells and of 4T1 murine mammary carcinoma cells by systemic administration of unconjugated anti-endoglin/CD105 mAb SN6j was also reported in immunocompetent tumour bearing BALB/c mice.⁹¹ Differences in tumour growth rate and therapeutic response were found to depend on the tumour location both in immunocompetent BALB/c mice and in immunodeficient SCID mice.⁹¹ The anti-endoglin/CD105 mAb was more effective against skin tumours with a low growth rate, as compared with rapidly growing intramuscular tumours.⁹¹ Consistent with these studies, the anti-tumour activity of a recently characterized anti-endoglin/CD105 mAb was also demonstrated in BALB/c and C57BL mice inoculated with H22 or Hepa1-6 hepatoma cells, respectively.⁹⁴ In both models, the administration of the mAb induced a significant inhibition of tumour growth and increased the survival rate.94 The analysis of the tumour tissues excised from treated mice showed an increase in apoptotic cells and a decrease in microvessels, compared with those removed from control mice.⁹⁴

The anti-tumour activity of anti-endoglin/CD105 mAb was found improved in immunocompetent compared with immunodeficient

mice.⁹⁰ Confirming the hypothesis that T cell immunity could play a role in enhancing the efficacy of anti-endoglin/CD105 antibody-based therapy, the administration of an immune activator CpG ODN, enhanced the anti-tumour efficacy of the anti-endoglin/CD105 mAb SN6j in immunocompetent BALB/c mice, but not in immunodeficient SCID mice. Supporting the notion that immune mechanisms are involved in antibody-based endoglin/CD105-targeted tumour therapy, the anti-tumour efficacy of the SN6j mAb was abrogated when CD4+ and/or CD8+ T cells were depleted from BALB/c mice.⁹⁰

Noteworthy for perspective clinical applications in patients, three mAb (SN6a, SN6j, and SN6 k) recognizing distinct epitopes of endoglin/CD105 showed therapeutic efficacy against tumour metastases in BALB/c mice inoculated with 4T1 murine mammary carcinoma or colon-26 murine adenocarcinoma cell lines.⁹³

In order to assess the potential clinical application of anti-endoglin/ CD105 antibodies, the recombinant human/mouse chimeric antibody of IgG1 isotype designated c-SN6j was generated from mAb SN6j, based on its strong anti-tumour efficacy *in vivo* and its strong ability to inhibit the proliferation of endothelial cells *in vitro*.⁹⁵ When administered in monkeys, mAb c-SN6j showed pharmacokinetic parameters comparable to those reported in humans for different therapeutic mAb.⁹⁵

In an attempt to construct a novel anti-endoglin/CD105 immunoconjugated mAb with limited unspecific toxicity, the type 2 ribosomeinactivating protein nigrin b was linked to the anti-human endoglin/ CD105 mAb 44G4. This 44G4 nigrin b immunotoxin specifically killed murine fibroblasts expressing the short isoform of human endoglin/CD105 and in a perinuclear region.⁹⁶

A phase I study, first-in human, with the human/murine chimeric anti-endoglin/CD105 mAb TRC105 in patients with refractory advanced or metastatic solid cancer is ongoing. Objectives of the study are to evaluate safety and tolerability of escalating doses of the therapeutic mAb, its pharmacokinetics and immunogenicity, as well as signs of clinical activity. Preliminary data were most recently reported showing that treatment is well tolerated at doses up to 1 mg/Kg and with evidence of clinical activity. Two dose-limiting toxicity were observed: a Grade 3 hypersensitivity reaction and a Grade 4 bleeding. Though initial, these findings indicate that anti-VEGF mAb and anti-endoglin/CD105-based treatment might share bleeding as a common adverse event.⁹⁷

8. Conclusions and future directions

In light of its involvement in vascular development, morphogenesis and physiology, and of its strong expression on blood vessels of tumour tissues, the potential of endoglin/CD105 as vascular target for diagnostic and therapeutic anti-angiogenetic strategies in cancer has been extensively investigated at pre-clinical level both *in vitro* and *in vivo* (summarized in *Table 1*).

