Impact of transmitted HIV-1 drug resistance on the efficacy of first-line antiretroviral therapy with two nucleos(t)ide reverse transcriptase inhibitors plus an integrase inhibitor or a protease inhibitor

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Running head: transmitted drug resistance and efficacy of first-line antiretroviral therapy

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Key words: transmitted drug resistance, HIV-1, first-line antiretroviral therapy, integrase inhibitors, protease inhibitors

Abstract

Objectives
The aim of this study was to examine the impact of transmitted drug resistance (TDR) on response to first-line regimens with integrase strand transfer inhibitors (INSTI) or boosted protease inhibitors (bPI).

Methods
From an Italian observational database (ARCA) we selected HIV-1 infected drug-naïve patients starting 2 nucleoside reverse transcriptase inhibitors (NRTI) and either an INSTI or a bPI, with available pre-ART resistance genotype. The endpoint was virological failure (VF: plasma HIV-1 RNA >200 copies/ml after week 24.). WHO surveillance drug resistance mutations and the Stanford algorithm were used to classify patients into three resistance categories: no TDR (A), TDR but fully-active ART prescribed (B), TDR and at least low-level resistance to one or more prescribed drug (C).

Results
We included 1,365 patients with a median follow-up of 96-weeks (IQR 54-110): 1,205 (88.3%) starting bPI and 160 (11.7%) INSTI. Prevalence of TDR was 6.1%, 12.5%, 0.5% and 0% for NRTI, NNRTI, bPI, and INSTI, respectively. Cumulative Kaplan-Meier estimates for VF at 48-weeks were 11% (10.1-11.9) for the bPI- and 7.7% (5.4-10) for the INSTI-group. In the INSTI-group, cumulative estimates for VF at 48-weeks were 6% (4-8) in resistance category A, 5% (1-10) in B and 50% (30-70) in C (p<0.001). Resistance category C (versus A, adjusted hazard ratio, aHR 12.6, 3.2-49.8, p<0.001) and nadir CD4 (+100 cells/µL, aHR 0.6, 0.4-0.9, p=0.03) predicted VF. In the bPI-group, VF rates were not influenced by baseline resistance.

Conclusions
Our data support the need of NRTI-resistance genotyping in patients starting an INSTI-based first-line ART.
Introduction

Transmission of drug resistant HIV-1 is a well-known phenomenon detected in around 8% of newly diagnosed individuals in Europe, with significant differences depending on viral subtype, geographic area, risk group and migration timeline.\textsuperscript{1-3} Transmitted drug resistance (TDR) is increasing in Southern and Eastern Africa, particularly to the antiretroviral class of non-nucleoside reverse transcriptase inhibitors (NNRTI), a cornerstone of recommended first-line antiretroviral therapy (ART) in these countries.\textsuperscript{4} TDR may significantly influence the outcome of ART,\textsuperscript{5-7} therefore drug resistance testing is recommended for the choice of the first-line regimen in resource-rich countries.\textsuperscript{8-11}

The risk of virological failure was increased in patients harboring pre-treatment drug resistance to at least one of the prescribed drugs in NNRTI-based regimens, as compared with individuals without pre-treatment drug resistance, but not in patients with pre-treatment drug resistance and fully active ART.\textsuperscript{12}

International panels currently recommend first-line ART regimens including integrase strand transfer inhibitors (INSTI) or boosted protease inhibitors (bPI), because of their efficacy and tolerability.\textsuperscript{9-11} However, in the absence of resistance testing, some authors suggest to use bPI due to their higher genetic barrier compared with INSTI.\textsuperscript{8} Indeed, the influence of TDR on the efficacy of INSTI-based first-line regimens has not yet been established, due to the exclusion of individuals carrying TDR from clinical trials and the sparse data from observational cohorts.\textsuperscript{5,6,13-16}

The aim of this study was to examine the impact of TDR on response to first-line regimens in naïve patients starting INSTI-based 3-drug antiretroviral therapy. As a reference, we also analyzed the impact of TDR on the efficacy of boosted PI-based regimens.

