# Nanoparticles of lignin and saccharides from fishery wastes as sustainable UV-shielding, antioxidant and antimicrobial bio-fillers

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### Supporting information S1 Production of Chit 33

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**Supporting information S4** Morphological analysis of **CH/KLNPs** (Figure S4.1) and dimensional analysis of **CH/KLNPs** (Table S4.2).

Supporting information S5 UV-absorbing spectra of CH/KLNPs

Supporting information S6 Antimicrobial activity for CH/OLNPs and CH/KLNPs

**Supporting information S7** UV-visible absorption spectra of **COS3/OLNPs** and OLNPs (Panel A), and translation of the UV-visible absorption spectra into Tauc's plots to calculate the HOMO-LUMO gap of **COS3/OLNPs** and OLNPs

Supporting information S1 Production of Chit 33

The gene chit33 from Trichoderma harzianum CECT2413 fused to the Saccharomyces cerevisiae MF1 secretion signal was previously cloned in plasmid pIB4 and expressed in Pichia pastoris as previously reported<sup>1</sup>. The *P. pastoris* fermentation was obtained in a 5 L bioreactor (Biostart BPluss Sartorius Ltd., Gottingen, Germany) containing 3.5 L of medium (40 g/L glycerol, 26.7 mL H<sub>3</sub>PO<sub>4</sub> 85%, 0.93 g/L CaSO<sub>4</sub>, 18.2 g/L K<sub>2</sub>SO<sub>4</sub>, 14.9 g/L MgSO<sub>4</sub>, 4.13 g/L KOH, 2 mL biotin (0.2 g/L), and 4.35 mL of PTM1 trace salts). The fermentation parameters were maintained at 30 °C, 600 rpm agitation, 20% dissolved oxygen and pH was controlled at 5.0 units with NH<sub>4</sub>OH 28% (v/v) during 24 h (~40  $OD_{600}$  units). Then 100% methanol was added continuously during 4 days at 20  $\mu$ L/ min L of fermentation volume to induce the expression of protein Chit33 (final  $\sim$ 300 OD<sub>600</sub> units). Culture growth were monitored spectrophotometrically at 600 nm ( $OD_{600}$ ) and protein concentration using NanoDrop at 280 nm. The cells were removed by centrifuging at  $6000 \times g$  for 15 min, then the extracellular fraction was concentrated using 10000 MWCO PES membranes in a Vivaflow 50 system (Sartorius, Gottingen, Germany).

Supporting information S2 DSC curves of KLNPs, and OLNPs (Figure S2.1), CH/KLNPs and CH/OLNPs (Figure S2.2) and COS/OLNPs (Figure S2.3), respectively.



Figure S2.1 Panel A: DSC curves of KLNPs (red line) *versus* KL (black line). Panel B: DSC curves of OLNPs (red line) *versus* OL (black line).



Figure S2.2 Panel A: DSC curves of CH/KLNPs *versus* KLNPs assigned with the following color code: KLNPs (yellow line), CH1/KLNPs (brown line), CH2/KLNPs (azure line), CH3/KLNPs (red line), CH4/KLNPs (black line), CH5/KLNPs (purple line), CH6/KLNPs (green line), CH7/KLNPs (blue line). Panel B: DSC curves of CH/OLNPs *versus* OLNPs assigned with the following color code: OLNPs (azure line), CH1/OLNPs (brown line), CH2/KLNPs (yellow line), CH3/KLNPs (red line), CH2/KLNPs (green line), CH3/KLNPs (brown line), CH2/KLNPs (yellow line), CH3/KLNPs (red line), CH4/KLNPs (black line), CH5/KLNPs (black line), CH3/KLNPs (brown line), CH2/KLNPs (yellow line), CH3/KLNPs (black line), CH5/KLNPs (black line), CH5/KLNPs (black line), CH3/KLNPs (black line), CH5/KLNPs (black line)





Figure S2.3 Panel A: DSC curves of COS1/OLNPs (black line) and COS2/OLNPs (red line). Panel B: DSC curves of COS3/OLNPs (black line), COS4/OLNPs (red line) and COS5/OLNPs (blue line). Panel C: DSC curves of COS6/OLNPs (black line), COS7/OLNPs (red line) and COS8/OLNPs (blue line). Panel D: DSC curves of COS9/OLNPs (black line) and COS10/OLNPs (red line).

Supporting information S3 FT-IR of original CH 1-7 (Panel A) and OL (Panel B)





**Figure S3.** FT-IR spectra of original CH 1-7 and OL. Panel A: amide I band (1649 cm-1) and amide II band (1585 cm-1) of CH. Panel B: 1614 cm<sup>-1</sup> (C=C Aromatic bending vibration), 1462 cm<sup>-1</sup> (C–H deformations in  $-CH_2$ – and  $-CH_3$  vibration band), 1332 cm<sup>-1</sup> (C=O stretching of syringyl unit), and 1116 cm<sup>-1</sup> (aromatic C–H in plane deformation of syringyl unit), respectively.

**Supporting information S4** Morphological analysis of **CH/KLNPs** (Figure S4.1) and dimensional analysis of **CH/KLNPs** (Table S4.2).



#### Figure S4.1. SEM images of CH/KLNPs. Panel A: CH/KLNPs bearing CH with high DD (CH5/KLNPs and CH1/KLNPs)

and low DD (CH4/KLNPs), respectively.

Table S4.2 DLS and ζ-potential of CH/OLNPs and COS/OLNPs

Entry	Sample	Size(nm)	PDI <sup>a</sup>	ζ-Potential (mV)
1	KLNPs	210	0,35	-42
2	CH1/KLNPs	586	0,40	+69
3	CH4/KLNPs	551	0,54	+50
4	CH5/KLNPs	160	0,23	+30

<sup>a</sup>Polydispersion index.

## Supporting information S5 UV-absorbing spectra of CH/KLNPs



Figure S5. UV-absorbing capacity of CH/KLNPs

## Supporting information S6 Antimicrobial activity of CH/OLNPs and CH/KLNPs



Figure S6. The doubling time for the referred Gram-negative and Gram-positive bacteria of CH/OLNPs and CH/KLNPs

**Supporting information S7** UV-visible absorption spectra of **COS3/OLNPs**, and OLNPs (Panel A) and translation of the the UV-visible absorption spectra into Tauc's plots to calculate the HOMO-

LUMO gap of COS3/OLNPs, and OLNPs



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