Endoglin/CD105 is overexpressed on proliferating endothelial cells^{1–5} and in the vascular endothelium within tumour tissues,^{1,4,6–}^{8,98} but also in endothelia of normal tissues.^{98,99} This latter notion raised concerns about potential side effects of clinical applications of endoglin/CD105 targeting in cancer patients.^{98–102} However, it was convincingly argued that most tissue antigens targeted by therapeutic antibodies that are successfully employed in the clinic are not tumour specific.¹⁰⁰ Nevertheless, not all available anti-endoglin/CD105 mAb are likely good candidates for therapeutic applications

Table I Pre-clinical studies with different anti-endoglin/CD105 mAb

mAb	In vitro		In vivo		
	Activity	Cell type	Activity	Tumour type	Host
MJ7/18	Target binding ⁷⁴	EC (M)	Cancer imaging ⁷² Vascular imaging ⁷⁵	Melanoma (M) Pancreatic adenocarcinoma (M)	C57BL/6 mice nu/nu mice
MAEND3	-		Cancer imaging ⁵	Spontaneous mammary carcinoma	Beagle dog, mixed breed dog
E9	-		Cancer imaging ⁷³	Renal carcinoma (H)	Excised human kidneys
SN6f	-		Anti-tumour ^{87,89,92} Anti-angiogenetic ^{87,89,92}	Breast cancer (H)	H skin/SCID mouse chimeras, SCID mice
SN6k	-		Anti-tumour ^{88,92,93} Anti-angiogenetic ^{88,92}	Mammary carcinoma (M), Colon adenocarcinoma (M), Breast cancer (H)	BALB/c mice, H skin/ SCID mouse chimeras, SCID mice
SN6J	Inhibition of proliferation ^{82,90} Induction of apoptosis ⁹⁰ ADCC ⁹³	HUVEC	Anti-tumour ^{90–93} Anti-angiogenetic ^{92,93}	Mammary carcinoma (M), Colon adenocarcinoma (M), Breast cancer (H)	BALB/c mice, H skin/ SCID mouse chimeras, SCID mice
SN6	Inhibition of proliferation ⁸²	HUVEC	-		
SN6a	Inhibition of proliferation ⁸²	HUVEC	Anti-tumour ⁹³ Anti-angiogenetic ⁹³	Mammary carcinoma (M), Colon adenocarcinoma (M)	BALB/c mice
SN6h	Inhibition of proliferation ⁸²	HUVEC	-		
TEC11	Inhibition of proliferation ^{4,83} Inhibition of urokinase production ⁸³	HUVEC, HDMC, vascular EC (H)	-		
44G4	Target binding ⁹⁶	Fibroblasts (M)	-		

EC, endothelial cells; HUVEC, human umbilical vein endothelial cells; ADCC, antibody-dependent cellular cytotoxicity, HDMC, human dermal microvascular endothelial cells; H, human; M, murine.

Table 2 Points to consider about anti-endoglin/CD105 mAb use for imaging and treatment of human tumours

Strenghts	Limitations		
Optimal accessibility from the blood stream	Presence of a soluble form		
Over-expression in tumour endothelia compared with endothelia of normal tissues	Extent of tumour vascularization		
Stable expression	Limited by tumour size? (imaging)		
Limited background (imaging)	Presence and extent of tumour necrosis (imaging)		
No tumour-histotype specificity	Bleeding as side effect? (therapy)		
Independence from the expression on neoplastic cells			
Detectable in mature and immature neovessels			
No major side effects in animal models			
Useful to evaluate the efficacy of anti-angiogenetic treatments			

in cancer since differences in their reactivity with the tumour vasculature have been observed. 92,98,100

As a whole, the pre-clinical data available undoubtedly point to endoglin/CD105 as an attractive vascular target to design novel antibody-based diagnostic and therapeutic strategies shared by different human malignancies, suggesting that times are mature to identify the most appropriate therapeutic applications of endoglin/ CD105 targeting in the clinical setting of cancer.¹⁰³ Along this line, it has been recently shown that, unlike other pan-endothelial markers, endoglin/CD105 is overexpressed in hepatic epithelioid hemangioendothelioma (HEH), a rare neoplasm of endothelial origin, clearly pointing to endoglin/CD105 as a possible candidate for the therapeutic targeting of HEH.¹⁰⁴

It is finally worth mentioning that, other than on endothelial cells, endoglin/CD105 is weakly expressed on selected cell types that indirectly contribute to tumour angiogenesis such as mural, stromal, and inflammatory cells.¹ Thus, it seems reasonable to envisage that the anti-angiogenetic efficacy of antibodies directed to endoglin/CD105 might be strengthened by their ability to target multiple cell types involved in the angiogenetic process besides endothelial cells.