Methods

Protease, reverse transcriptase and integrase genotype sequences from treatment-naïve HIV-1 infected adults starting a first-line therapy including 2 nucleoside or nucleotide
reverse transcriptase inhibitors (NRTI) plus 1 INSTI or 2 NRTI plus 1 bPI from January 2008 to June 2016 were selected from the Antiviral Response Cohort Analysis (ARCA), an Italian multicenter virological and clinical database [http://www.dbarca.net], including cases with at least 1 plasma HIV-1 RNA value after 24 weeks of follow up. The database was approved by the local Ethics Committees and written informed consent was obtained from all patients before participation. The study was performed in accordance with the ethical guidelines of the Declaration of Helsinki (7th revision).

Plasma genotypic resistance was determined by Sanger’s population sequencing using commercially available or homebrew systems. TDR was defined as the detection of at least one mutation among those included in the WHO-recommended surveillance drug resistance mutation (SDRM) list for NRTI, NNRTI, bPI\[^{17}\] and those included in the Stanford HIVdb SDRM Worksheet for INSTI [https://hivdb.stanford.edu/pages/SDRM.worksheet.INI.html]. The Stanford HIVdb algorithm (version 8.4, https://hivdb.stanford.edu) was used to classify patients into three resistance categories:\[^{4}\] absence of TDR mutations (resistance category A), presence of TDR mutations but use of a fully-active ART regimen (B), or presence of TDR mutations and at least low-level resistance to at least one prescribed drug (C). HIV-1 subtyping was available as automatically performed by BLAST upon sequence upload and further analyzed by phylogenetic analysis in case of <95% homology to the pure subtype reference panel.

The primary outcome was virological failure, defined as a plasma HIV-1 RNA >200 copies/mL after week 24, ignoring treatment changes. Survival analysis, using Kaplan-Meier curves, was employed to estimate the probability of virological failure. Predictors of virological failure were investigated using Cox regression models. All analyses were performed using SPSS (version 22, IBM, Armonk, NY).

**Results**

A total of 1,365 patients were included, 1,205 (88.3%) treated with 2 NRTI plus 1 bPI and 160 (11.7%) treated with 2 NRTI plus 1 INSTI. Baseline patients’ characteristics are shown in table 1. The main differences between the two treatment groups were a higher baseline plasma HIV-1 RNA and lower baseline and nadir CD4 cells counts in
the bPI group. Patients in the INSTI group were cared more frequently in Southern Italy and started therapy more recently. The most frequently prescribed INSTI was raltegravir (RAL) (39%), followed by dolutegravir (DTG) (35%) and elvitegravir/cobicistat (EVG/c) (26%). The most frequently used bPI was lopinavir/ritonavir (LPV/r) (41%), followed by atazanavir/ritonavir (ATV/r) (30%) and darunavir/ritonavir (DRV/r) (28%).

The overall prevalence of any TDR mutation was 18.4%, without differences between groups. NRTI, NNRTI, PI and INSTI resistance mutations were detected in 83 (6.1%), 171 (12.5%), 35 (2.6%) and 0 (0.0%) patients, respectively. While there was a similar prevalence of NRTI TDR in the two treatment groups, NNRTI TDR was more frequent in the bPI group (13.1% versus 8.1% in the INSTI group, p=0.043), whereas PI TDR was less frequent in the PI group (2.1% versus 6.3%, p=0.05).

During a median follow-up time of 96 weeks (IQR 54-110) virological failure occurred in 195 individuals in the PI-group and in 11 in the INSTI-group, with an estimated cumulative probability at 48 weeks of 11% (CI 95% 10.1-11.9) and 7.7% (CI 95% 5.4-10), respectively (p=0.01 by log-rank test).