In the last few years significant advances have been made on the biology of endoglin/CD105 in cancer, allowing to identify potential advantages and drawbacks associated with its diagnostic and therapeutic use (*Table 2*); altogether, time seems mature to translate the results emerged from pre-clinical research to the clinical setting. Along this line, a multicenter clinical trial is in progress⁹⁷ and its results are eagerly awaited to validate endoglin/CD105 as a novel target for cancer treatment.

Conflict of interest: none declared.

Funding

This work was supported in part by grants from the Associazione Italiana per la Ricerca sul Cancro (H.J.M.N. fellow), by the Istituto Superiore di

Sanità (ISS) and Alleanza Contro il Cancro, and by the Programma Italia-USA 'Malattie Rare' from the ISS (A.C. fellow).

References

- Fonsatti E, Del Vecchio L, Altomonte M, Sigalotti L, Nicotra MR, Coral S et al. Endoglin: an accessory component of the TGF-beta-binding receptor-complex with diagnostic, prognostic, and bioimmunotherapeutic potential in human malignancies. J Cell Physiol 2001;188:1–7.
- Wong SH, Hamel L, Chevalier S, Philip A. Endoglin expression on human microvascular endothelial cells association with betaglycan and formation of higher order complexes with TGF-beta signalling receptors. *Eur J Biochem* 2000;267:5550–5560.
- Wikstrom P, Lissbrant IF, Stattin P, Egevad L, Bergh A. Endoglin (CD105) is expressed on immature blood vessels and is a marker for survival in prostate cancer. *Prostate* 2002;**51**:268–275.
- Burrows FJ, Derbyshire EJ, Tazzari PL, Amlot P, Gazdar AF, King SW et al. Up-regulation of endoglin on vascular endothelial cells in human solid tumors: implications for diagnosis and therapy. *Clin Cancer Res* 1995;1:1623–1634.
- Fonsatti E, Jekunen AP, Kairemo KJ, Coral S, Snellman M, Nicotra MR et al. Endoglin is a suitable target for efficient imaging of solid tumors: *in vivo* evidence in a canine mammary carcinoma model. *Clin Cancer Res* 2000;6:2037–2043.
- Miller DW, Graulich W, Karges B, Stahl S, Ernst M, Ramaswamy A et al. Elevated expression of endoglin, a component of the TGF-beta-receptor complex, correlates with proliferation of tumor endothelial cells. *Int J Cancer* 1999;81:568–572.
- Wang JM, Kumar S, Pye D, van Agthoven AJ, Krupinski J, Hunter RD. A monoclonal antibody detects heterogeneity in vascular endothelium of tumours and normal tissues. *Int J Cancer* 1993;**54**:363–370.
- Wang JM, Kumar S, Pye D, Haboubi N, al-Nakib L. Breast carcinoma: comparative study of tumor vasculature using two endothelial cell markers. J Natl Cancer Inst 1994;86:386–388.
- Kumar S, Ghellal A, Li C, Byrne G, Haboubi N, Wang JM et al. Breast carcinoma: vascular density determined using CD105 antibody correlates with tumor prognosis. *Cancer Res* 1999;**59**:856–861.
- Shariat SF, Karam JA, Walz J, Roehrborn CG, Montorsi F, Margulis V et al. Improved prediction of disease relapse after radical prostatectomy through a panel of preoperative blood-based biomarkers. *Clin Cancer Res* 2008;14:3785–3791.
- ten Dijke P, Goumans MJ, Pardali E. Endoglin in angiogenesis and vascular diseases. Angiogenesis 2008;11:79–89.
- Bernabeu C, Lopez-Novoa JM, Quintanilla M. The emerging role of TGF-beta superfamily coreceptors in cancer. *Biochim Biophys Acta* 2009; [Epub ahead of print] PubMed PMID: 19607914.
- Haruta Y, Seon BK. Distinct human leukemia-associated cell surface glycoprotein GP160 defined by monoclonal antibody SN6. Proc Natl Acad Sci USA 1986;83: 7898–7902.
- Bernabeu C, Conley BA, Vary CPH. Novel biochemical pathways of endoglin in vascular cell physiology. J Cell Biochem 2007;102:1375–1388.
- Gougos A, Letarte M. Primary structure of endoglin, an RGD-containing glycoprotein of human endothelial cells. J Biol Chem 1990;265:8361–8364.
- Cheifetz S, Bellon T, Cales C, Vera S, Bernabeu C, Massague J et al. Endoglin is a component of the transforming growth factor-beta receptor system in human endothelial cells. J Biol Chem 1992;267:19027–19030.
- Barbara NP, Wrana JL, Letarte M. Endoglin is an accessory protein that interacts with the signaling receptor complex of multiple members of the transforming growth factor-beta superfamily. J Biol Chem 1999;274:584–594.
- Jovine L, Darie CC, Litscher ES, Wassarman PM. Zona Pellucida domain proteins. Annu Rev Biochem 2005;74:83–114.
- Llorca O, Trujillo A, Blanco FJ, Bernabeu C. Structural model of human endoglin, a transmembrane receptor responsible for hereditary hemorrhagic telangiectasia. *J Mol Biol* 2007;365:694–705.
- Lastres P, Martin-Perez J, Langa C, Bernabeu C. Phosphorylation of the human-transforming-growth-factor-beta-binding protein endoglin. *Biochem J* 1994; 301:765–768.
- Ge AZ, Butcher EC. Cloning and expression of a cDNA encoding mouse endoglin, an endothelial cell TGF-beta ligand. *Gene* 1994;138:201–206.
- Yamashita H, Ichijo H, Grimsby S, Moren A, ten Dijke P, Miyazono K. Endoglin forms a heteromeric complex with the signaling receptors for transforming growth factorbeta. J Biol Chem 1994;269:1995–2001.
- Letamendia A, Lastres P, Botella LM, Raab U, Langa C, Velasco B et al. Role of endoglin in cellular response to transforming growth factor-beta. A comparative study with betaglycan. J Biol Chem 1998;273:33011–33019.
- Fernández-Ruiz E, St-Jacques S, Bellón T, Letarte M, Bernabéu C. Assignment of the human endoglin gene (END) to 9q34→qter. *Cytogenet Cell Genet* 1993;64:204–207.
- Graulich W, Nettelbeck DM, Fischer D, Kissel T, Muller R. Cell type specificity of the human endoglin promoter. *Gene* 1999;227:55–62.
- Velasco B, Ramirez JR, Relloso M, Li C, Kumar S, Lopez-Bote JP et al. Vascular gene transfer driven by endoglin and ICAM-2 endothelial-specific promoters. *Gene Ther* 2001;8:897–904.