In the INSTI group, resistance category C showed a significantly higher estimated probability of 48-week virological failure (50%, 95% CI 30-70) versus A (6%, 95% CI 4-8) and B (5%, 1-10) (p<0.001). By contrast, in the bPI group the estimated probability of virological failure at 48 weeks was similar in three categories: category A 11% (95% CI 10-12), B 12% (95% CI 10-14) and C 9% (95% CI 5-13) (p=0.390) (Fig.1). In the INSTI group, but not in the PI group, resistance category C (versus A, adjusted hazard ration, aHR 12.6, 3.2-49.8, p<0.001) and nadir CD4 (+100 cells/µL higher, aHR 0.6, 0.4-0.9, p=0.03) independently predicted virological failure. In the PI group, in a multivariable model adjusting for gender, nationality, TDF/FTC use, viral subtype, type of bPI and TDR to NRTI, independent predictors of virological failure were AZT/3TC use (aHR 2.3, CI 95% 1.4-3.9, p=0.002), calendar year (per 1 year more recent, aHR 0.9, CI 95% 0.8-0.9, p=0.04) and LPV/r use (versus DRV/r, aHR 1.4, CI 95% 1.0-2.0, p=0.03).
Eleven patients, mostly (9/11) harboring viral subtype B, experienced virological failure in the INSTI group: 8 were on treatment with RAL, 2 with DTG and 1 with EVG/c. At failure, plasma HIV-1 RNA ranged between 210 and 213,200 copies/mL and higher values were detected in patients with lower baseline CD4 counts. Three of the 11 failing INSTI carried TDR to NRTI: 2 M41L and 1 M184V, while none carried resistance to INSTI. Seven patients changed antiretroviral therapy after virological failure, with 6 patients switching to a bPI-based regimen. Among those that continued the previous regimen, 3 reached virological re-suppression at the subsequent visit and 1 was lost to follow up.

Discussion

The key finding of this study is the impact of pre-treatment HIV-1 drug resistance on the risk of virological failure in patients initiating ART with 2 NRTI plus INSTI. Despite the small number of cases, the magnitude of this effect was very relevant, with a more than 10-fold higher adjusted hazard of virological failure as compared to patients without TDR. To our knowledge, this is the first report showing a significant impact of TDR to NRTI on the activity of first-line regimens with 2 NRTI plus INSTI, the current standard of care of first-line ART. Indeed, previous observational studies on the influence of TDR did not include INSTI-based regimens and clinical trials with INSTI excluded patients with TDR. Interestingly, in the same group, TDR not affecting the activity of the prescribed drugs did not show any impact on virological efficacy. This finding is reassuring, suggesting that even in the presence of TDR, INSTI-based first-line regimens are effective when fully active accompanying drugs are selected based on the resistance test result.

The overall prevalence of TDR in this cohort was 18.4%, higher than usually reported in European cohorts, and was primarily driven by NNRTI resistance. This could be explained by the fact that, detection of TDR may have advised to use high-genetic barrier bPI therapy, resulting in an overestimate of TDR in the case file. In addition, bPI-based regimens were preferentially prescribed to more challenging patients, such as those with higher baseline viral load and lower CD4 counts, possibly explaining at least in part the higher virological efficacy of INSTI-based as compared with PI-based regimens observed here. The relatively long period of observation (2008-2016) may
also explain some imbalance observed between the two treatment groups reflecting drug availability over time. In the PI-group, AZT/3TC and LPV/r use were associated to more frequent virological failure, suggesting a crucial role of their lower tolerability and efficacy.

The main limitations of this study are the retrospective nature, the small number of patients treated with INSTIs and the relatively limited sample size in the INSTI treatment groups, which did not allow a sufficient power to detect differences among drugs with different genetic barrier. Future analyses including a larger and balanced INSTI group are necessary to confirm our findings and clarify whether NRTI TDR has a different impact on virological efficacy using different types of INSTI.

In conclusion, our findings support the need of pre-treatment drug resistance testing to NRTI in order to optimize antiretroviral therapy in patients starting first-line INSTI-based regimens.

Acknowledgments

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Funding

This study was conducted as part of our routine work.