- Cowan PJ, Shinkel TA, Fisicaro N, Godwin JW, Bernabeu C, Almendro N et al. Targeting gene expression to endothelium in transgenic animals: a comparison of the human ICAM-2, PECAM-1 and endoglin promoters. *Xenotransplantation* 2003;10: 223–231.
- Bellon T, Corbi A, Lastres P, Cales C, Cebrian M, Vera S et al. Identification and expression of two forms of the human transforming growth factor-beta-binding protein endoglin with distinct cytoplasmic regions. *Eur J Immunol* 1993;23: 2340–2345.
- Perez-Gomez E, Eleno N, Lopez-Novoa JM, Ramirez JR, Velasco B, Letarte M et al. Characterization of murine S-endoglin isoform and its effects on tumor development. Oncogene 2005;24:4450-4461.
- Qu R, Silver MM, Letarte M. Distribution of endoglin in early human development reveals high levels on endocardial cushion tissue mesenchyme during valve formation. *Cell Tissue Res* 1998;292:333–343.
- Barresi V, Grosso M, Vitarelli E, Triolo O, Barresi G. Endoglin (CD105) expression in the human heart throughout gestation: an immunohistochemical study. *Reprod Sci* 2008;**15**:1018–1026.
- Li DY, Sorensen LK, Brooke BS, Urness LD, Davis EC, Taylor DG et al. Defective angiogenesis in mice lacking endoglin. Science 1999;284:1534–1537.
- Dallas NA, Samuel S, Xia L, Fan F, Gray MJ, Lim SJ et al. Endoglin (CD105): a marker of tumor vasculature and potential target for therapy. *Clin Cancer Res* 2008;14: 1931–1937.
- St-Jacques S, Forte M, Lye SJ, Letarte M. Localization of endoglin, a transforming growth factor-beta binding protein, and of CD44 and integrins in placenta during the first trimester of pregnancy. *Biol Reprod* 1994;**51**:405–413.
- Nagatsuka H, Hibi K, Gunduz M, Tsujigiwa H, Tamamura R, Sugahara T et al. Various immunostaining patterns of CD31, CD34 and endoglin and their relationship with lymph node metastasis in oral squamous cell carcinomas. J Oral Pathol Med 2005; 34:70–76.
- Fonsatti E, Altomonte M, Nicotra MR, Natali PG, Maio M. Endoglin (CD105): a powerful therapeutic target on tumor-associated angiogenetic blood vessels. *Onco*gene 2003;22:6557–6563.
- Li C, Hampson IN, Hampson L, Kumar P, Bernabeu C, Kumar S. CD105 antagonizes the inhibitory signaling of transforming growth factor beta1 on human vascular endothelial cells. FASEB J 2000;14:55–64.
- Guerrero-Esteo M, Sanchez-Elsner T, Letamendia A, Bernabeu C. Extracellular and cytoplasmic domains of endoglin interact with the transforming growth factor-beta receptors I and II. J Biol Chem 2002;277:29197–29209.
- Lebrin F, Goumans MJ, Jonker L, Carvalho RL, Valdimarsdottir G, Thorikay M et al. Endoglin promotes endothelial cell proliferation and TGF-beta/ALK1 signal transduction. EMBO J 2004;23:4018–4028.
- Goumans MJ, Valdimarsdottir G, Itoh S, Rosendahl A, Sideras P, ten Dijke P. Balancing the activation state of the endothelium via two distinct TGF-beta type I receptors. *EMBO J* 2002;**21**:1743–1753.
- Goumans MJ, Liu Z, ten Dijke P. TGF-beta signaling in vascular biology and dysfunction. Cell Res 2009;19:116–127.
- Blanco FJ, Santibanez JF, Guerrero-Esteo M, Langa C, Vary CP, Bernabeu C. Interaction and functional interplay between endoglin and ALK-1, two components of the endothelial transforming growth factor-beta receptor complex. J Cell Physiol 2005;204:574–584.
- Koleva RI, Conley BA, Romero D, Riley KS, Marto JA, Lux A et al. Endoglin structure and function: Determinants of endoglin phosphorylation by transforming growth factor-beta receptors. J Biol Chem 2006;281:25110–25123.
- Lastres P, Letamendia A, Zhang H, Rius C, Almendro N, Raab U et al. Endoglin modulates cellular responses to TGF-beta 1. J Cell Biol 1996;133:1109–1121.
- 45. Ma X, Labinaz M, Goldstein J, Miller H, Keon WJ, Letarte M et al. Endoglin is overexpressed after arterial injury and is required for transforming growth factor-beta-induced inhibition of smooth muscle cell migration. Arterioscler Thromb Vasc Biol 2000;20:2546–2552.
- Guerrero-Esteo M, Lastres P, Letamendia A, Perez-Alvarez MJ, Langa C, Lopez LA et al. Endoglin overexpression modulates cellular morphology, migration, and adhesion of mouse fibroblasts. *Eur J Cell Biol* 1999;**78**:614–623.
- Diez-Marques L, Ortega-Velazquez R, Langa C, Rodriguez-Barbero A, Lopez-Novoa JM, Lamas S et al. Expression of endoglin in human mesangial cells: modulation of extracellular matrix synthesis. *Biochim Biophys Acta* 2002;**1587**:36–44.
- Botella LM, Sanz-Rodriguez F, Sanchez-Elsner T, Langa C, Ramirez JR, Vary C et al. Lumican is down-regulated in cells expressing endoglin. Evidence for an inverse correlationship between Endoglin and Lumican expression. *Matrix Biol* 2004;22: 561–572.
- Li C, Issa R, Kumar P, Hampson IN, Lopez-Novoa JM, Bernabeu C et al. CD105 prevents apoptosis in hypoxic endothelial cells. J Cell Sci 2003;116:2677–2685.
- Jerkic M, Rodríguez-Barbero A, Prieto M, Toporsian M, Pericacho M, Rivas-Elena JV et al. Reduced angiogenic responses in adult Endoglin heterozygous mice. *Cardiovasc* Res 2006;69:845–854.
- Jerkic M, Rivas-Elena JV, Prieto M, Carron R, Sanz-Rodriguez F, Perez-Barriocanal F et al. Endoglin regulates nitric oxide-dependent vasodilatation. FASEB J 2004;18: 609–611.

- Toporsian M, Gros R, Kabir MG, Vera S, Govindaraju K, Eidelman DH *et al.* A role for endoglin in coupling eNOS activity and regulating vascular tone revealed in hereditary hemorrhagic telangiectasia. *Circ Res* 2005;**96**:684–692.
- Santibanez JF, Letamendia A, Perez-Barriocanal F, Silvestri C, Saura M, Vary CP et al. Endoglin increases eNOS expression by modulating Smad2 protein levels and Smad2-dependent TGF-beta signaling. J Cell Physiol 2007;210:456–468.
- Jerkic M, Rivas-Elena JV, Santibanez JF, Prieto M, Rodríguez-Barbero A, Perez-Barriocanal F et al. Endoglin regulates cyclooxygenase-2 expression and activity. Circ Res 2006;99:248–256.
- Lebrin F, Mummery CL. Endoglin-mediated vascular remodeling: mechanisms underlying hereditary hemorrhagic telangiectasia. Trends Cardiovasc Med 2008;18:25–32.
- van den Driesche S, Mummery CL, Westermann CJ. Hereditary hemorrhagic telangiectasia: an update on transforming growth factor beta signaling in vasculogenesis and angiogenesis. *Cardiovasc Res* 2003;**58**:20–31.
- van Laake LW, van den Driesche S, Post S, Feijen A, Jansen MA, Driessens MH et al. Endoglin has a crucial role in blood cell-mediated vascular repair. *Circulation* 2006; 114:2288–2297.
- Tanaka F, Otake Y, Yanagihara K, Kawano Y, Miyahara R, Li M et al. Correlation between apoptotic index and angiogenesis in non-small cell lung cancer: comparison between CD105 and CD34 as a marker of angiogenesis. *Lung Cancer* 2003;**39**: 289–296.
- Liu Y, Jovanovic B, Pins M, Lee C, Bergan RC. Over expression of endoglin in human prostate cancer suppresses cell detachment, migration and invasion. *Oncogene* 2002; 21:8272–8281.
- Jovanovic BD, Huang S, Liu Y, Naguib KN, Bergan RC. A simple analysis of gene expression and variability in gene arrays based on repeated observations. *Am J Pharmacogenomics* 2001;**1**:145–152.
- Wong VC, Chan PL, Bernabeu C, Law S, Wang LD, Li JL et al. Identification of an invasion and tumor-suppressing gene, Endoglin (ENG) silenced by both epigenetic inactivation and allelic loss in esophageal squamous cell carcinoma. Int J Cancer 2008;**123**:2816–2823.
- Oxmann D, Held-Feindt J, Stark AM, Hattermann K, Yoneda T, Mentlein R. Endoglin expression in metastatic breast cancer cells enhances their invasive phenotype. *Oncogene* 2008;27:3567–3575.
- Brewer CA, Setterdahl JJ, Li MJ, Johnston JM, Mann JL, McAsey ME. Endoglin expression as a measure of microvessel density in cervical cancer. *Obstet Gynecol* 2000;96:224–228.
- Saad RS, Jasnosz KM, Tung MY, Silverman JF. Endoglin (CD105) expression in endometrial carcinoma. Int J Gynecol Pathol 2003;22:248–253.