Transparency declarations

CSR, LP, GP, VB, MP, GP and GZ none to declare.
BR declare consultant fees from Janssen, ViiV Healthcare, Abbvie, MSD and Gilead, all outside the submitted work.
MC received speakers’ honoraria and support for travel to meetings from BMS, Gilead, Merck Sharp &Dohme ( MSD), ViiV Healthcare and JC.
MMS has received funds for attending symposia, speaking, organizing educational activities and participating at Scientific Board from ViiV, Janssen Cilag and Gilead.

AA received fees for consultancy from Abbvie, Bristol Myers Squibb, Gilead Sciences, Janssen-Cilag, Merck, ViiV Healthcare, and research institutional grants from Bristol Myers Squibb, Gilead Sciences, Janssen-Cilag, ViiV Healthcare.

MZ grants from ViiV Healthcare e Gilead Sciences, consultancy fees from ViiV Healthcare, Gilead Sciences e Janssen-Cilag.

ADL Received unrestricted research grants from ViiV, Merck and Gilead (Fellowship Program) and was a paid consultant for ViiV, Merck and Gilead and Janssen-Cilag.

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Fig. 1 Kaplan–Meier curves showing the impact of the different pre-treatment HIV-1 drug resistance category on the virological outcome of first-line regimens based on 2 NRTI plus either a boosted PI (a) or an integrase inhibitor (b).

a. PI-group

b. INSTI-group
Table 1. Baseline characteristics of the patient population.