- Schimming R, Marme D. Endoglin (CD105) expression in squamous cell carcinoma of the oral cavity. *Head Neck* 2002;24:151–156.
- Akagi K, Ikeda Y, Sumiyoshi Y, Kimura Y, Kinoshita J, Miyazaki M et al. Estimation of angiogenesis with anti-CD105 immunostaining in the process of colorectal cancer development. Surgery 2002;31:S109–S113.
- Düwel A, Eleno N, Jerkic M, Arevalo M, Bolaños JP, Bernabeu C et al. Reduced tumor growth and angiogenesis in endoglin-haploinsufficient mice. *Tumour Biol* 2007;28:1–8.
- Bockhorn M, Tsuzuki Y, Xu L, Frilling A, Broelsch CE, Fukumura D. Differential vascular and transcriptional responses to anti-vascular endothelial growth factor antibody in orthotopic human pancreatic cancer xenografts. *Clin Cancer Res* 2003;9: 4221–4226.
- Davis DW, Inoue K, Dinney CP, Hicklin DJ, Abbruzzese JL, McConkey DJ. Regional effects of an antivascular endothelial growth factor receptor monoclonal antibody on receptor phosphorylation and apoptosis in human 253J B-V bladder cancer xenografts. *Cancer Res* 2004;**64**:4601–4610.
- Henriksen R, Gobl A, Wilander E, Oberg K, Miyazono K, Funa K. Expression and prognostic significance of TGF-beta isotypes, latent TGF-beta 1 binding protein, TGF-beta type I and type II receptors, and endoglin in normal ovary and ovarian neoplasms. *Lab Invest* 1995;**73**:213–220.
- Hobson B, Denekamp J. Endothelial proliferation in tumours and normal tissues: continuous labelling studies. Br J Cancer 1984;49:405–413.
- Bredow S, Lewin M, Hofmann B, Marecos E, Weissleder R. Imaging of tumour neovasculature by targeting the TGF-beta binding receptor endoglin. *Eur J Cancer* 2000; 36:675–681.
- Costello B, Li C, Duff S, Butterworth D, Khan A, Perkins M et al. Perfusion of 99Tcm-labeled CD105 Mab into kidneys from patients with renal carcinoma suggests that CD105 is a promising vascular target. Int J Cancer 2004;109:436–441.
- Korpanty G, Grayburn PA, Shohet RV, Brekken RA. Targeting vascular endothelium with avidin microbubbles. *Ultrasound Med Biol* 2005;31:1279–1283.
- Korpanty G, Carbon JG, Grayburn PA, Fleming JB, Brekken RA. Monitoring response to anticancer therapy by targeting microbubbles to tumor vasculature. *Clin Cancer Res* 2007;**13**:323–330.
- 76. Neri D, Bicknell R. Tumour vascular targeting. Nat Rev Cancer 2005;5:436-446.
- 77. Molema G, Griffioen AW. Rocking the foundations of solid tumor growth by attacking the tumor's blood supply. *Immunol Today* 1998;**19**:392–394.
- Burrows FJ, Thorpe PE. Eradication of large solid tumors in mice with an immunotoxin directed against tumor vasculature. *Proc Natl Acad Sci USA* 1993;**90**: 8996–9000.