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Overall</th>
<th>bPI group</th>
<th>INSTI group</th>
<th>P-value*</th>
</tr>
</thead>
<tbody>
<tr>
<td>N= 1,365</td>
<td>N= 1,205</td>
<td>N= 160</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male, n (%)</td>
<td>1006/1355 (73.7)</td>
<td>880/1196 (73.0)</td>
<td>126/159 (78.8)</td>
<td>0.30</td>
</tr>
<tr>
<td>Age (year), median (IQR)</td>
<td>40 (33-48)</td>
<td>40 (33-48)</td>
<td>40 (30-48)</td>
<td>0.60</td>
</tr>
<tr>
<td>Italian born, n (%)</td>
<td>982/1365 (71.9)</td>
<td>882/1205 (73.2)</td>
<td>100/160 (62.5)</td>
<td>0.05</td>
</tr>
<tr>
<td>Risk factor, n (%):</td>
<td></td>
<td></td>
<td></td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Heterosexual contacts</td>
<td>444 (32.5)</td>
<td>405 (33.6)</td>
<td>39 (24.4)</td>
<td></td>
</tr>
<tr>
<td>MSM</td>
<td>272 (19.9)</td>
<td>245 (20.3)</td>
<td>27 (16.8)</td>
<td></td>
</tr>
<tr>
<td>Injection drug users</td>
<td>109 (8.0)</td>
<td>101 (8.4)</td>
<td>8 (5.0)</td>
<td></td>
</tr>
<tr>
<td>Other/Unknown</td>
<td>540 (39.6)</td>
<td>454 (37.7)</td>
<td>86 (53.8)</td>
<td></td>
</tr>
<tr>
<td>Geographical area, n (%):</td>
<td></td>
<td></td>
<td></td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Northern Italy</td>
<td>584 (42.8)</td>
<td>540 (44.8)</td>
<td>44 (27.5)</td>
<td></td>
</tr>
<tr>
<td>Central Italy</td>
<td>522 (38.2)</td>
<td>465 (38.6)</td>
<td>57 (35.6)</td>
<td></td>
</tr>
<tr>
<td>Southern Italy and Islands</td>
<td>259 (19.0)</td>
<td>200 (16.6)</td>
<td>59 (36.9)</td>
<td></td>
</tr>
<tr>
<td>Calendar year of treatment start, median (IQR)</td>
<td>2011 (2009-2013)</td>
<td>2011 (2009-2012)</td>
<td>2015 (2014-2016)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Time from HIV diagnosis (years), median (IQR)</td>
<td>0.3 (0.1-2.5)</td>
<td>0.3 (0.1-2.5)</td>
<td>0.5 (0.2-2.9)</td>
<td>0.60</td>
</tr>
<tr>
<td>Baseline plasma HIV-1 RNA (log_{10} copies/mL), median (IQR)</td>
<td>4.9 (4.3-5.4)</td>
<td>4.9 (4.4-5.4)</td>
<td>4.7 (4.5-2)</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td>Baseline CD4 cell count (cells/mm$^3$), median (IQR)</td>
<td>258.5 (103-383)</td>
<td>240 (96-364)</td>
<td>380 (198-557)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>CD4 nadir cell count (cells/mm$^3$), median (IQR)</td>
<td>230 (95-346)</td>
<td>222 (89-334)</td>
<td>323 (167-496)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Subtype, n (%):</td>
<td></td>
<td></td>
<td></td>
<td>0.30</td>
</tr>
<tr>
<td>B</td>
<td>944 (69.2)</td>
<td>839 (69.6)</td>
<td>105 (65.6)</td>
<td></td>
</tr>
<tr>
<td>non B</td>
<td>421 (30.8)</td>
<td>366 (30.4)</td>
<td>55 (34.4)</td>
<td></td>
</tr>
<tr>
<td>Backbone, n (%):</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>TDF/FTC</td>
<td>1011 (74.1)</td>
<td>895 (74.3)</td>
<td>116 (72.5)</td>
<td>0.63</td>
</tr>
<tr>
<td>ABC/3TC</td>
<td>240 (17.6)</td>
<td>198 (16.4)</td>
<td>42 (26.2)</td>
<td>0.002</td>
</tr>
<tr>
<td>AZT/3TC</td>
<td>102 (7.5)</td>
<td>102 (8.5)</td>
<td>0 (0)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>other</td>
<td>12 (0.9)</td>
<td>10 (0.8)</td>
<td>2 (1.3)</td>
<td>0.59</td>
</tr>
<tr>
<td>Anchor drug</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>DRV/r</td>
<td>339 (28.1)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Drug</td>
<td>Any class</td>
<td>NRTI</td>
<td>NNRTI</td>
<td>PI</td>
</tr>
<tr>
<td>---------------</td>
<td>-----------</td>
<td>----------</td>
<td>--------</td>
<td>------</td>
</tr>
<tr>
<td>LPV/r</td>
<td>499 (41.4)</td>
<td>222 (18.4)</td>
<td>158 (13.1)</td>
<td>25 (2.1)</td>
</tr>
<tr>
<td>ATV/r</td>
<td>367 (30.5)</td>
<td>74 (6.1)</td>
<td>9 (5.6)</td>
<td>0 (0.0)</td>
</tr>
<tr>
<td>RAL</td>
<td>63 (39.4)</td>
<td>9 (5.6)</td>
<td>13 (8.1)</td>
<td>0 (0.0)</td>
</tr>
<tr>
<td>EVG</td>
<td>41 (25.6)</td>
<td>74 (6.1)</td>
<td>9 (5.6)</td>
<td>0 (0.0)</td>
</tr>
<tr>
<td>DTG</td>
<td>56 (35.0)</td>
<td>9 (5.6)</td>
<td>13 (8.1)</td>
<td>0 (0.0)</td>
</tr>
</tbody>
</table>

Patients with transmitted drug resistance, n (%):

ABC/3TC, abacavir/lamivudine; ATV/r, atazanavir/ritonavir; AZT/3TC, zidovudine/lamivudine; DRV/r, darunavir/ritonavir; DTG, dolutegravir; EVG, elvitegravir; INSTI, Integrase strand transfer inhibitors; LPV/r, lopinavir/ritonavir; MSM, man who have sex with man; NRTI, nucleoside reverse transcriptase inhibitor; NNRTI, non-nucleoside reverse transcriptase inhibitor; bPI, boosted protease inhibitor; RAL, raltegravir; TDF/FTC, tenofovir/emtricitabine.