- Ferrara N. VEGF as a therapeutic target in cancer. Oncology 2005;69(Suppl. 3): 11-16.
- Ferrara N, Kerbel RS. Angiogenesis as a therapeutic target. Nature 2005;438: 967–974.
- Cohen MH, Gootenberg J, Keegan P, Pazdur R. FDA drug approval summary: bevacizumab plus FOLFOX4 as second-line treatment of colorectal cancer. *Oncologist* 2007;**12**:356–361.
- She X, Matsuno F, Harada N, Tsai H, Seon BK. Synergy between anti-endoglin (CD105) monoclonal antibodies and TGF-beta in suppression of growth of human endothelial cells. *Int J Cancer* 2004;**108**:251–257.
- Maier JA, Delia D, Thorpe PE, Gasparini G. In vitro inhibition of endothelial cell growth by the antiangiogenic drug AGM-1470 (TNP-470) and the anti-endoglin antibody TEC-11. Anticancer Drugs 1997;8:238-244.
- Korn T, Muller R, Kontermann RE. Bispecific single-chain diabody-mediated killing of endoglin-positive endothelial cells by cytotoxic T lymphocytes. *J Immunother* 2004; 27:99–106.
- Volkel T, Holig P, Merdan T, Muller R, Kontermann RE. Targeting of immunoliposomes to endothelial cells using a single-chain Fv fragment directed against human endoglin (CD105). *Biochim Biophys Acta* 2004;**1663**:158–166.
- Ahmadvand D, Rasaee MJ, Rahbarizadeh F, Mohammadi M. Production and characterization of a high-affinity nanobody against human endoglin. *Hybridoma (Larchmt)* 2008;27:353–360.
- Seon BK, Matsuno F, Haruta Y, Kondo M, Barcos M. Long-lasting complete inhibition of human solid tumors in SCID mice by targeting endothelial cells of tumor vasculature with antihuman endoglin immunotoxin. *Clin Cancer Res* 1997;3:1031–1044.
- Matsuno F, Haruta Y, Kondo M, Tsai H, Barcos M, Seon BK. Induction of lasting complete regression of preformed distinct solid tumors by targeting the tumor vasculature using two new anti-endoglin monoclonal antibodies. *Clin Cancer Res* 1999;5: 371–382.
- Tabata M, Kondo M, Haruta Y, Seon BK. Antiangiogenic radioimmunotherapy of human solid tumors in SCID mice using (125)I-labeled anti-endoglin monoclonal antibodies. *Int J Cancer* 1999;82:737–742.
- Tsujie M, Tsujie T, Toi H, Uneda S, Shiozaki K, Tsai H et al. Anti-tumor activity of an anti-endoglin monoclonal antibody is enhanced in immunocompetent mice. Int J Cancer 2008;122:2266–2273.
- Tsujie M, Uneda S, Tsai H, Seon BK. Effective anti-angiogenic therapy of established tumors in mice by naked anti-human endoglin (CD105) antibody: differences in growth rate and therapeutic response between tumors growing at different sites. *Int J Oncol* 2006;29:1087–1094.
- Takahashi N, Haba A, Matsuno F, Seon BK. Antiangiogenic therapy of established tumors in human skin/severe combined immunodeficiency mouse chimeras by antiendoglin (CD105) monoclonal antibodies, and synergy between anti-endoglin antibody and cyclophosphamide. *Cancer Res* 2001;61:7846–7854.
- Uneda S, Toi H, Tsujie T, Tsujie M, Harada N, Tsai H et al. Anti-endoglin monoclonal antibodies are effective for suppressing metastasis and the primary tumors by targeting tumor vasculature. Int J Cancer 2009;125:1446–1453.
- Tan GH, Huang FY, Wang H, Huang YH, Lin YY, Li YN. Immunotherapy of hepatoma with a monoclonal antibody against murine endoglin. World J Gastroenterol 2007;13:2479–2483.
- Shiozaki K, Harada N, Greco WR, Haba A, Uneda S, Tsai H et al. Antiangiogenic chimeric anti-endoglin (CD105) antibody: pharmacokinetics and immunogenicity in nonhuman primates and effects of doxorubicin. *Cancer Immunol Immunother* 2006;55:140–150.
- Muñoz R, Arias Y, Ferreras JM, Rojo MA, Gayoso MJ, Nocito M et al. Targeting a marker of the tumour neovasculature using a novel anti-human CD105-immunotoxin containing the non-toxic type 2 ribosome-inactivating protein nigrin b. Cancer Lett 2007;256:73-80.
- Rosen L, Gordon MS, Hurwitz HI, Wong MK, Adams BJ, Alvarez D et al. Early evidence of tolerability and clinical activity from a phase I study of TRC105 (anti-CD105 antibody) in patients with advanced refractory cancer (Abstract 3518). J Clin Oncol 2009;27 (Suppl.):15s.
- Duff SE, Li C, Garland JM, Kumar S. CD105 is important for angiogenesis: evidence and potential applications. FASEB J 2003;17:984–992.
- Balza E, Castellani P, Zijlstra A, Neri D, Zardi L, Siri A. Lack of specificity of endoglin expression for tumor blood vessels. *Int J Cancer* 2001;94:579–585.
- Seon BK. Expression of endoglin (CD105) in tumor blood vessels. Int J Cancer 2002; 99:310–311.
- Thorpe PE, Burrows FJ. Antibody-directed targeting of the vasculature of solid tumors. Breast Cancer Res Treat 1995;36:237–251.
- Griffioen AW, Damen CA, Blijham GH, Groenewegen G. Endoglin/CD105 may not be an optimal tumor endothelial treatment target. *Breast Cancer Res Treat* 1996;39: 239–242.
- Maio M, Altomonte M, Fonsatti E. Is it the primetime for endoglin (CD105) in the clinical setting? *Cardiovasc Res* 2006;**69**:781–783.
- 104. Calabrò L, Di Giacomo AM, Altomonte M, Fonsatti E, Mazzei MA, Volterrani L et al. Primary hepatic epithelioid hemangioendothelioma progressively responsive to interferon-alpha: is there room for novel anti-angiogenetic treatments? J Exp Clin Cancer Res 2007;26:145–150.